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### Research Article

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# Metabolism of Prostaglandins A<sub>1</sub> and E<sub>1</sub> in Man

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**ABSTRACT** To investigate the *in vivo* whole blood metabolic clearance rates and sites of metabolism of prostaglandins A<sub>1</sub> and E<sub>1</sub> in man, constant infusions of the tritiated compounds were administered to normal subjects and to patients undergoing cardiac catheterization. The whole blood metabolic clearance rate of [<sup>3</sup>H]prostaglandin A<sub>1</sub> in eight men was 5,003±864 liters/day (SD) or 2,546±513 liters/day per m<sup>2</sup> (SD). Nonradioactive prostaglandin A<sub>1</sub> was similarly infused in two subjects, and the metabolic clearance rates were determined, utilizing a specific radioimmunoassay. The clearance rates with this method correlated closely with those determined by the isotope infusions. Extraction studies of prostaglandin A<sub>1</sub> showed that pulmonary, splanchnic, renal, and extremity perfusions resulted in 8.1±4.1, 56.1±10.1, 50.3±3.4, and 34.4±5.9% (SEM) removal, respectively. With [<sup>3</sup>H]prostaglandin E<sub>1</sub>, the whole blood metabolic clearance rate was determined from the pulmonary artery concentration in three patients and averaged 4,832±1,518 liters/day (SD) or 2,686±654 liters/day per m<sup>2</sup> (SD). Pulmonary extraction was 67.8±6.8% (SEM) and extremity removal averaged 6.6±4.9% (SEM).

These results indicate that A prostaglandins are metabolized by several organs, such as the liver and kidney, and possibly by intravascular pathways as well. In man, the E prostaglandins are primarily metabolized by the lung, but extraction is not complete and approximately one-third may escape lung metabolism. Thus, these findings suggest that both E and A prostaglandins in the venous circulation may reach the systemic circulation in man.

This work was presented in part at the 56th Annual Meeting of The Endocrine Society at Atlanta, Ga. on 14 June 1974 and published in abstract form (1).

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## INTRODUCTION

The hypothesis that renal prostaglandins reach the arterial circulation could have important physiologic consequences. The E and A prostaglandins, when infused intravenously at some doses, have natriuretic and vaso-depressor actions (2). However, at certain infusion rates, prostaglandin A<sub>1</sub> (PGA<sub>1</sub>)<sup>1</sup> appears to selectively stimulate aldosterone secretion (3, 4). This suggests the possibility that A or E prostaglandins and angiotensin II could act in concert to regulate mineralocorticoid production while antagonizing each others effect on blood pressure (5). The presence of circulating prostaglandins could therefore have important effects upon both blood pressure and fluid volume control.

In the cat and dog, it has been demonstrated that E prostaglandins are rapidly metabolized by the lung, whereas A prostaglandins appear to escape this degradation (6, 7). This difference in metabolism has led to the speculation that A prostaglandins may function as circulating hormones. In man, Hamberg and Samuelsson have shown that tritiated prostaglandin E<sub>2</sub> (PGE<sub>2</sub>), injected as a bolus, is rapidly cleared from the circulation with the formation of the less polar metabolite, 13, 14-dihydro-15-keto PGE<sub>2</sub>, in blood (8). As pulmonary tissue contains the enzymes necessary for this conversion (9), it has been presumed that the lung is the first organ of metabolism of E prostaglandins, and this has recently been confirmed *in vivo* in the dog (10). Studies of the clearance of A prostaglandins from the human circulation or the sites of this removal have not been available.

<sup>1</sup>Abbreviations and trivial names used in this paper: MCR, metabolic clearance rate; PG, prostaglandin; PGA<sub>1</sub>, prostaglandin A<sub>1</sub> (15-hydroxy-9-ketoprost-10,13-dienoic acid); PGA<sub>2</sub>, prostaglandin A<sub>2</sub> (15-hydroxy-9-ketoprost-5,10,13-trienoic acid); PGE<sub>1</sub>, prostaglandin E<sub>1</sub>, (11 $\alpha$ ,15-dihydroxy-9-ketoprost-13-enoic acid); PGE<sub>2</sub>, prostaglandin E<sub>2</sub> (11 $\alpha$ ,15-dihydroxy-9-ketoprost-5,13-dienoic acid); 13, 14-dihydro-15-keto PGE<sub>2</sub>, 11 $\alpha$ -hydroxy-9,15-diketoprost-5-enoic acid.

This study was designed to determine the metabolic clearance rate in blood and the major *in vivo* sites of metabolism of  $\text{PGA}_1$  and  $\text{PGE}_1$  in man.

## METHODS

**Materials.** Spectrograde methanol and dichloromethane were purchased from Matheson Coleman & Bell (East Rutherford, N. J.).

$[^3\text{H}]\text{PGA}_1$  (sp act 384.6  $\mu\text{Ci}/\mu\text{g}$ ),  $[^3\text{H}]\text{PGE}_1$  (sp act 312.5  $\mu\text{Ci}/\mu\text{g}$ ), and  $[^{14}\text{C}]\text{PGE}_1$  (sp act 1.41  $\mu\text{Ci}/\mu\text{g}$ ) were purchased from New England Nuclear (Boston, Mass.).  $[^{14}\text{C}]\text{PGA}_1$  was chemically converted from  $[^{14}\text{C}]\text{PGE}_1$  by acidification according to the method of Andersen (11) and then purified by an LH-20 Sephadex chromatography system as previously described (12).

**Radiochemical purity.** The radioactive prostaglandins were periodically checked for purity by the demonstration of a single elution peak from an  $80 \times 1\text{-cm}$  LH-20 Sephadex column using the system methylene chloride, methanol (95:5). In addition, the  $[^3\text{H}]\text{PGA}_1$  was checked against  $[^{14}\text{C}]\text{PGA}_1$  by mixing quantities of them together. An aliquot of the mixture was counted and the remainder was chromatographed. The "A fraction" (from 65 to 85 ml) was collected and counted. (Original  $^3\text{H}/^{14}\text{C}$  ratio: 4.10; chromatographed ratio: 4.08.) Similarly,  $[^3\text{H}]\text{PGE}_1$  and  $[^{14}\text{C}]\text{PGE}_1$  were mixed, and an aliquot was chromatographed with collection and counting of the "E fraction" (105–135 ml). (Original  $^3\text{H}/^{14}\text{C}$  ratio: 3.84; chromatographed ratio: 3.75.)

**Subjects.** The metabolic clearance rate (MCR) of  $\text{PGA}_1$  was determined in five healthy men who gave informed consent.

Studies of the sites of metabolism of both  $\text{PGA}_1$  and  $\text{PGE}_1$  were performed on 10 patients (6 men and 4 women) who were scheduled for elective cardiac catheterization. All the patients were given a full explanation of the study, and all volunteered to participate. At the time of the study, the patients were clinically stable with no evidence of congestive heart failure, hypertension, or significant regurgitant valvular lesions. Serum creatinine and liver function tests were normal. The cardiac index of these patients was only slightly reduced, averaging 2.9 liters/min per  $\text{m}^2$ . The cardiologic diagnoses are presented in Table III.

**Constant infusion method.** Constant rate infusions of  $[^3\text{H}]\text{PGA}_1$  and  $[^3\text{H}]\text{PGE}_1$  were performed in a manner similar to that employed for the determination of the metabolic clearance rates of various steroids (13). All infusions were administered to the subjects in the morning, after an overnight fast. The patients maintained the supine position throughout the infusion period. Infusion solutions were prepared by dissolving 20  $\mu\text{Ci}$  of  $[^3\text{H}]\text{PGA}_1$  or 80  $\mu\text{Ci}$  of  $[^3\text{H}]\text{PGE}_1$  in 4 ml of absolute ethyl alcohol and then diluting this in 50 ml of normal saline. The infusions were begun in an antecubital vein with a 10% priming dose of this solution and they were continued at a rate of 0.382 ml/min (infusion pump, Harvard Apparatus Co., Inc., Millis, Mass.). The calibration of the pump was monitored and found to be accurate. During the infusions, 5 ml of heparinized whole blood was collected on ice from the opposite arm at 90, 105, and 120 min. At the conclusion of each study, infusion fluid was collected from the patient's intravenous tubing. As the concentration of tritiated prostaglandin in this fluid was nearly identical to that calculated from the amount of counts and diluent added, it did not appear that there was appreciable adsorption of the labeled prostaglandins onto the surfaces of the infusion system.

Samples of the infusion solution were rechromatographed and retained their characteristic elution pattern.

In two subjects, the MCR was also calculated by infusions of nonradioactive  $\text{PGA}_1$ . The first study was performed by the infusion of four different doses for 1-h periods. At the end of each period, blood was obtained for the measurement of  $\text{PGA}_1$  concentration with a specific radioimmunoassay (14). The infusion fluid was appropriately diluted and measured in the same radioimmunoassay. The second subject's MCR was calculated from a 1-h infusion of a single dose of nonradioactive  $\text{PGA}_1$ .

In the 10 catheterization studies, the constant infusions were begun in the antecubital vein as soon as the patient was positioned on the radiography table. The cardiology staff then proceeded with indicated studies with catheter positioning performed by the percutaneous technique. In all the studies, catheters were placed in the pulmonary artery and the left ventricle or ascending aorta. In four studies, a catheter was placed in the hepatic vein and in three studies in a renal vein. Blood was drawn from these catheters and a peripheral vein at 90, 105, and 120 min, when possible.

**Sample handling.** Whole blood was utilized because of the possibility that infused  $\text{PGA}_1$  may diffuse into the cellular elements of blood (14). Additionally, we have reported evidence for metabolism of A prostaglandins by whole blood *in vitro* (15). The use of whole blood allowed the immediate introduction of the second isotope to correct for this potential *in vitro* source of error.

The 5-ml blood samples were collected in iced heparinized syringes and immediately lysed by addition to a 100-ml conical centrifuge tube containing 5 ml of deionized water and  $^{14}\text{C}$ -labeled indicator (approximately 500 cpm). This mixture was acidified with the addition of 1 ml of 0.5 M phosphate buffer, pH 4.5. Extraction was quickly performed by adding 80 ml of 5% methanol in dichloromethane and shaking vigorously. After centrifugation at 2,000 rpm for 10 min, the aqueous layer was decanted, and the organic solvent was separated from the overlying precipitate by filtration through glass wool and anhydrous sodium sulfate. After the organic solvent was evaporated in a vacuum apparatus (Rinco Instruments Co., Inc., Greenville, Ill.), the samples were chromatographed on a Sephadex LH-20 system (12). The A fraction (from 65 to 85 ml) or E fraction (105–135 ml) was collected, dried under nitrogen, and counted in 10 ml of 2,5-diphenyloxazole-1,4-bis[2-(5-phenyloxazolyl)]benzene (Spectrafluor, Amersham/Searle Corp., Arlington Heights, Ill.) in toluene with a Mark I dual channel scintillation counter (Nuclear-Chicago Corp., Des Plaines, Ill.) ( $^3\text{H}$  efficiency, 35%.  $^{14}\text{C}$  efficiency 68%;  $^{14}\text{C}$  to  $^3\text{H}$  feedthrough, 15%). Samples were counted for a period long enough to reduce the error of counting to less than 2% (40 min, three times) (16). The recoveries for  $\text{PGA}_1$  and  $\text{PGE}_1$  averaged 44.9% (38.3–55.3) and 34.2% (30.5–41.4), respectively.

The purity of the chromatographic peaks was demonstrated on several studies by fractionation of the collection of the peaks and showing uniformity of the  $^3\text{H}/^{14}\text{C}$  ratio.

When plasma measurements were made, iced heparinized blood was immediately centrifuged at  $0^\circ\text{C}$  for 10 min at 2,000 rpm. Rapid preparation of the plasma was felt to be critical in minimizing *in vitro* metabolism. The extraction procedure for plasma was identical to that of whole blood.

**Calculations.** Whole blood MCRs were determined by the formula  $\text{MCR (liters/day)} = \text{infusion rate (counts/day)} / \text{whole blood concentration (counts/liter)}$ . The infusion rate was calculated by counting several 0.1-ml dried samples of infusion fluid and multiplying by the infusion

rate of the pump. The whole blood concentration of tritiated prostaglandin was determined from the counts of the appropriate chromatographic fraction by subtracting the counts due to background and  $^{14}\text{C}$ - $^3\text{H}$  feedthrough and correcting for losses as calculated from the  $^{14}\text{C}$  counts.

The concentration of prostaglandin counts at various points in the circulation was used to calculate the extraction accomplished by the circulation through several organs. Thus, the percent extracted by the lung was equal to: (pulmonary artery concn.)-(left ventricle concn.)/(pulmonary artery concn.) $\times 100$ . The percent extracted by a peripheral organ was equal to: (left ventricle concn.)-(venous drainage of organ concn.)/(left ventricle concn.) $\times 100$ .

The clearance rate of an organ was calculated as the product of the extraction fraction and the estimated blood flow through the organ in the basal state.

## RESULTS

**MCRs.** The whole blood MCR of  $\text{PGA}_1$  in five normal men was  $5,052 \pm 914$  liters/day (SD). When expressed in terms of body surface area, the value was  $2,600 \pm 482$  liters/day per  $\text{m}^2$  (SD). In three men undergoing catheterization for cardiac diagnosis, the same calculations could be performed, and no significant difference in the data was noted. The overall MCR in the eight men was  $5,003 \pm 864$  liters/day (SD) or  $2,546 \pm 513$  liters/day per  $\text{m}^2$  (SD) (Table I). The MCR of  $\text{PGA}_1$  was calculated in only one woman (M. F.), a cardiac patient with a slightly decreased cardiac index. Her MCR of 2,469 liters/day or 1,543 liters/day per  $\text{m}^2$  was the lowest individual value measured, although it was

within 2 SDs of the mean of the male clearances when corrected for body surface area.

There was a trend in the data with an average difference of  $15.8 \pm 11.2\%$  (SD) between the 90- and 120-min samples. As a priming dose was used, failure to reach equilibrium should be reflected by falling concentrations. The actual trend was in the opposite direction and may reflect a recirculation effect or a changing cardiac output. The latter was felt to be the major factor in the greater variability seen in those patients undergoing cardiac catheterization. In one subject (D. R.), the infusion was extended to a total of 4 h and additional samples were drawn at 210 and 240 min. The MCR calculated at 90 min differed only by 5.9% from that determined at 240 min. The maximum variation from the mean of all five samples was 6.1%. This indicates that equilibrium most probably had been reached by 90 min, as would be expected of a compound with a high MCR, and that the method was reproducible with the subject in a basal state.

Aside from the  $\text{PGA}_1$  region, no significant tritium was eluted from the chromatography columns in these studies except for a minor ( $< 5\%$ ), less polar peak.

In a healthy man (G. B.), 1-h infusions of four different doses of nonradioactive  $\text{PGA}_1$  were used to calculate the MCR. The MCR results (Table II) were consistent at all four doses, with a maximum variation of 7.5% from the mean. The average MCR of the four infusion rates was  $5,784 \pm 363$  liters/day (SD) or  $3,324 \pm$

TABLE I  
MCR of  $[^3\text{H}]\text{PGA}_1$

Subject	Sex	Age	Infusion rate	Blood counts/liter $\times 10^{-3}$				MCR	MCR
				90 min	105 min	120 min	Mean		
		<i>yr</i>	<i>counts/day</i> $\times 10^{-3}$					<i>liter/day</i>	<i>liters/day per m<sup>2</sup></i>
Normal subjects									
G. L.	M	26	1.39		28.4	32.7	30.6	4,562	2,258
M. T.	M	26	1.47	37.9		37.7	37.8	3,889	2,102
N. R.	M	30	1.04	15.2	17.9	18.7	17.3	6,099	3,227
R. H.	M	41	1.12	22.1		23.7	22.9	4,870	2,435
D. R.	M	36	2.01	33.8	36.6	36.5	35.5*	5,841	2,980
Mean ( $\pm$ SD)								5,052 $\pm$ 914	2,600 $\pm$ 482
Catheterized patients									
R. B.	M	45	1.75	28.7	32.0	37.8	32.8	5,411	2,552
V. G.	M	26	1.75	40.5	49.2	49.1	46.3	3,813	1,749
V. R.	M	41	1.79		32.3	32.3	32.3	5,540	3,061
Overall mean ( $\pm$ SD)								5,003 $\pm$ 864	2,546 $\pm$ 513
Female patient									
M. F.	F	56	1.62	58.6	69.1	70.4	66.0	2,469	1,543

\* Includes values from 210 and 240 min.

TABLE II  
MCR of Nonradioactive  $\text{PGA}_1$

Subject	Infusion rate	$\text{PGA}_1$ concentration by radio- immunoassay*	MCR
		$\mu\text{g}/\text{liter}$	
G. B.	$\mu\text{g}/\text{day}$		
	7,020	1.19	5,899
	14,040	2.59	5,421
	28,080	4.50	6,240
	56,160	10.07	5,577
		Mean ( $\pm$ SD)	5,784 $\pm$ 363
D. R.	7,401	1.21	6,117

\* Endogenous concentration of 0.028  $\mu\text{g}/\text{liter}$  (G. B.) and 0.025  $\mu\text{g}/\text{liter}$  (D. R.) subtracted.

209 liters/day per  $\text{m}^2$  (SD). Simultaneous plasma concentrations were measured at the completion of two of the infusions, and the values were very similar to the whole blood concentrations (1.23 vs. 1.19  $\mu\text{g}/\text{liter}$  and 5.01 vs. 4.50  $\mu\text{g}/\text{liter}$ ), suggesting that  $\text{PGA}_1$  distributes equally throughout blood. In a second subject (D. R.), the MCR via the nonradioactive technique was 6,117 liters/day compared to 5,841 determined previously by the isotopic method; a difference of only 5%.

The whole blood MCR of  $\text{PGE}_1$  could not be determined from peripheral blood concentrations as this site is distal to a major organ of metabolism (the lung). When the pulmonary artery concentration was used to calculate the MCR, the average in three cardiac patients was  $4,832 \pm 1,518$  liters/day (SD) or  $2,686 \pm 654$  liters/day per  $\text{m}^2$  (SD).

**Organ extraction and clearance of  $\text{PGA}_1$  and  $\text{PGE}_1$ .** In seven studies, extraction of  $\text{PGA}_1$  by the lung averaged  $8.1 \pm 4.1\%$  (SEM), which was not statistically different from 0, ( $P > 0.05$ ). In contrast, sampling from the venous drainage of the liver, kidney, and extremities showed significant extraction by these organs. The extraction by the splanchnic circulation averaged  $56.1 \pm 10.1\%$  (SEM) in four patients ( $P < 0.05$ ), the renal extraction was  $50.3 \pm 3.4\%$  (SEM) in three patients ( $P < 0.01$ ), and the extremity extraction was  $34.4 \pm 5.9\%$  (SEM) in six patients ( $P < 0.01$ ). (Fig. 1 and Table III).

Organ clearance of  $\text{PGA}_1$  was calculated as the product of extraction and blood flow through each organ, assuming a basal cardiac output of 3.3 liters/min per  $\text{m}^2$  (4,752 liters/day per  $\text{m}^2$ ) (17) and that the perfusion of the pulmonary, splanchnic and renal circulations represented 100, 38, and 20%, respectively, of the cardiac output (18, 19). The averages were: pulmonary clearance 270 liters/day per  $\text{m}^2$ ; splanchnic clearance, 700 liters/day per  $\text{m}^2$ ; and renal clearance, 330 liters/day

per  $\text{m}^2$ . Thus, the total clearance attributable to the pulmonary, splanchnic, and renal circulations totaled 1,300 liters/day per  $\text{m}^2$ , only slightly more than half of the total clearance.

In three patients, the pulmonary extraction of infused tritiated  $\text{PGE}_1$  averaged  $67.8 \pm 6.8\%$  (SEM) ( $P < 0.01$ ). The extraction of the upper extremity averaged  $6.6 \pm 4.9\%$  (SEM) ( $P > 0.05$ ) (Fig. 1 and Table III).

## DISCUSSION

In several species, the biologic potencies of E prostaglandins have been found to be much greater via intra-aortic injection than by the intravenous route (20, 21). Thus, it became clear that the lung probably played a major role in degrading these compounds. The blood-bathed organ perfusion technique subsequently confirmed the capacity of the lung to metabolize E and F prostaglandins (22). However, the A prostaglandins have nearly equal effects when injected intra-arterially or intravenously (6, 7). The action of the enzyme 15-hydroxyprostaglandin dehydrogenase appears to be the first step in the degradation of E and F prostaglandins. This enzyme, which metabolizes A prostaglandins at a somewhat slower rate (23), has been found in high concentrations in the lung, spleen and kidney cortex of the pig (24). The 15-keto derivatives of E and F prostaglandins may be further modified by reduction at the 13, 14-double bond by a reductase enzyme present in several tissues of the swine (9). The 13,14-dihydro-15-keto derivative of  $\text{PGE}_2$  was found to accumulate rapidly in the blood of man after an intravenous injection of the tritiated prostaglandin (8). Further metabolism of the E and F prostaglandins appears to be oxidative

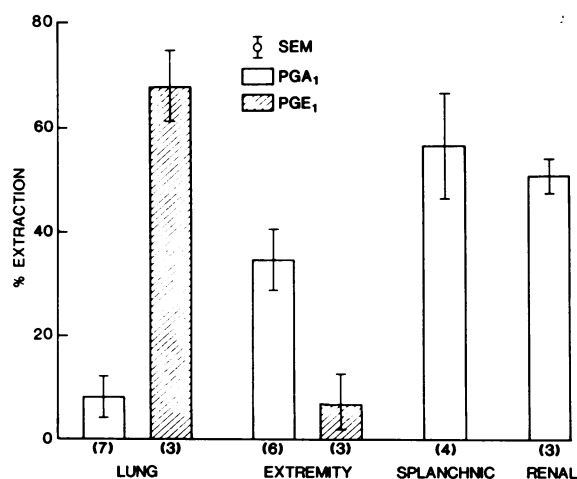


FIGURE 1 Percent extraction of  $\text{PGA}_1$  and  $\text{PGE}_1$  by perfusion of the lung, extremity, splanchnic region, and kidney. Numbers in parentheses indicate number of studies performed.

TABLE III  
Extraction Studies of [ $^3\text{H}$ ]PGA<sub>1</sub> and [ $^3\text{H}$ ]PGE<sub>1</sub>

Subject	Sex	Age	Diagnosis	Sites counts/liters $\times 10^{-3}$ *					Extraction, %			
				Pulmonary artery	Left ventricle	Periph-eral vein	Hepatic vein	Renal vein	Pulmonary	Extremity	Splanchnic	Renal
[ <sup>3</sup> H]PGA <sub>1</sub>												
M. F.	F	56	Mitral stenosis	121.4	93.5	66.0	53.4		23.0	29.3	42.3	
			Atrial fibrillation									
R. B.	M	45	Angina	50.8	45.3	32.8	13.8		10.8	27.5	69.5	
F. W.	M	37	Ventricular aneurysm	41.9	43.3	19.5‡	10.1		0	55.0‡	76.7	
V. G.	M	26	Ventricular aneurysm	52.8	56.8	46.3	36.5		0	13.6	35.7	
M. P.	F	29	Prosthetic mitral valve	123.8	95.2			48.5	23.1			49.1
			Atrial fibrillation									
K. H.	M	43	Angina	47.7	48.5	26.5‡		21.0	0	45.4‡		56.7
V. R.	M	41	Mitral stenosis	49.8	50.1	32.3		27.5	0	35.5		45.1
							Mean ( $\pm$ SEM)		8.1 $\pm$ 4.1	34.4 $\pm$ 5.9	56.1 $\pm$ 10.1	50.3 $\pm$ 3.4
							P§		>0.05	<0.01	<0.05	<0.01
[ <sup>3</sup> H]PGE <sub>1</sub>												
C. B.	F	71	Mitral stenosis	136.2	62.1	52.1			54.4	16.1		
W. R.	M	62	Angina	82.4	19.5	18.8			76.3	3.6		
L. F.	F	51	Angina	138.3	37.7	38.1			72.7	0		
							Mean ( $\pm$ SEM)		67.8 $\pm$ 6.8	6.6 $\pm$ 4.9		
							P§		<0.01	>0.05		

\* Mean value of determinations at 90, 105, and 120 min.

‡ Peripheral blood obtained from the femoral vein.

§ Values compared to 0.

with the formation of polar metabolites which are cleared into the urine. The liver, a source of oxidative enzymes, is also capable of inactivating a large fraction of E and A prostaglandins in single passage perfusion studies in the cat (6).

Little is known of the metabolic pathway for PGA degradation, but the kidney is a potential organ of metabolism. In the rabbit, injection of PGA<sub>2</sub> into the renal artery has resulted in quantitative conversion to less polar metabolites in the renal venous blood (25).

Enzymes in the blood may also play a role in prostaglandin interconversions and degradation. Jones and Cammock have demonstrated that the plasma of several species is capable of converting A to B prostaglandins, probably with C prostaglandins as an intermediary product (26). However, in recent studies it has been shown that the reaction rates of A to C isomerization and E to A conversion are quite slow in human serum (27). In vitro studies with whole blood in our laboratory indicate that the cellular elements of the blood may be capable of metabolizing A prostaglandins (15).

The existence of A prostaglandins in vivo remains an open question (28, 29). Although several radioimmunoassays measure A prostaglandins, a recent study with gas chromatography-mass spectrometry was unable to detect PGA<sub>2</sub> in human plasma (sensitivity of 200 pg/ml)

(30). If they are present, A prostaglandins may result from the enzymatic conversion of E prostaglandins. Robertson has reported that the infusion of PGE<sub>2</sub> in man elevates plasma radioimmunoassayable prostaglandin A concentrations (31). In the present study, chromatography of the blood extracts from the [ $^3\text{H}$ ]PGE<sub>1</sub> infusions demonstrated the presence of three peaks, one with the mobility of [ $^3\text{H}$ ]PGE<sub>1</sub> and two that were less polar. The larger and more polar of these two other peaks had a mobility similar to that of an A prostaglandin. An antibody quite specific for PGA<sub>1</sub> (14) was added in high concentration to the counts of this peak, and the bound fraction was precipitated with ammonium sulfate. Compared to [ $^3\text{H}$ ]PGA<sub>1</sub>, the counts of this peak were only partially precipitated with the antibody.

Two methods were used to measure the clearance rate of PGA<sub>1</sub> in this study. The infused dosage (0.125-1.0  $\mu\text{g/kg}$  per min) of nonradioactive PGA<sub>1</sub> has been shown to increase renal blood flow (2) and was observed to increase the pulse without changing the blood pressure in our subjects. Although the clearance rate was similar for four different infusion rates, the biological activity of this material could have influenced the clearance rate. However, this method verified the results obtained with the radioactive method in which tracer quan-

titities were infused (less than 25 ng/h of PGA<sub>1</sub> and less than 125 ng/h of PGE<sub>1</sub>). No changes in pulse or blood pressure were observed with these infusions.

The results of the present study indicate that the clearance rates of PGA<sub>1</sub> and PGE<sub>1</sub> are quite high, in excess of the blood flow to any organ except the heart and lung. The identification of the extracted isotopes was made chromatographically. Thus, if there were conversion to a metabolite with nearly identical chromatographic properties (i.e. A to B isomerization), this technique would not recognize this metabolism. In this sense, the calculated MCR represents a minimal value.

In this context, the high clearances of PGA<sub>1</sub> and PGE<sub>1</sub> would lead us to predict that these compounds are cleared either by pulmonary metabolism or by metabolism in multiple organs. A third possibility could be intravascular metabolism. In addition, the high clearances would be compatible with the previously demonstrated low affinity albumin binding of these substances (32).

The extraction studies have demonstrated that man metabolizes E and A prostaglandins in different ways. With PGA<sub>1</sub>, there was minimal evidence of pulmonary clearance, but perfusion of the splanchnic and renal circulations resulted in significant extraction. As these extractions can explain only a portion of PGA<sub>1</sub> clearance, these findings can be interpreted to indicate that multiple peripheral organs are involved in the metabolism of PGA<sub>1</sub>. However, the extraction in the extremities and the demonstrated capacity of the blood to metabolize A prostaglandins in vitro suggests that intravascular metabolism may also play a role.

In contrast, the studies with PGE<sub>1</sub> showed that the lung was a major organ of metabolism and that there was little or no extraction by the extremities. PGE<sub>1</sub> in the arterial circulation presumably could be metabolized by the liver or other organs as well.

Perhaps the most significant findings of these studies was the demonstration that constant intravenous infusion of both substances resulted in the recovery of appreciable counts of the infused material in the peripheral circulation. Although the extraction of PGE<sub>1</sub> by the lung was considerable, it was not complete. This may be important in view of the findings that various stimuli are capable of releasing E prostaglandins into the renal vein (29). Thus, prostaglandins of either the A or E series, if released into the venous circulation, have the potential to reach and affect the systemic vasculature and distant organs.

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