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Research Article

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Quantitation of Human Gamma Globin Genes and Gamma Globin mRNA with Purified Gamma Globin Complementary DNA

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ABSTRACT Complementary DNA (cDNA) specific for γ -globin nucleotide sequences has been prepared by hybridizing total cDNA made from cord blood messenger RNA (mRNA) as template to an excess of normal adult human globin mRNA and recovering the single-stranded cDNA from hydroxylapatite. The specificity of the γ cDNA for γ mRNA sequences is strongly supported by the hybridization of this cDNA at low C_0t values (C_0 , concentration of RNA and t , time in seconds) to RNA samples containing large amounts of functional γ globin mRNA and the lack of hybridization to RNA samples containing little, if any, γ -globin mRNA. The absence of cross-hybridization of γ cDNA with α , β , and δ mRNAs is demonstrated by the complete hybridization of the γ cDNA to mRNA samples completely lacking either α or β and δ mRNA. An estimate of the number of γ -globin genes in human cellular DNA was obtained by hybridization of purified γ cDNA to DNA from spleen and white blood cells of normal and β -thalassemia subjects and measurement of the percent of γ cDNA hybridized at saturation. The results indicate that there are between one and two γ -globin genes per total haploid gene DNA equivalent obtained from both normal and β -thalassemia subjects. These values are consistent with genetic evidence for the presence of multiple γ gene loci in human cells. The finding that the number of γ -globin genes in β -thalassemia DNA is similar to that in nonthalassemia DNA indicates that a deletion of γ -globin genes cannot account for either

the inadequate γ -globin synthesis or indirectly for the decreased or absent β -globin synthesis in β -thalassemia cells.

INTRODUCTION

The use of highly radioactive complementary DNA (cDNA)¹ has recently permitted the measurement of the number of globin genes in a variety of mammalian species (1-3). In addition, purified α and β cDNAs from human cells have been used to quantitate the relative number of α - and β -globin genes in human DNA (4,5). The results indicate that there are between one and four α -globin genes, and one and five β -globin genes per haploid human genome. More recently, the specificity of the hybridization of purified α and β cDNA probes to complementary nucleotide sequences in human cellular DNA has been strengthened by the detection of deletions of α - and β -like globin genes in certain forms of α - and β -thalassemia (4, 6-8) and in hereditary persistence of fetal hemoglobin (8-10). To quantitate the number of γ -globin genes in human cells, we have prepared specific γ -globin cDNA by hybridizing the cDNA synthesized with cord blood messenger RNA (mRNA) as substrate (α , β , and γ mRNA) to an excess of normal adult reticulocyte mRNA containing only α and β mRNA. After α and β cDNA were allowed to become hybridized, the cDNA remaining single stranded was isolated. This single-stranded material specifically hybridizes with mRNA from cells known to contain large amounts of γ mRNA, whether of adult or fetal origin, while there is a lack of hybridization of this γ cDNA when hybridized to mRNAs containing small amounts of γ mRNA.

¹Abbreviations used in this paper: C_0 , concentration of RNA; cDNA, complementary DNA; mRNA, messenger RNA; t , time in seconds.

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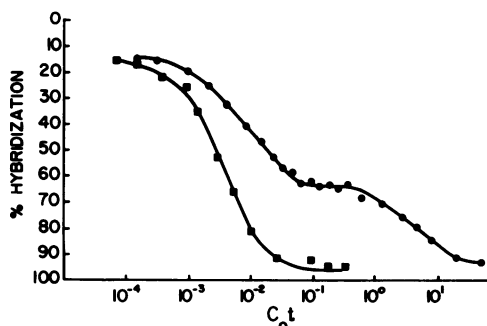


FIGURE 1 Hybridization of mRNA from cord blood and normal adult reticulocytes to cord blood cDNA: 32 pg–20 μ g of cord blood mRNA was annealed to 2,000 cpm (0.143 ng) of cord blood cDNA for 4 h at 68°C to obtain the C_0t values indicated. The percentage of hybridization of cDNA was assayed as described in Methods (■ — ■). 32 pg–0.2 μ g of normal adult reticulocyte mRNA was annealed to 2,000 cpm (0.143 ng) of cord blood cDNA as described above (● — ●).

With this highly specific γ cDNA, the relative amounts of γ -globin gene material in DNA from spleens and peripheral blood cells of patients with nonthalassemic conditions and with β -thalassemia have been compared. Calculation of the number of γ -globin genes indicate that there are between one and two γ -globin genes per total haploid gene DNA equivalent. There is no evidence for deletion of γ -globin gene material in β -thalassemia which could account for the inadequate compensation by γ -globin synthesis for the decrease or absence of β -globin in these cells. In addition, there is no evidence that γ gene deletion is indirectly responsible for the relative decrease in β -globin mRNA production in β^+ -thalassemia.

METHODS

Human globin mRNAs were obtained by extraction of either peripheral blood or liver of a patient with hydrops fetalis as described previously (4). The 6-16S RNA region was routinely isolated for use in the preparation of globin cDNA and for hybridization to purified and total cDNA. α and β cDNA were purified as previously reported (4). Globin synthesis in intact cells and translation of globin mRNA in a Krebs ascites tumor cell-free system was as previously described (11).

Preparation of γ -globin cDNA. The preparation of γ -globin cDNA was analogous to that described previously (12). In our experiments, 10 μ g of cord blood mRNA was used as template for the production of α -, β -, and γ -globin cDNAs. 4×10^6 cpm of this cord blood cDNA with a calculated specific activity of approximately 14,000 cpm/ng (3) was then hybridized to 5 μ g of globin mRNA (α and β) prepared from the reticulocytes of an adult patient with a hemolytic anemia. After 4 h of hybridization equivalent to a C_0t (C_0 , concentration of RNA times t , time in seconds) of 0.25, the hybridized and unhybridized cDNA and mRNA species were separated on hydroxylapatite. The single-stranded material was eluted with 0.12 M sodium phosphate

in 0.4% sodium dodecyl sulfate, while the double-stranded material was eluted with 0.4 M phosphate in 0.4% sodium dodecyl sulfate. The hydroxylapatite eluates were passed through a column of Sephadex G-50 (Pharmacia Laboratories, Inc., Piscataway, N.J.) and the excluded volume was collected. The material was then treated with alkali for 18 h at 37°C. The cDNA eluted initially as the double strand was used for subsequent purification of α - and β -globin cDNA (4), while the single-stranded material was used as γ -globin cDNA. The γ cDNA migrated as a single homogeneous peak of 7S DNA on alkaline sucrose gradients (13) and contained approximately 500 nucleotides when analyzed by polyacrylamide gel electrophoresis in formamide (14).

Preparation of human DNA. DNA was prepared from human spleen and liver of patients with and without β -thalassemia, as described previously (3, 4). DNA from peripheral blood was prepared similarly, except that the blood sample containing white blood cells was washed three times with 10 vol of isotonic saline, frozen, and thawed before washing with 5% sucrose containing 1 mM $MgCl_2$, 1 mM sodium phosphate (pH 6.5), 0.4% deoxycholate, and 0.8% Nonidet P40 (Shell International Ltd., London, England).

Hybridization of cDNA with human mRNA and cellular DNA. Globin cDNA was hybridized with mRNA under conditions described previously (15) and for 4 h unless otherwise specified. With this methodology, it has been shown previously (16) that the relative C_0t 1/2 values obtained (Table I) are proportional to the amount of mRNA present over a wide range of RNA concentrations. Globin mRNA was prepared by isolating 6–16S RNA from sucrose density gradients unless otherwise indicated (13). Hybridization of globin cDNA with spleen, liver, or white cell DNA was also previously described (4, 5).

RESULTS

Properties of cord blood cDNA. The hybridization kinetics of total cord blood cDNA to cord blood mRNA and normal adult reticulocyte globin mRNA is shown in Fig. 1. When cord blood cDNA and cord blood mRNA are annealed, a monophasic hybridization curve is obtained, indicating that the relative proportions of the different nucleotide sequences (α , β , and γ) are similar in both mRNA and cDNA. A similar monophasic curve is obtained when normal reticulocyte mRNA is hybridized with total cDNA prepared from normal adult reticulocyte mRNA (4). By contrast, when cord blood cDNA is annealed to normal reticulocyte mRNA containing predominantly α and β mRNA, the hybridization curve is diphasic (Fig. 1). The initial hybridization to 65% is due to the re-annealing of α and β complementary sequences. Only after an additional 50–100-fold excess of normal mRNA is added is the remaining cDNA hybridized. This is presumably due to the small amount of γ mRNA present in normal reticulocyte mRNA, which requires that a large excess of this mRNA be added to hybridize the γ cDNA of the cord blood cDNA. Presumed γ cDNA was isolated from the total cord blood cDNA by separating hybridized and unhybridized cDNA at a C_0t of 0.25 (Fig. 1) by hydroxylapatite chromatography.

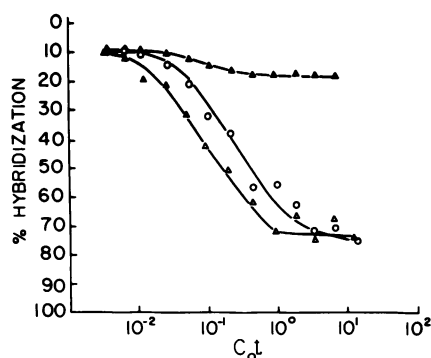


FIGURE 2 Hybridization of $\delta\beta$ -thalassemia mRNA with α , β , and γ cDNA. 0.1 ng–0.36 μ g of mRNA was hybridized to 2,000 cpm (0.143 ng) of α , β , or γ cDNA. Hybridization was for 24 h at 68°C to obtain the indicated C_0t values. The percentage of hybridization was assayed as described in Methods. (Δ — Δ), α cDNA; (\blacktriangle — \blacktriangle), β cDNA; (\circ — \circ), γ cDNA.

The unhybridized cDNA isolated from cord blood cDNA after hybridization with normal reticulocyte mRNA will subsequently be called γ cDNA. To demonstrate the specificity of the γ cDNA for γ mRNA sequences, the hybridization of α , β , and γ cDNA was measured in RNAs containing different amounts of functional γ mRNA.

Hybridization of $\delta\beta$ -thalassemia mRNA. Globin mRNA was isolated from the peripheral blood cells of a patient homozygous for $\delta\beta$ -thalassemia. The peripheral blood of this patient contains only HbF and when the reticulocytes of this patient were incubated with [3 H]leucine, only γ - and α -globin were synthesized, in a ratio of 0.62:1 (17). Cell-free translation of this mRNA led to only γ - and α -globin synthesis in a ratio of 0.33:1. With purified β cDNA, only 10% hybridization above background is obtained, even at C_0t values of 10^2 (Fig. 2). By contrast, over 80% of the γ cDNA is hybridized at a C_0t value approximately double that at which α cDNA is hybridized. These results indicate that the γ cDNA does not hybridize with β mRNA (since no β mRNA is present) and contains more than 80% of an mRNA species present in reticulocytes in amounts comparable to α mRNA. These observations also show that the γ cDNA is not a “nonglobin” cDNA species present in cord blood, since it hybridizes at C_0t values comparable to α cDNA with RNA from an adult patient known to have comparable amounts of α and γ mRNA. In addition, when γ cDNA is hybridized to either 6–16S RNA or total RNA isolated from human fibroblast cell lines, less than 10% of the γ cDNA is hybridized at C_0t values of 10^3 .

Hybridization to hydrops fetalis mRNA. The possibility that the γ cDNA cross-hybridizes to α mRNA sequences was studied by hybridization of the γ cDNA

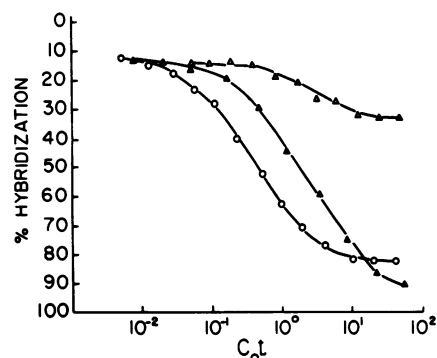


FIGURE 3 Hybridization of mRNA from hydrops fetalis liver with β and γ cDNA. 32 ng–32 μ g of mRNA was hybridized to 2,000 cpm (0.143 ng) of γ and β cDNA, respectively. Hybridization was for 4 h at 68°C to obtain the indicated C_0t values. The percentage of hybridization was assayed as described in Methods. (\blacktriangle — \blacktriangle), β cDNA; (\circ — \circ), γ cDNA; (Δ — Δ), α cDNA.

to hydrops fetalis mRNA, which contains no α mRNA (4). When γ and β cDNA are hybridized to mRNA prepared from the liver of a patient with hydrops fetalis (Fig. 3), there is six times more γ mRNA than β mRNA based on the differences in C_0t 1/2 of mRNA required to protect similar amounts of each cDNA probe. These relative amounts of γ and β mRNA are consistent with the 5–10-fold greater γ as compared to β -globin present and synthesized in hydrops fetalis cells (Table I, ref. 4). The α cDNA hybridizes to 20% above background even at high C_0t values (Fig. 3, ref. 4). The hybridization results with γ and β probes indicate that the γ cDNA hybridizes to γ mRNA at a rate independent of the amount of β mRNA present, and conversely, that β cDNA hybridizes to β mRNA at a rate independent of the amount of γ mRNA present.

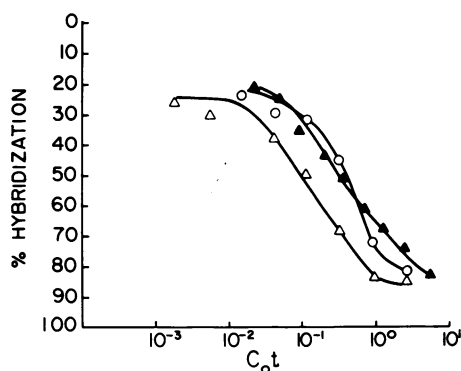


FIGURE 4 Hybridization of mRNA from reticulocytes of a homozygous β -thalassemia patient with α , β , and γ cDNA. 0.8 ng–0.28 μ g of mRNA was hybridized to 2,000 cpm (0.143 ng) of α , β , or γ cDNA. Hybridization was for 4 h at 68°C to obtain the indicated C_0t values. The percentage of hybridization was assayed as described in Methods. (Δ — Δ), α cDNA; (\blacktriangle — \blacktriangle), β cDNA; (\circ — \circ), γ cDNA.

TABLE I
Relative Globin mRNA Content and Globin
Synthesis in Human Erythroid Cells

Source	mRNA content*			Globin synthesis†		
	α	β	γ	α	β	γ
Normal—1 (Fig. 5b)	1.0	1.0	0.01	1.0	1.0	0.01
Normal—2	1.0	1.1	0.005	1.0	1.0	0.01
Normal—3	1.0	1.0	0.003	—	—	—
Normal—4	1.0	1.0	0.025	—	—	—
Homozygous $\delta\beta$ -thal (Fig. 2)	1.0	—	0.33	1.0	—	0.62
Hydrops fetalis (Fig. 3)	—	1.0	5.9	—	1.0	6.0
Cord blood (Fig. 5a)	1.0	1.0	1.8	1.0	0.4	0.6
Homozygous β^+ thal (Fig. 4)	1.0	0.072	0.08	1.0	0.27	0.27
Homozygous β^+ thal	1.0	0.25	0.25	1.0	0.23	0.21
HbH—1	1.0	3.3	0.01	1.0	1.8	0.01
HbH (bone marrow)—1	1.0	3.8	0.004	1.0	1.8	0.01

* The relative mRNA content is calculated from the C_{ot} $1/2$ values obtained by hybridization to α , β , and γ cDNAs. The relative mRNA content is the reciprocal of the C_{ot} $1/2$ s; for example, α/β mRNA content = $C_{ot} \ 1/2 \ \beta/C_{ot} \ 1/2 \ \alpha$. The results have been normalized compared to an α mRNA value of 1.0, except for hydrops fetalis mRNA, for which the value for β mRNA is set as 1.0.

† Globin synthesis in intact cells with [3 H]leucine as described in the text. α -globin synthesis has been set at 1.0, except for hydrops fetalis, for which β -globin synthesis is set 1.0.

Over 90% of the γ cDNA hybridizes at a low C_{ot} , indicating that over 90% of the γ cDNA is not α cDNA.

Hybridization to α and β thalassemia mRNA. When mRNA isolated from reticulocytes of two patients with homozygous β^+ thalassemia is hybridized with α , β , and γ cDNAs, the results indicate that there is a significant amount of γ mRNA present, 0.08 and 0.25 times as much γ mRNA as α mRNA (Fig. 4, Table I). In these experiments, it is also clear that the amount of γ cDNA present in the cells measured by comparing hybridization to α , β , and γ cDNA is comparable to the relative amounts of γ -, β -, and α -globin synthesized by these cells or by mRNA isolated from them and

translated in cell-free systems. In addition, with the mRNA of five patients with β^0 -thalassemia from Ferrara described elsewhere (8), the ratio of γ to α mRNA obtained by hybridization was 0.23 to 0.77, comparable to the relative amounts of γ - and α -globin present in intact cells. When the mRNA from a patient with hemoglobin H disease (HbH), a form of α -thalassemia, is hybridized to α , β , and γ cDNA, there is a threefold excess of β mRNA over α mRNA, and very little γ mRNA (Table I). Again, there is no evidence for hybridization of γ cDNA to the β mRNA present in this case. These results are consistent with the negligible γ -globin synthesis in HbH cells.

Hybridization to normal and cord blood mRNA. Globin mRNA from reticulocytes of fetuses at birth (cord blood) was hybridized to purified α , β , and γ cDNAs (Table I, Fig. 5a). When cord blood mRNA is hybridized to α , β , and γ cDNA, the C_{ot} $1/2$ with α , β , and γ mRNA are similar (Fig. 5a, Table I), indicating that approximately equal amounts of these RNAs are present in the cord blood mRNA. These results are consistent with the relatively equal synthesis of α -, β -, and γ -globin in intact cells of cord blood and by isolated mRNA in cell-free systems (Table I). When γ cDNA is annealed with normal adult reticulocyte mRNA, there is slow gradual rate of hybridization of the γ cDNA until a C_{ot} of 10^{-2} , and more rapid hybridization at C_{ot} values above 10^{-2} , until the probe is fully hybridized at C_{ot} values 50–100 times that required to hybridize α or β cDNA (Fig. 5b). The hybridization of γ cDNA with normal mRNA at low C_{ot} values (less than 0.01) is probably due to contamination of the γ -globin cDNA with α or β cDNA. This level of contamination of γ cDNA by α and β cDNA (about 20%) is consistent with that obtained with $\delta\beta$ -thalassemia and hydrops fetalis mRNA.

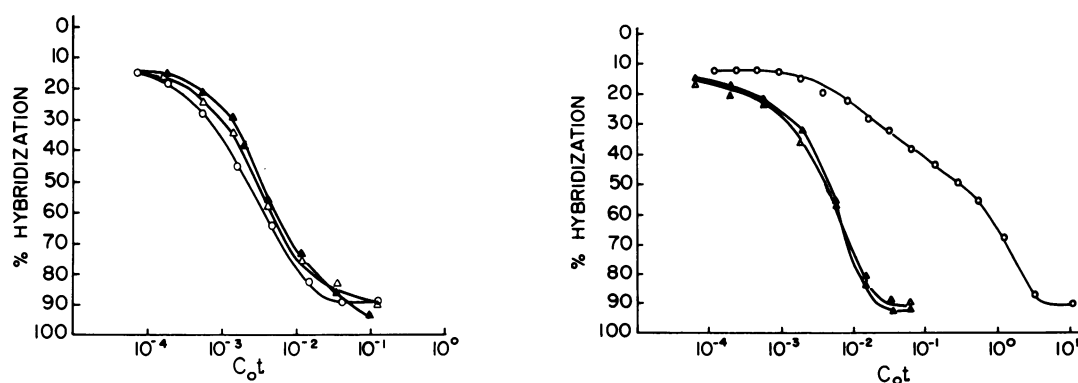


FIGURE 5 (a) Hybridization of mRNA from cord blood with α , β , and γ cDNA. 36 pg–48 ng of mRNA was hybridized to 2,000 cpm (0.143 ng) of α , β , or γ cDNA for 4 h at 68°C to obtain the indicated C_{ot} . The percentage of hybridization was assayed as described in Methods. (Δ — Δ), α cDNA; (\blacktriangle — \blacktriangle), β cDNA; (\circ — \circ), γ cDNA. (b) Hybridization of mRNA from normal adult reticulocytes with α , β and γ cDNA: 32 pg–4 μ g of normal adult reticulocyte mRNA was hybridized to 2,000 cpm (0.143 ng) of purified α , β , and γ cDNA for 4 h at 68°C to obtain the C_{ot} values indicated. (a), left; (b), right.

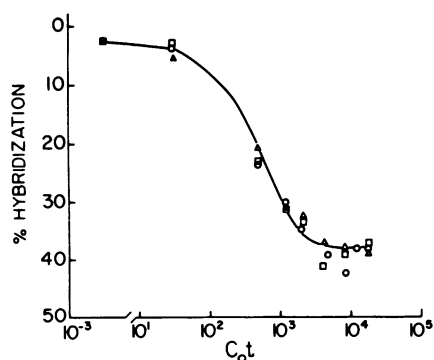


FIGURE 6 Hybridization of normal and thalassemia cellular spleen DNA to γ cDNA. 45- μ l aliquots of a 300- μ l incubation mixture containing 0.9 mg of DNA and 4,200 cpm (0.3 ng) of γ cDNA in 0.12 M sodium phosphate (pH 6.8), 0.3 M NaCl, 1 mM EDTA, and 0.4% sodium dodecyl sulfate were heated in a boiling water bath for 15 min and incubated at 68°C for various times to attain the C_0t values shown. The samples were assayed by hydroxylapatite as described previously (3). (○ — ○), normal spleen DNA; (△ — △), β^0 -thalassemia spleen DNA; (□ — □), β^+ -thalassemia spleen DNA.

Number of γ -globin genes in human DNA. We have previously reported that the relative numbers of α - and β -globin genes in normal DNA (4). In these hybridizations with relative cDNA excess (see Table II, legend), a competition between the cDNA and one of the cellular DNA strands for the complementary cellular DNA strand limits the extent of cDNA hybridization (Fig. 6, Table II) (3). By contrast, in vast cellular DNA excess, over 80% of the γ cDNA is hybridized. At inputs of cDNA per cellular DNA of 3.3×10^{-7} (relative cDNA excess), the hybridization of normal and thalassemia DNA to γ cDNA was measured (Fig. 6, Table II). At saturation hybridization with these inputs, approximately 40% of the γ cDNA was hybridized. When DNA from patients with β -thalassemia is hybridized to γ cDNA, results similar to those with DNA from normal cells are obtained (Fig. 6, Table II). The C_0t 1/2 of the unique sequences in unlabelled cellular DNA in these studies is approximately 1,100, while that of the γ cDNA was between 350 and 450 (Fig. 6). These data indicate no detectable deletion of γ -globin genes in β -thalassemia of either the β^+ or β^0 type in the patients studied.

DISCUSSION

We have previously separated α and β cDNA from total normal adult human cDNA by using mRNA from patients with hydrops fetalis that completely lacks α mRNA sequences. More recently, we have separated α and β mRNAs by acrylamide gel electrophoresis and have prepared α and β cDNA using these mRNAs.²

² Nudel, U., F. Ramirez, and A. Bank. In preparation.

The hybridization kinetics of the α and β cDNA prepared by hybridization to hydrops fetalis mRNA and by electrophoresis are comparable. In the present study, a similar procedure has been used to isolate γ cDNA from cord blood by hybridizing total cord cDNA to normal adult reticulocyte mRNA containing negligible amounts of γ mRNA. The single-stranded cDNA recovered has been used as purified γ cDNA. The specificity of the γ cDNA has been demonstrated by its complete hybridization at relatively low C_0t values to a variety of mRNAs obtained from both adult and fetal cells known to contain large amounts of functional γ mRNA. The complete hybridization of the γ cDNA to mRNA from a patient homozygous for $\delta\beta$ -thalassemia and completely lacking functional or structurally intact δ and β mRNA indicates less than 20% contamination of γ cDNA with δ or β mRNA. In addition, the relatively similar C_0t values at which γ and α cDNAs are hybridized with $\delta\beta$ mRNA, and its lack of hybridization to fibroblast RNA make it extremely unlikely that the γ cDNA is due to "non-globin cDNA" resulting from nonglobin mRNA in the cord blood mRNA used to prepare the γ cDNA. If the

TABLE II
Hybridization of γ -Globin cDNA with Human Cellular DNA

Source	γ^* Hybrid (gene no.)†
	%
Normal spleen—1 (Fig. 6)	39 (2)
Normal spleen—2	36 (1–2)
Peripheral blood—1	35 (1–2)
β^0 thalassemia spleen—1 (Fig. 6)	40 (2)
β^0 thalassemia spleen—1	38 (2)
β^0 thalassemia peripheral blood—2	31 (1–2)
β^+ thalassemia spleen—1 (Fig. 6)	39 (2)
β^+ thalassemia spleen—2	36 (1–2)
β^+ thalassemia spleen—3	35 (1–2)

* Each hybridization contained 3.3×10^{-7} as much cDNA as cellular DNA (see legend to Fig. 6 for typical inputs). The percent hybrid is the average of the values obtained at the plateau of hybridization at C_0t values of 8,600 or greater. At least two determinations at different C_0t values have been used in each experiment and in most cases, duplicate samples have been analyzed.

† The number of gene copies per haploid genome shown in parentheses was calculated as described previously (3) as follows: The fraction of a single globin gene in the human genome was calculated as 1.1×10^{-7} (3). Thus, the above experiments were performed in "relative cDNA excess" with three times as much cDNA as for a single globin gene ($3.3 \times 10^{-7}/1.1 \times 10^{-7} = 3$). The number of globin genes (N) was then calculated from the equation $1/(N+3) = \% \text{ hybridization}$. For example: for normal spleen—1: $N/(N+3) = 39\%$, $N = 2$.

γ cDNA was primarily nonglobin cDNA specifically present in cord blood cells, the nonglobin mRNA it represented would not be present in amounts comparable to that of α mRNA in the cells of an adult patient with $\delta\beta$ -thalassemia. Similarly, it should not be present in large amounts in the cells of adult patients with β -thalassemia (Fig. 4, Table I). With these considerations, the γ cDNA is approximately 80% purified γ cDNA. The high C_{ot} values required to fully hybridize the γ cDNA to normal reticulocyte mRNA and hemoglobin H mRNA is additional evidence that the γ cDNA is primarily γ cDNA.

The lack of significant cross-hybridization of γ cDNA with β mRNA sequences under the conditions used is quite remarkable, since the amino acid sequence of γ and β chains differ by only 38 of 146 amino acids (18). Thus, it appears that there are more mutations in the nucleotide sequence coding for these globin genes than are reflected by changes in the amino acid sequence of γ - and β -globin. The nucleotide changes are apparently numerous enough to prevent stable hybrids between γ and β nucleotide sequences. It is possible that γ cDNA sequences homologous to α and β mRNA exist but have been removed by our method of preparing γ cDNA. However, the specificity of the γ cDNA isolated suggests that such sequences do not exist in the cDNA we have obtained, which contains about 500 nucleotides. The results also indicate that the amount of γ mRNA present in globin mRNA from human cells is roughly proportional to the relative amount of γ -globin translated in both intact cells and by mRNA in cell-free systems (Table I).

With purified α - and β -globin cDNA, it has been possible to quantitate the relative numbers of α - and β -globin genes in normal and β -thalassemia cellular DNA. With these probes, it has been possible to demonstrate specific deletions of either α - or β -like globin gene material in α - and $\delta\beta$ -thalassemia (4, 6–8) and in hereditary persistence of fetal hemoglobin (8–10). Similar results have been obtained with measurements of C_{ot} kinetics in cellular DNA excess (6, 7, 9, 10) and by limited cDNA excess hybridization (4, 8). We prefer this latter method, which relies on the saturation plateaus obtained, since it requires less cellular DNA and is quite reproducible (3, 4, 8; Table II).

In this paper, we have used the γ cDNA probe to compare the amount of γ -globin DNA in thalassemia and nonthalassemia DNA. The number of γ -globin genes in cellular DNA from patients with β -thalassemia is similar to those in nonthalassemia DNA (Table II). Comparable numbers of α - and β -globin genes have been previously reported in β^+ -thalassemia DNA as in nonthalassemia DNA (3, 4). In the studies reported here, both β^+ - and β^0 -thalassemia DNA were used and shown to contain γ -globin genes in amounts compar-

able to normal DNA. These results indicate that there is no detectable deletion of structural γ -globin genes in the DNA of these patients. On the basis of its size, the γ cDNA represents about 500 nucleotides homologous to the γ mRNA and starting at the 3' end of the RNA. Since there are approximately 100 adenylate residues at the 3' end of globin mRNAs, the γ -globin cDNA contains about 400 nucleotides excluding the polyadenylate regions. Approximately 450 nucleotides are required to encode structural γ -globin; in addition, by analogy with α and β mRNA, there are an additional 100 untranslated nucleotides at the 3' end of γ cDNA. Thus, the γ cDNA probably represents about 70% of the structural γ -globin gene sequence (400/550). It is, therefore, possible that deletions in the γ -globin gene homologous to the 5' end of the γ -globin mRNA will be undetected by this probe. With this reservation in mind, the results suggest that deletion of γ -globin genes cannot account for the decreased amount of γ -globin present in the cells of β -thalassemia patients. In addition, they indicate that a deletion of γ -globin genes is apparently unrelated to the decreased production of β -globin mRNA by β -globin genes in β^+ -thalassemia.

It is difficult to compare the absolute number of α -, β -, and γ -globin genes with the methodology in this paper, since the precise gene numbers depend on the absolute purity and length of the cDNA probes used, the absolute specific activity of these probes (which can only be calculated), the hybridization of the cDNA probes to nonglobin gene material or untranscribed globin gene material in cellular DNA, and variations in the relative hybridization of cDNA to DNA at varying cDNA to DNA inputs (3). For these reasons, it is difficult to compare the relative hybridization of cellular DNA to different cDNAs as a measure of the relative or absolute numbers of globin genes. It is more reliable to simply compare the hybridization of different cellular DNAs with the same probe at similar cellular DNA to cDNA inputs as reported here. Calculations of specific gene numbers can be made, within the limitations of this type of analysis. Comparing our present data to that we have published previously (4), we find that at similar levels of input of cDNA per cellular DNA, there is consistently less γ cDNA hybridization obtained at saturation than with α and β cDNA. Hybridization of α cDNA to human DNA may reflect not only the number of α genes but other embryonic α -like genes as well. Similarly, hybridization of β cDNA may well include δ gene hybridization as well as that of other embryonic β -like genes. Our previous data (4) and that of others (5) are consistent with strong genetic evidence for one β , one δ , and two α loci per haploid human genome. The lack of cross-hybridization of γ cDNA with β or α nucleotide sequences demonstrated in this study makes

it unlikely that hybridization of the γ cDNA is due to contamination with α or β cDNA sequences or cross-hybridization to α or β genes. However, it is possible that the γ cDNA hybridizes to embryonic or other γ -like genes that may be present in native human DNA. The C_{θ} 1/2 of the γ cDNA hybridization is two- to threefold less than that of the total unique sequence DNA and indicate that γ -globin genes are present in very few copies. From the data obtained by saturation hybridization analysis, one to two γ globin genes per haploid genome are calculated in these studies (Table II). All of the results are consistent with the two to four γ globin genes per haploid genome, estimated from genetic data utilizing the relative amounts of γ^{Glv} and γ^{ala} in human subjects (19).

Since this work was completed, γ -globin gene measurements similar to ours have been reported with hybridization in a vast excess of cDNA (20). In this report (20), the authors state that there is less variability in gene numbers and greater sensitivity with a vast excess of cDNA. However, these same authors were unable to detect a deletion of β -like globin genes in a heterozygote for $\delta\beta$ -thalassemia with excess cDNA hybridization (5), while we have been able to do so easily using the methodology described in this paper (8).

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