On the Influence of Extracellular Fluid Volume Expansion on

Bicarbonate Reabsorption in the Rat

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ABSTRACT Bicarbonate reabsorption is classically regarded as a rate-limited process characterized by saturation kinetics. The tubular maximum (Tm), however, varies with glomerular filtration rate. Thus bicarbonate reabsorption, in common with sodium reabsorption, is characterized by glomerulo-tubular balance. The examination of bicarbonate reabsorption is accomplished using the bicarbonate titration technique; however, this method in its traditional form leads to marked expansion of extracellular fluid (ECF) volume. The possibility exists, therefore, that glomerulo-tubular balance for bicarbonate is altered by the volume expansion and thus that the classic pattern of reabsorption may actually reflect inhibited bicarbonate reabsorptive capacity. The present studies were performed in rats to examine this possibility. Bicarbonate titration studies were performed in two groups of animals: (a) those in which ECF volume expansion was minimized; and (b) those in which ECF volume expansion was exaggerated. In the first group, no Tm for bicarbonate was observed either in the majority of individual rats studied or in a group plot for all rats studied despite the fact that plasma bicarbonate concentrations were increased to values in excess of 60 mEq/liter. In the second group, a clear Tm was demonstrated both in individual animals and in group data and there was a lowered threshold for the excretion of bicarbonate. The data thus lend support to the view that the "normal" Tm for bicarbonate may actually represent an inhibited level of bicarbonate reabsorption induced by ECF volume expansion.

INTRODUCTION

The reabsorption of bicarbonate by the mammalian kidney is believed to be characterized by saturation kinetics (1, 2). The relationship between the apparent maximum velocity of transport and the substrate (i.e. bicarbonate) concentration of the glomerular filtrate is not a simple one however, in that the tubular maximum (Tm) for bicarbonate varies with glomerular filtration rate (GFR). Thus, under experimental conditions wherein a Tm is demonstrable, there appears to exist a form of glomerulo-tubular balance for bicarbonate which is analogous to glomerulo-tubular balance for sodium. To delineate the normal pattern for bicarbonate reabsorption, the bicarbonate titration technique has been employed. However, this procedure in its traditional form involves substantial expansion of extracellular fluid (ECF) volume. Since expansion of extracellular fluid volume has been shown to have profound effects on proximal tubular sodium reabsorption, and indeed to reset glomerulo-tubular balance for sodium, the possibility exists that the accepted "normal" pattern for bicarbonate reabsorption may be influenced by the experimental method. Accordingly, the present studies were undertaken to reexamine bicarbonate reabsorption using a bicarbonate titration technique which minimized extracellular fluid volume expansion. Studies also were performed during exaggerated ECF volume expansion.

METHODS

Experiments were performed on unanesthetized female Sprague-Dawley rats weighing between 225 and 250 g. Two types of titration experiments were performed. The first was designed to restrict the expansion of ECF volume to the minimal level consistent with obtaining the appropriate stepwise elevation of plasma bicarbonate concentrations. In the second type of titration experiment ECF expansion was exaggerated.

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The preparation of the animals for study was accomplished in the manner described in detail previously (3). The rats were anesthetized lightly with ether to allow for the insertion of arterial, venous, and bladder catheters. After completion of the surgical procedures, the anesthesia was discontinued and a period of $1\frac{1}{2}$ -2 hr was allowed for the animals to recover completely from the anesthetic. Urine was collected through a soft silastic catheter (0.D. 1.25 mm).

All of the sustaining infusions contained a sodium concentration of 140 mEq/liter. The rate of bicarbonate administration was adjusted by increasing the bicarbonate concentration of the infusate progressively from 0 (in the control periods)

TABLE I

Representative Bicarbonate Titration Experiment during Minimal ECF Volume Expansion

Clearance				Plasma			Urine					
period	Time	GFR	pН	HCO3	pCO2	pН	HCO3	pCO2	HCO3 ez	kcretion	HCO3 rea	bsorption
	min	ml/min		µEq/ml	mm Hg		µEq/ml	mm Hg	µEq/min	µEq/ml GFR	µEq/min	µEq/ml GFR
	-175	Light e	ther an	esthesia fo animal in	or insertio holder ()	on of tail duratior	vein and f 25 min)	emoral ar	tery cannı	ılae, blad	lder cathet	er, and
	- 60	Inulin	prime ($0.71 \ \mu c$ in	ulin- ¹⁴ C in	n 1 ml n 1 wo MC	ormal salin	ne normal sa	line at 0 1	11 ml/m		
1	0-30	3 00	7 4 1	24 0	40 0	6 70	2 83	27 0	0 46	0 15	77 4	25.1
-	31-49	Sustai	ning sol	ution con	taining N	JaHCO.	30 mFa/li	iter at 0 1	1 ml/min	0.15	11.1	25.1
2	49-65	2 88	7 44	24 0	36.5	6 88	3 Q7	74 8	0 70	0.24	71 0	25.0
3	65-77	3.61	7 4 5	24.0	36.0	6.90	4 84	24.0	0.10	0.24	80 0	23.0
Ŭ	82-186	Prime	0.3 ml 1	21.0 15 x Nak	4CO. (04	15 mFa	HC0) e	ustaining	solution c	ontainin		. 40 mFa/
	02 100	52-150 Prime U.S mi 1.5 M NAHCU ₃ (U.45 mEq HCU ₃ ⁻), sustaining solution containing NaHCO ₃ 40 mEq/									3 40 mEq/	
4	186-200	2 64	7 53	29.0	36.5	7 20	9.88	28.5	1 77	0.67	78.6	20.8
5	200-211	3.61	7 54	29.5	36.0	7 25	10.4	20.5	1.80	0.52	100.8	30.5
Ũ	218-238	Prime	0.3 ml	15 M Na	HCO. (() 45 mE		sustainir	ng solution	1 contair	ing NaH(CO. 40
	210 200	mE	ı∕liter a	1.0 m 100 m 1000 m 100	/min		q/1100,),	, sustaini	ig solution	i contan	ing mari	0,10
6	238-250	3 52	7 54	29.5	36.0	7.30	14.0	28.5	2 21	0.63	106.8	30.4
7	250-261	3.27	7.53	29.7	37.0	7.30	11.2	25.0	2.03	0.62	99.9	30.6
•	266-289	Prime	0.3 ml	15 M Na	нсо, (() 45 mE		sustainir	or solution	1 contair	ning NaH(CO. 80
	200 207	mE	∖liter a	1.0 m 100 m 100	/min		q 1100 .),	, sustanni	ig solution	reontan	ing runi	20,00
8	289-317	3 11	7 60	32.2	34.0	7.60	36.8	36.0	3 94	1 27	101.6	327
Q	317-330	3 78	7 50	33.1	35.8	7.60	36.3	38.0	4 46	1 1 8	126.0	33.6
	334-350	Prime	0.4 ml	15 M Na		0.60 mF		sustainin	a solution	containi	ing NaHC	0.100
	001 000	mE	ı∕liter a	1.0 M I.	/min	0.00 mL	,q 11003),	Sustannin	5 Solution	contain	ing marie	03 100
10	350-362	2 60	7 61	39.0	40.5	7 85	84 7	45.5	5 68	2.18	100.8	38.8
11	362-374	3.18	7.59	38.7	41.0	7.85	80.0	45.0	8.64	2.72	120.6	37.9
	379-392	Prime	0.4 ml	15 м Na		60 mEo		sustaining	r solution	containii	ng NaHC(-100
	0	mEe	/liter a	at 0.11 ml	/min			ouocurre	, contraction	001111111		
12	392-403	2.99	7.61	40.8	41.5	7.95	117.2	50.0	8.56	2.86	119.5	40.0
13	403-416	2.55	7.60	39.0	41.3	7.85	72.9	41.0	5.90	2.31	98.5	38.7
	421-436	Prime	0.4 ml	м NaHC()₃ (0.60 r	nEa HC	O ₂). sustai	ning solut	tion conta	ining Na	HCO ₃ 100	mEa/
		liter	0.11 m	1/min			- •//					
14	436-447	3.16	7.62	43.0	43.3	7.85	82.9	44.5	6.63	2.10	136.0	43.1
15	447-461	2.78	7.62	43.0	43.3	7.90	82.9	42.5	8.29	2.98	117.2	42.2
	466-491	Prime	0.4 ml	1.5 м Na	HCO ₃ (0	.60 mEa	h HCO₁ [−]).	sustainin	g solution	containi	ing NaHC	O ₂ 120
		mEe	ı/liter a	nt 0.11 ml	/min				8			
16	491-502	3.12	7.65	47.0	45.5	7.75	75.9	55.0	8.28	2.65	145.7	46.7
17	502-523	2.58	7.64	45.6	45.5	7.75	69.6	54.0	8.63	3.34	114.9	44.5
	526-549	Prime	0.4 ml	1.5 м Na	HCO ₂ ((0.60 mE	a HCO₃).	sustaining	g solution	containi	ng NaHC	O ₂ 120
		mE	a∕liter ≄	at 0.11 ml	/min		·1 · 3/)				0	
18	549-564	2.46	7.65	49.2	46.8	7.80	113.5	70.0	11.4	4.61	115.8	47.0
19	564-575	2.57	7.65	49.7	46.5	7.90	94.6	52.0	13.7	5.34	120.4	46.9
	577-601	Prime	0.4 ml	1.5 м Na	HCO ₂ (0	.60 mEa	HCO₃ [−]).	sustainin	g solution	contain	ing NaHC	O ₃ 120
		mE	g/liter a	at 0.11 ml	/min		//				0	•
20	601-620	2.59	7.71	57.7	47.8	7.80	132.8	80.0	13.3	5.13	143.6	55.5
21	620-637	2.49	7.68	57.5	50.0	7.80	65.8	43.5	8.16	3.28	142.2	57.1

Rat weight, 220 g.



FIGURE 1 Mean bicarbonate titration curves for nine animals studied under conditions of minimized ECF volume expansion.

to 120 mEq/liter. The concentration of chloride, the only other anion, was changed reciprocally. 15-20 clearance periods, each 10-30 min in duration, were obtained. Observations were made over a range of plasma bicarbonate concentrations extending from values as low as 11 mEq/liter to values as high as 64 mEq/liter. Two to three clearance periods were obtained at each level of bicarbonate infusion. In approximately half of the experiments 2.5% ammonium chloride was administered in the drinking water the night before study to effect a decrease in the initial plasma bicarbonate concentrations to subnormal levels.

Glomerular filtration rate was measured using carboxyllabeled inulin-¹⁴C. A priming dose of 0.7 μ c of inulin-¹⁴C in 1 ml of isotonic saline was administered intravenously. Sufficient inulin-¹⁴C was added to the sustaining solutions to provide counting rates at least 10 times greater than background in 10- μ l samples of plasma.

For experiments in which ECF volume expansion was minimized, the sustaining solutions containing NaHCO₃, NaCl, and inulin were infused at a rate of 0.09 or 0.11 ml/min. Before each increment in the rate of bicarbonate infusion, a single injection of 0.4 or 0.6 mEq of HCO₃ was infused in a volume of 0.3 or 0.4 ml. An equilibration period of at least 12 min was allowed after initiating each new sustaining solution. Exaggerated extracellular fluid volume expansion was accomplished as follows: after obtaining two control clearance periods, isotonic sodium chloride containing appropriate concentrations of inulin-14C was infused at 0.15 ml/min for 20 min, 0.375 ml/min for the next 20 min, and 0.75 ml/min for the following 20 min. The infusion then was continued at 0.46 ml/min and two new control clearance periods were obtained. At this point, a bicarbonate-containing solution was substituted for the sodium chloride and the titration studies were performed using a pattern of increasing bicarbonate concentrations in the infusate similar to that described in the first group of experiments. The infusion rate for these experiments, however, was maintained at 0.46 ml/min instead of 0.1 ml/min.

All urine samples were collected under oil, and blood samples were obtained directly from the indwelling femoral arterial cannula. The pH and pCO₂ determinations were made immediately after collection of blood and urine using an Instrumentation Laboratory, Inc. microgas analyzer (Model IL 113-FL). Bicarbonate concentrations in urine and plasma were calculated using the Henderson-Hasselbalch equation with a pk' value of 6.1 and an α value of 0.0301 for plasma. An α value of 0.0309 was used for urine and pk' values were calculated for each urine sample using the formula pk' = 6.33 $-0.5 \times \sqrt{B}$, where B represents the total cation concentration estimated as the sum of sodium concentration plus



FIGURE 2 Mean bicarbonate titration curves for six animals studied under conditions of exaggerated ECF volume expansion.

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potassium concentration. Inulin-¹⁴C in plasma and urine samples was counted in a Packard Tri-Carb liquid scintillation counter (Model 3214); at least 10,000 counts were obtained in all urine and plasma samples. Bicarbonate reabsorption was calculated as the difference between the amount filtered and the amount excreted. A Donnan factor of 1.05 was employed for estimating the concentration of bicarbonate in the ultrafiltrate. Sodium was determined using an Instrumentation Laboratories flame photometer.

RESULTS

The results of a representative bicarbonate titration study in an animal in which extracellular fluid volume expansion was minimized are shown in Table I. The plasma bicarbonate concentration was increased from an initial level of 24 mEq/liter to a final value of 58 mEq/ liter. The arterial pCO₂ was 40 mm Hg in the control period; it then decreased to 36 mm Hg and rose pro-

 TABLE II

 Representative Bicarbonate Titration Experiment during Exaggerated ECF Volume Expansion

C1	· · · · · · · · · · · · · · · · · · ·			Plasma			Urine					
period	Time	GFR	pН	HCO3	pCO ₂	pH	HCO3	pCO2	HCO3 e	xcretion	HCO3 reab	sorption
	min	ml/min		µEq/ml	mm Hg		µEq/ml	mm Hg	µEq/min	µEq/ml _GFR	µEq/min	µEq/ml GFR
	-187	Light and	ether and position	esthesia fo ning anim	or insertio al in hole	on of tail der (du	l vein and f ration 40 r	emoral a nin)	rtery cann	ulae and	bladder cat	theter,
	- 62	Inulin µci	prime 0 nulin-14C	.7 μc inul C in 100 n	in- ¹⁴ C in 1 nl normal	l ml nor l saline	mal saline	, i.v. Sust	aining sol	ution beg	un contain	ing 71
1	0–23	1.65	7.37	18.8	33.5	5.80	0.30	20.0	0.01	0.01	32.5	19.7
2	23-43	1.99	7.37	19.1	34.0	5.78	0.29	21.0	0.01	0.01	39.8	20.0
	43-63	Norma	l saline o	containing	g inulin-14	С (17 µ	c in 100 m	l normal	saline) at	0.15 ml/	'min	
	63-83	Norma	l saline o	containing	z inulin-14	C (17 µ	c in 100 m	l normal	saline) at	0.375 m	/min	
	83-103	Norma	l saline	containin	g inulin-1	4C (17	uc in 100 n	nl norma	l saline) a	t 0.75 m	/min	
	103-125	Norma	l saline	containin	g inulin-1	•C (17	uc in 100 r	nl norma	l saline) a	t 0.46 m	/min	
3	125-133	2.88	7.34	19.5	37.5	6.00	0.57	25.0	0.32	0.11	58.7	20.3
4	133-143	2.56	7.35	19.8	37.5	6.00	0.55	24.0	0.25	0.10	53.0	20.6
	148-168	Sustain rate	ing solu 0.46 ml/	tion with /min	17 μc in	ulin-14C	/100 ml a	nd NaHO	CO₃ 30 ml	Eq/liter s	solution, in	fusion
5	168-178	2.78	7.42	23.5	37.5	6.12	0.80	26.5	0.32	0.12	68.3	24.6
6	178-189	2.72	7.45	25.6	37.5	6.03	0.70	29.3	0.29	0.11	72.8	26.8
	195–210	Prime NaH	0.3 ml 1 CO ₂ 30	l.5 м Na mEq/lite	HCO₃ (iı r, infusio	n D. W n rate (.) (0.45 m).46 ml/mi	iEq HCC n)₃ [−]), susta	uning sol	ution cont	aining
7	210/218	2.89	7.55	34.7	40.8	6.95	6.58	31.5	2.30	0.70	103.0	35.6
8	218-228	3.05	7.56	35.3	41.0	7.00	8.06	36.0	3.71	1.21	109.3	35.7
	233–248	Sustain 0.46	ing solu ml/min	tion wit	h 17 μc	inulin-14	C/100 ml	and Na	HCO3 40	mEq/lite	er, infustio	n rate
9	248-258	2.99	7.61	39.0	40.5	7.10	9.64	35.0	4.34	1.45	118.1	39.5
10	258-273	2.39	7.62	38.3	39.0	7.24	16.1	41.5	6.29	2.63	89.8	37.6
	277–287	Sustain 0.46	ing solu ml/min	ition with	h 17 μc	inulin-14	C/100 ml	and Na	HCO3 60	mEq/lit	er, infusio	n rate
11	287–297	1.97	7.64	42.3	40.8	7.42	22.4	38.0	6.71	3.39	80.8	40.8
12	297–308	1.68	7.65	43.2	40.8	7.50	23.9	34.5	6.44	3.38	69.8	41.5
	312-322	Prime NaH	0.3 ml 1 CO3 60	l.5 м Na mEq/lite	HCO₃ (in r, infusio	n D. W n rate (′.) (0.45 m).46 ml/mi	nEq HCC .n)₃ [−]), susta	aining sol	lution cont	aining
13	322-334	2.50	7.67	45.8	41.5	7.60	41.5	43.5	14.1	5.65	106.1	42.4
14	334-344	2.25	7.66	46.2	42.5	7.64	43.5	43.5	15.2	6.77	93.9	41.7
	344-357	Prime	0.3 ml 1	1.5 м Na	HCO ₂ (in	n D. W	.) (0.45 m	Eq HCC)₃ [−]), susta	aining sol	lution cont	aining
		NaH	CO ₃ 80	mEq/lite	r, infusio	n rate ().46 ml/mi	n		-		
15	357-375	2.73	7.70	49.0	42.0	7.62	49.5	47.2	16.3	5.98	124.1	45.5
16	375-387	1.96	7.69	51.5	44.5	7.65	57.3	53.5	18.9	9.65	87.1	44.4
	392-402	Prime NaH	0.3 ml ICO3 80	1.5 м Na mEq/lite	HCO3 (in r, infusion	n D. W n rate 0.	7.) (0.45 m 46 ml/min	nEq HCC	O₃ [−]), susta	aining so	lution cont	taining
17	402-413	1.41	7.69	54.5	46.0	7.71	69.2	55.0	20.8	14.7	59.9	42.5
18	413-426	1.92	7.68	52.5	46.0	7.80	83.7	54.0	29.3	15.3	76.6	39.9

Rat weight, 190 g;

		Plasma					
	CIn	HCO3	pH	pCO ₂			
	ml/min	µEq/ml		mm Hg			
Control							
"Minimal" expansion $(n = 9)$	2.78 ± 0.19	19.7 ± 1.96	7.38 ± 0.03	33.5 ± 1.81			
"Exaggerated" expansion $(n = 6)$							
Before expansion	2.27 ± 0.14	20.4 ± 2.04	7.36 ± 0.03	36.6 ± 2.46			
After expansion	2.87 ± 0.14	19.6 ± 1.64	7.34 ± 0.03	37.2 ± 1.37			
Bicarbonate diuresis							
"Minimal" expansion	2.47 ± 0.21	46.1 ± 1.86	7.66 ± 0.01	42.1 ± 1.04			
"Exaggerated" expansion	2.73 ± 0.03	47.5 ± 2.84	7.67 ± 0.02	42.9 ± 1.23			

Values represent means \pm SE of means. Values during control conditions were selected from a compilation of one or more control clearance periods from each rat under the specified conditions. Values during bicarbonate diuresis were derived by selecting clearance periods in which plasma bicarbonate concentrations in the minimal and exaggerated expansion groups were comparable and then recording the values for plasma pH and pCO₂ urine pH, pCO₂, bicarbonate excretion, and sodium excretion.

gressively thereafter to a final value of 50. Bicarbonate reabsorption increased over the entire range of plasma bicarbonate concentrations and no Tm or tendency for a Tm was observed even at the highest plasma level achieved. Thus with a plasma bicarbonate concentration of 57.5 mEq/liter, bicarbonate reabsorption was 57.1 μ Eq/ml GFR. Bicarbonate excretion increased gradually with the increments in the plasma bicarbonate concentrations but the highest rate of excretion was 13.7 μ Eq/min with a filtered load of 134 μ Eq/min.

Fig. 1 depicts a composite titration curve for nine animals in which ECF volume expansion was minimized. Each point is the mean of from 2 to 25 individual observations. Consistent with the result shown in Table I, mean bicarbonate reabsorption increased progressively over a range of plasma bicarbonate concentrations extending from 12 to 66 mEq/liter. No Tm was demonstrable for the group data despite multiple observations at plasma levels over 45 mEq/liter. Bicarbonate excretion did not begin until plasma bicarbonate concentrations exceeded 27 mEq/liter.

A representative titration study in which extracellular fluid volume expansion was exaggerated is shown in Table II. Plasma bicarbonate concentrations increased from 19 to 54.5 mEq/liter. pCO₂ values rose from 33.5 to 46 mm/Hg. In contrast to the pattern presented in Table I and pictured in Fig. 1, a clearly discernible tendency towards stabilization of bicarbonate reabsorption is evident with a Tm value between 40 and 45 μ Eq/ml GFR. In the final portion of the experiment, bicarbonate excretion approximated 15 μ Eq/ml GFR in contrast to the maximal value of 5.3 μ Eq/ml GFR shown in the representative experiment in Table I for minimized expansion. A composite titration curve for six rats studied during exaggerated extracellular fluid volume expansion is shown in Fig. 2. The pattern for the animals



FIGURE 3 Bicarbonate excretion for animals with minimized and exaggerated ECF volume expansion.

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Urine								
pH pCO2		UHCO2V	UHCO3V GFR	Na	Filtered Na ⁺ excreted			
	mm Hg	µEq/min		µEq/min	%			
6.31 ± 0.17	22.3 ±1.61	0.11 ± 0.49	0.04 ± 0.05	8.0 ± 2.32	1.92 ± 0.49			
6.28 ± 0.24 6.23 ± 0.12	21.8 ± 1.70 25.4 ± 1.44	0.15 ± 0.08 0.56 ± 0.14	0.06 ± 0.03 0.20 ± 0.05	10.4 ± 2.10 71.3 ±4.65	3.37 ± 0.75 17.82 ± 1.44			
8.01 ± 0.04 7.69 ±0.05	50.9 ± 5.64 55.5 + 3.79	$\begin{array}{r} 11.34 \ \pm 1.68 \\ 27.70 \ + 3.97 \end{array}$	4.75 ± 0.76 10.19 ±1.56	19.7 ± 2.43 78.2 ± 7.41	6.32 ± 0.90 20.76 ± 2.58			

with minimized expansion is superimposed for comparison. In the animals with exaggerated expansion, reabsorption tended to stabilize above a plasma bicarbonate concentration of 45 μ Eq/ml and there is an apparent Tm for bicarbonate reabsorption with a value of approximately 46 μ Eq/ml GFR. Bicarbonate excretion began earlier in the animals with exaggerated expansion than in those with minimized expansion and slope of the excretion curve is much steeper at higher plasma bicarbonate levels.

In Fig. 3, urinary excretion of bicarbonate at increasing plasma bicarbonate concentrations is compared in the animals with minimized and exaggerated expansion of ECF volume. Bicarbonate excretion began at a lower plasma level in the more expanded group and at all levels of plasma bicarbonate above the respective thresholds, excretion rates were greater in animals with exaggerated expansion than in those with minimized expansion. Table III presents comparative data for both groups of animals for plasma pH, pCO₂, fractional sodium excretion, and certain other relevant parameters during the control periods, and after brisk bicarbonate excretion was in effect. At the same plasma bicarbonate concentration, bicarbonate excretion per unit of GFR was approximately twice as great in the animals with exaggerated expansion. However no differences in either pH or pCO₂ were evident between the two groups. The patterns of sodium excretion, on the other hand, were markedly different with the exaggerated expansion group excreting 21% of the filtered sodium while the minimal expansion group excreted only 6.0%. The absolute rates of sodium excretion (in #Eq/min) were 78.2 and 19.7 respectively; only about 25% of this difference could be attributed to bicarbonate as an impermeant anion.

DISCUSSION

The standard procedure for examining bicarbonate reabsorption consists of infusing bicarbonate solutions so as to effect a gradual but progressive increment in plasma bicarbonate concentrations. Since bicarbonate is infused as a sodium salt, large quantities of sodium are administered during the course of a classical titration experiment. Hence, expansion of the extracellular fluid volume is an inescapable consequence of the experimental method. There is now compelling evidence that ECF volume expansion leads to striking alterations of proximal tubular functions. The best characterized of these is the inhibition of fractional sodium reabsorption (4, 5). However glucose reabsorption is altered (6) and in the dog, maximum tubular absorption rate for p-aminohippuric acid (TmPAH) is diminished.¹ There are also observations that suggest that calcium (7), magnesium (7), and urate reabsorption (8) may be influenced by ECF expansion. The reabsorption of bicarbonate in the proximal tubule presumably is coupled to sodium reabsorption either directly or indirectly whether this reabsorption occurs in consequence of the secretion of hydrogen ions into the tubular lumen or the transport of bicarbonate as an ion. Thus the possibility exists that the pattern of bicarbonate reabsorption which has been accepted as normal may in fact represent an altered pattern which conceals substantial inhibition of bicarbonate reabsorptive capacity. In micropuncture studies, recently described by Kunau, Frick, Rector, and Seldin (9) estimated tubular fluid/plasma ratios for bicarbonate (estimated from tubular fluid/plasma chloride ratios) in su-

¹ Shapiro, H., M. Lao, C. Manley, R. G. Schultze, and N. S. Bricker. Unpublished observations.

perficial nephrons of rats were substantially higher during saline loading than in the hydropenic state. These data indicate that ECF volume does inhibit bicarbonate reabsorption in the proximal tubule. This inhibition could affect not only the apparent Tm for bicarbonate but also the threshold level at which bicarbonate first appears in the urine.

The present studies support the foregoing possibility. When ECF volume expansion was minimized during the execution of titration experiments, no Tm was demonstrated in the majority of individual rats studied or in a group plot from nine experiments despite the fact that plasma bicarbonate concentrations were elevated to values in excess of 60 mEq/liter. Conversely, when extracellular fluid volume expansion was exaggerated, a clear Tm could be demonstrated and the threshold for bicarbonate excretion was diminished. The data, therefore, suggest that the titration procedure as typically employed leads to inhibition of proximal tubular reabsorption of bicarbonate; thus the apparent Tm observed in such conditions is not a true index of the maximum capacity for proximal tubular reabsorption of bicarbonate. Why the values for bicarbonate reabsorption were so high even in the expanded rats is not evident, but this presumably relates to a species difference between the rat and other species previously studied (in man and dogs).

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