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# ELECTROPHORETIC STUDIES OF THE VITAMIN B<sub>12</sub>-BINDING PROTEIN OF NORMAL AND CHRONIC MYELOGENOUS LEUKEMIA SERUM \*

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The serum in chronic myelogenous leukemia (CML) has an increased concentration of vitamin B<sub>12</sub> (1, 2) and an increased capacity to bind added vitamin B<sub>12</sub> either *in vivo* (3-5) or *in vitro* (2, 6, 7). The vitamin B<sub>12</sub> of normal and CML serum moves with the  $\alpha$ -globulins when serum is electrophoresed at pH 8.6 (1, 8, 9). The seromuroid, the fraction of serum containing glycoproteins which comprises about 1.5 per cent of the serum proteins (10), moves with the  $\alpha$ -globulins when electrophoresed at pH 8.6 (11). Furthermore, it has been found that the seromuroid fraction isolated from CML serum is capable of binding amounts of added cobalt<sup>60</sup>-labeled vitamin B<sub>12</sub> (B<sub>12</sub>\*) sufficient to account for the increased binding capacity of the whole serum (12). The possibility that the binding protein of both normal and CML serum for native vitamin B<sub>12</sub> (B<sub>12</sub>BP) is in the seromuroid fraction was investigated.

## METHODS

Venous blood was collected in the fasting state from normal subjects and from patients with CML in hematological and clinical relapse. About one to two hours after clotting, the serum was separated and either used immediately or stored at -20°C. The seromuroid fraction of serum was prepared by the method of Winzler, Devor, Mehl and Smyth (10) as follows: one volume of serum was added to one volume of 0.4 M sulfosalicylic acid, thoroughly mixed for five minutes, and then centrifuged. The supernatant containing the seromuroid was separated and neutralized with 1 N NaOH. The neutralized seromuroid and the original serum from which it had been prepared were dialyzed against large volumes of 0.15 M NaCl for 72 hours at 4 to 6°C. and its vitamin B<sub>12</sub> concentration then determined by *Euglena gracilis* assay (13). When used for electrophoresis, the seromuroid was dialyzed for 72 hours against large volumes of distilled water, lyophilized, and the dried

protein then dissolved in the appropriate buffer. Seromuroid or serum containing bound (nondialyzable) B<sub>12</sub>\*<sup>1</sup> was prepared by adding an amount of B<sub>12</sub>\* in excess of the binding capacity (4 m $\mu$ g. B<sub>12</sub>\* per ml. of normal serum or its equivalent seromuroid and 12 m $\mu$ g. per ml. of CML serum or its equivalent seromuroid). The seromuroid was then dialyzed against large volumes of distilled water for 72 hours at 4 to 6°C. and lyophilized, while the serum was dialyzed against 0.15 M NaCl.

Lyophilized Cohn plasma fractions (I through VI),<sup>2</sup> prepared by Method VI (14), were dissolved in sufficient 0.15 M NaHCO<sub>3</sub> to bring them to their equivalent plasma volumes, dialyzed for 48 hours against 0.15 M NaCl, and the vitamin B<sub>12</sub> concentration determined as was the vitamin B<sub>12</sub> concentration of the original plasma.

Zone electrophoretic separation of serum or lyophilized seromuroid was performed using starch blocks as the supporting medium. The general technique described by Kunkel (15) was used except that the electrophoresis was carried out at pH 4.5 which permitted separation of the B<sub>12</sub>BP from the bulk of serum proteins. The starch blocks were prepared from slurries of starch<sup>3</sup> that had been previously washed once with distilled water and twice with an acetate-sodium chloride buffer pH 4.5, ionic strength 0.10. The buffer consisted of 0.04 M sodium acetate and 0.06 M NaCl with concentrated HCl added to bring the pH to 4.5. The following samples were electrophoresed: a) a mixture of 1.0 ml. of serum and 0.5 ml. of acetate buffer; b) serum containing bound (nondialyzable) B<sub>12</sub>\* in a volume of 1.5 to 2 ml.; c) lyophilized seromuroid dissolved in a solution of 1 ml. of 0.15 M NaCl and 0.5 ml. acetate buffer; and d) lyophilized seromuroid containing bound B<sub>12</sub>\* dissolved as in c. Samples were applied to a 4 to 5 mm. transverse slit at the center of the block. In some experiments, serum containing bound B<sub>12</sub>\* was run in parallel on the same starch block as the native serum. Electrophoresis was carried out for 16 hours at 5 to 8°C. using a current flow of 36 ma. for a 31 cm. long, 10 cm. wide, 1.5 cm. thick starch block. At the termination of electrophoresis a maximum

<sup>1</sup> Kindly supplied by Dr. N. S. Ritter, Merck and Co., Rahway, N. J.

<sup>2</sup> Kindly prepared by the Plasma Protein Foundation, Boston, Mass.

<sup>3</sup> Fisher hydrolyzed potato starch, Fisher Scientific Co., New York, N. Y.

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TABLE I  
The native vitamin B<sub>12</sub> concentration of the seromuroid of normal and chronic myelogenous leukemic (CML) sera

Subject		Serum B <sub>12</sub> concentration	Seromuroid B <sub>12</sub> con- centration	Seromuroid fraction
		μg./ml.	μg./ml.	%
Normal				
1	S.W.	525	435	83
2	L.S.	125	122	98
3	A.N.	732	451	62
4	M.B.	732	539	74
5	T.C.	681	544	80
6	S.J.	332	208	63
	Mean	521	383	77
	Range	125-732	123-544	62-98
CML				
1	H.W.	6,096	5,976	98
2	A.M.	15,121	15,101	100
3	C.M.	9,220	9,010	98

pH change of 0.2 pH unit was found at either end of the gel. After drying at room temperature (0.5 to one hour) the gel was cut into transverse segments so that the origin was included in one segment (O segment). Other segments were designated as anodal (A) or cathodal (C) and identified by the number of segments from the O segment. Sections of the block at the cathodal and anodal ends well outside the range of migration of the proteins were removed to serve as blanks. Each segment was suspended in 8 ml. of distilled water, the contents periodically mixed for 1.5 hours, and then centrifuged. The supernatant was removed and centrifuged again. The vitamin B<sub>12</sub> concentration of these eluates was then determined by *E. gracilis* assay. The per cent recovery of vitamin B<sub>12</sub> from the starch block in 19 electrophoretic analyses of serum or seromuroid was equal to an average of 93 per cent ± 14, range, 75 to 115 per cent.

The protein concentration of starch eluates was determined by Kunkel and Tiselius' modification (16) of the Folin-Ciocalteu procedure (17). Protein concentration was plotted in terms of optical density with necessary corrections made for the volume of eluate analyzed. Sialic acid was determined by the method of Werner and Odin (18) using 2 ml. of the eluates. Because of the negligible reading of the blank with Baker's iso-amyl alcohol, further purification of the iso-amyl alcohol was not necessary.

In preliminary experiments, it was found that albumin and the bulk of the globulins remained at the origin or migrated cathodally when serum was electrophoresed at pH 4.5. Acidic α<sub>1</sub>-glycoprotein,<sup>4</sup> isolated from Cohn Fraction VI (19), was dissolved in a solution composed of 1 ml. 0.15 M NaCl and 0.5 ml. acetate buffer, electrophoresed on a starch block (pH 4.5) and the protein

<sup>4</sup> Kindly supplied by Dr. Karl Schmid, Massachusetts General Hospital, Boston, Mass.

and sialic acid concentration of 1.0 cm. segments of the block determined.

After electrophoresis, the starch segments containing nondialyzable B<sub>12</sub>\* were placed in test tubes and the radioactivity determined in a plastic scintillation well counter (20) having a background of 385 to 400 counts per minute. Samples with higher counting rates, such as a) the origin and first four cathodal segments of normal sera, b) anodal segments of the seromuroid and c) the anodal and cathodal segments of CML sera, were counted long enough to give a 10 to 15 per cent counting error. The anodal segments of the normal sera contained small amounts of radioactivity (as little as five to 10 counts per minute over background) and were counted for three to four 10 minute periods with 10 minute counts of background alternating between each sample count. The radioactivity of the B<sub>12</sub>\* solution added to serum was counted in the same well counter using similar geometry, and the concentration of vitamin B<sub>12</sub> in this solution was also determined. From the a) specific activity of the added B<sub>12</sub>\* and b) radioactivity found in each starch segment, the amount of B<sub>12</sub>\* (μg.) per segment was estimated. The total vitamin B<sub>12</sub> bound by the proteins in any starch segment was calculated by summing of the native vitamin B<sub>12</sub> (determined by *E. gracilis* assay) and of the nondialyzable B<sub>12</sub>\*.

## RESULTS

### Vitamin B<sub>12</sub> concentration of the seromuroid

The seromuroid contained an average of 77 per cent, range 62 to 98 per cent, of the vitamin B<sub>12</sub> of whole normal serum (Table I). Despite a great increase in serum vitamin B<sub>12</sub> concentration, 98 to 100 per cent of the vitamin was recovered from the seromuroid in three patients with CML.

### Vitamin B<sub>12</sub> concentration of Cohn plasma fractions

Vitamin B<sub>12</sub> was found in all six Cohn plasma fractions. Fraction V contained the highest percentage of the plasma vitamin (27 per cent) while Fraction VI had only 14 per cent.

### Vitamin B<sub>12</sub> concentration of fractions of the seromuroid separated by electrophoresis

The distribution of native vitamin B<sub>12</sub>, protein and sialic acid in the seromuroid, obtained from normal serum following starch gel electrophoresis at pH 4.5, is shown in Figure 1. The protein and sialic acid were concentrated in segments anodal to the origin with peak levels found in the A-3 and A-4 segments. A similar distribution of protein and sialic acid was found when a crystalline preparation of the acidic α<sub>1</sub>-glycoprotein was electro-

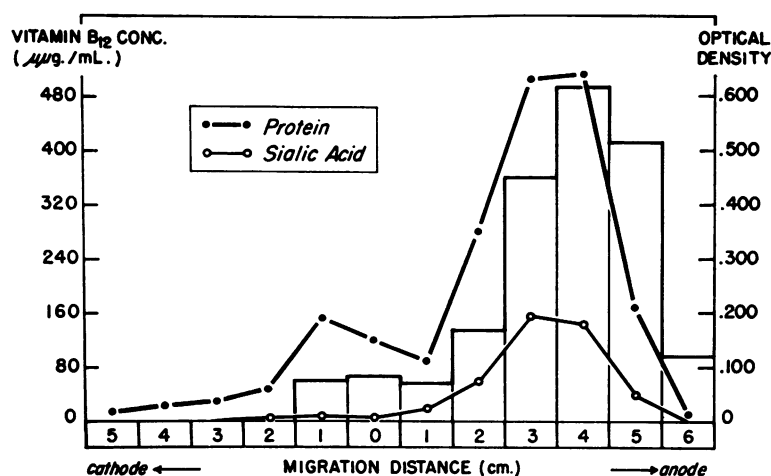


FIG. 1. NATIVE VITAMIN B<sub>12</sub> CONCENTRATION OF FRACTIONS OF NORMAL SEROMUCOID SEPARATED BY ELECTROPHORESIS ON STARCH GEL AT pH 4.5

Vitamin B<sub>12</sub> concentration is plotted on the left ordinate and is represented by the open boxes. Optical density is plotted on the right ordinate with protein the solid dots ●—●, and sialic acid the open dots ○—○.

phoresed at pH 4.5. The vitamin B<sub>12</sub> was also found anodally with peak concentrations at A-3 to A-5 segments. The B<sub>12</sub>\* bound to the normal seromucoid had a distribution similar to the native vitamin, with peak concentrations at A-2 to A-4 segments (Figure 2).

The distribution of protein, sialic acid and vitamin B<sub>12</sub> in the CML seromucoid was similar to that of the normal, with peak concentrations found at A-3 to A-4 segments (Figure 3). Furthermore, the distribution of added B<sub>12</sub>\* bound by

CML seromucoid was similar to that of the native vitamin, with peak concentrations located at A-2 to A-4 segments (Figure 4).

#### *Vitamin B<sub>12</sub> concentration of fractions of serum separated by electrophoresis*

Electrophoresis of a normal serum at pH 4.5 (Figure 5) is representative of the findings in a group of eight such sera. In sharp contrast to the seromucoid, there was a negligible amount of protein in A-2 to A-6 segments with the peak pro-

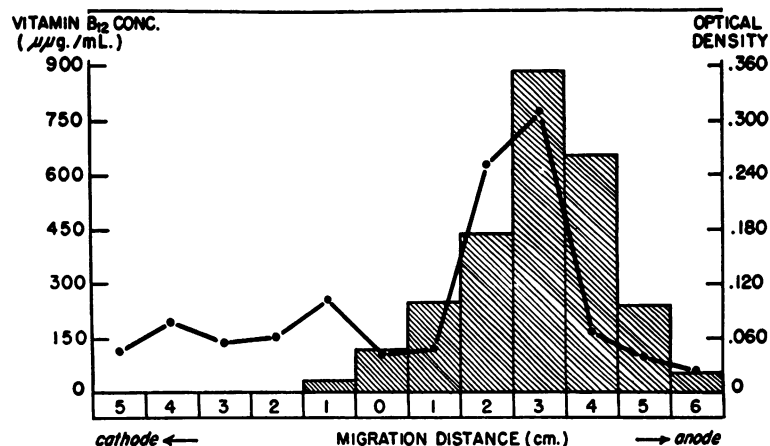


FIG. 2. DISTRIBUTION OF BOUND (NONDIALYZABLE) B<sub>12</sub>\* AMONG FRACTIONS OF NORMAL SEROMUCOID SEPARATED BY STARCH GEL ELECTROPHORESIS AT pH 4.5

The concentration of B<sub>12</sub>\* is plotted on the left ordinate and is represented by the ruled boxes. Optical density of protein is plotted on the right ordinate and is represented by the solid dots ●—●.

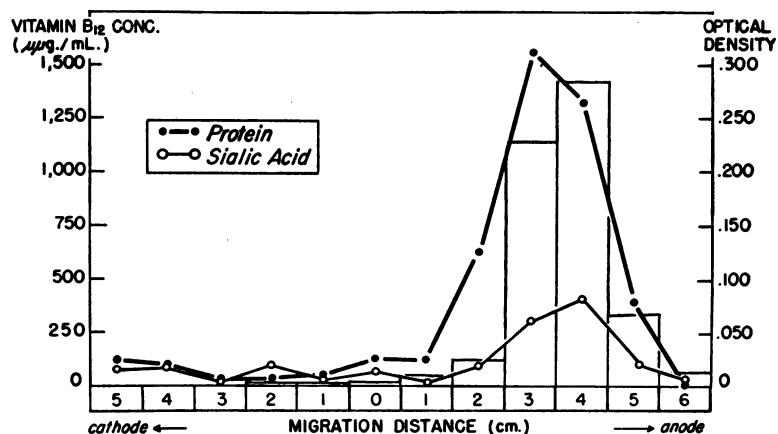


FIG. 3. NATIVE VITAMIN B<sub>12</sub> CONCENTRATION OF FRACTIONS OF CHRONIC MYELOGENOUS LEUKEMIA SEROMUCOID SEPARATED BY ELECTROPHORESIS ON STARCH GEL AT PH 4.5

Vitamin B<sub>12</sub> concentration plotted on the left ordinate is represented by the open boxes. Optical density is plotted on the right ordinate with protein the solid dots ●—● and sialic acid the open dots ○—○.

tein concentration at the O segment (A-1 or C-1 in some sera). However, virtually all of the native vitamin B<sub>12</sub> was recovered anodally (83 to 92 per cent found in segments A-2 to A-6), with peak concentrations in A-3 to A-5 segments. The distribution of the native vitamin B<sub>12</sub> and of the nondialyzable B<sub>12</sub>\* bound to the same serum is shown in Figure 6 and is representative of the findings in a group of seven such sera. The native vitamin had the same anodal distribution as previously described. Only a small amount of

B<sub>12</sub>\* was found anodally, segments A-2 to A-6 containing an average of 80 µµg. per ml. ± 17, range, 28 to 183 µµg. per ml. The proteins with the greatest binding ability for the added B<sub>12</sub>\* were found at the origin and C-1 to C-3 segments. The binding of B<sub>12</sub>\* by A-2 to A-6 segments was equal to an average of 6 per cent ± 4, range, 3 to 15 per cent of the total bound radiovitamin. The saturation of the B<sub>12</sub>BP (located at A-2 to A-6 segments) as estimated from the ratio of the native bound vitamin to the total vitamin that could be

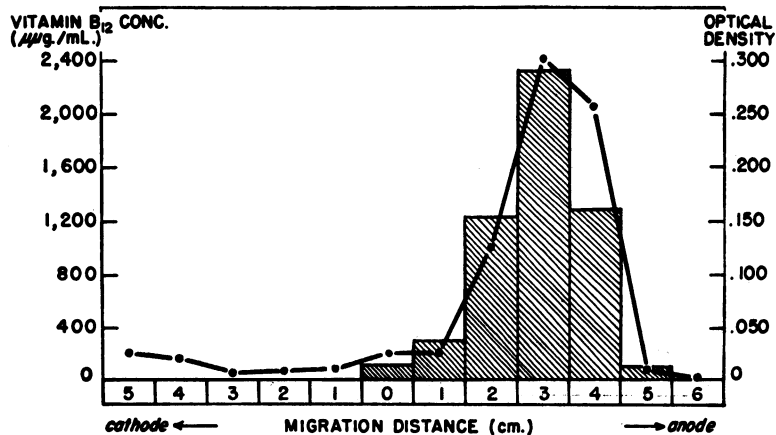


FIG. 4. DISTRIBUTION OF BOUND (NONDIALYZABLE) B<sub>12</sub>\* AMONG FRACTIONS OF CHRONIC MYELOGENOUS LEUKEMIA SEROMUCOID SEPARATED BY STARCH GEL ELECTROPHORESIS AT PH 4.5

The concentration of B<sub>12</sub>\* plotted on the left ordinate is represented by the ruled boxes. Optical density of protein is plotted on the right ordinate and is represented by the solid dots ●—●.

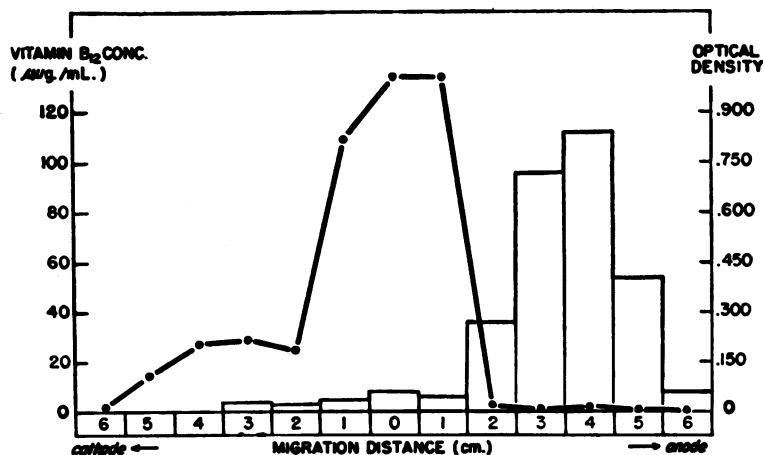


FIG. 5. NATIVE VITAMIN B<sub>12</sub> CONCENTRATION OF PROTEIN FRACTIONS OF NORMAL SERUM SEPARATED BY STARCH GEL ELECTROPHORESIS AT PH 4.5

Vitamin B<sub>12</sub> concentration is plotted on the left ordinate and is represented by the open boxes. Optical density of protein is plotted on the right ordinate and is represented by the solid dots ●—●.

bound and was equal to an average of 78 per cent  $\pm$  13, range, 57 to 94 per cent.

The distribution of vitamin B<sub>12</sub> in a CML serum (Figure 7) is representative of a group of four such sera. As in the normal, the protein concentration of the A-2 to A-6 segments was

negligible, while the native vitamin B<sub>12</sub> was virtually all found anodally with peak concentrations at A-3 to A-5 segments. The distribution of the native vitamin and of the nondialyzable B<sub>12</sub>\* (Figure 8) is representative of a group of three such sera. The native vitamin had the same anodal

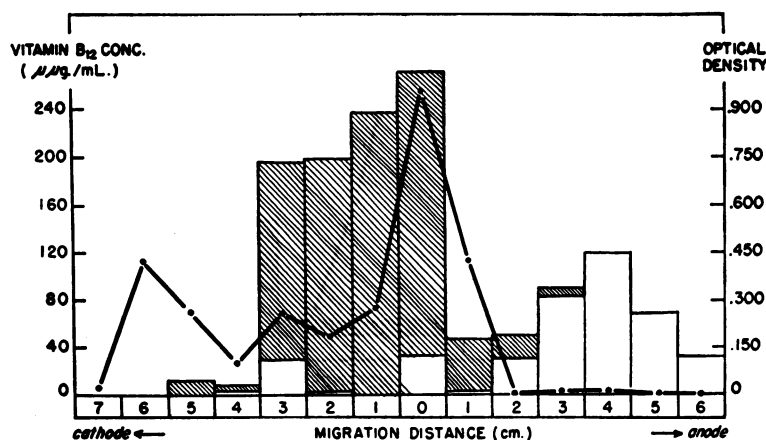


FIG. 6. DISTRIBUTION OF NATIVE VITAMIN B<sub>12</sub> AND OF BOUND (NON-DIALYZABLE) B<sub>12</sub>\* AMONG THE PROTEINS OF NORMAL SERUM SEPARATED BY ELECTROPHORESIS ON STARCH GEL AT PH 4.5

Normal serum and an aliquot of the same serum containing bound (nondialyzable) B<sub>12</sub>\* were electrophoresed on the same starch gel. The native vitamin shown by the open boxes was measured by *Euglena gracilis* assay while the bound radiovitamin, shown by the ruled boxes, was measured by counting the radioactivity in each segment. The solid dots ●—● refer to protein concentration.

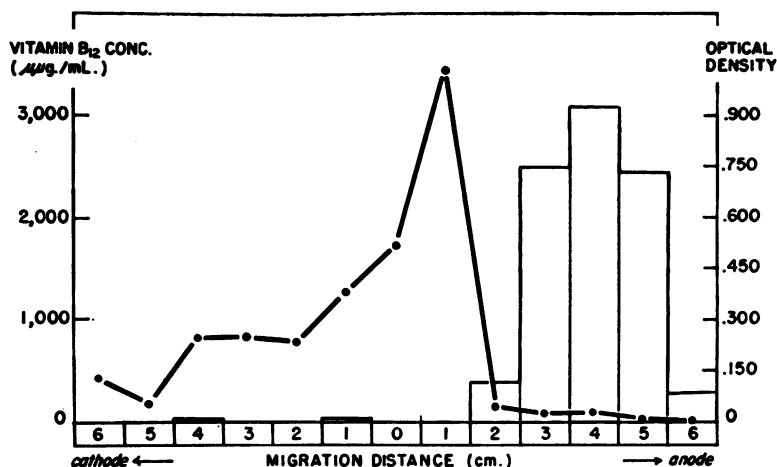


FIG. 7. NATIVE VITAMIN B<sub>12</sub> CONCENTRATION OF PROTEIN FRACTIONS OF CHRONIC MYELOGENOUS LEUKEMIC SERUM SEPARATED BY STARCH GEL ELECTROPHORESIS AT PH 4.5

Vitamin B<sub>12</sub> concentration is plotted on the left ordinate and is represented by the open boxes. Optical density of protein is plotted on the right ordinate and is represented by the solid dots ●—●.

distribution as described above. However, in contrast to the normal, a large amount of the added B<sub>12</sub>\* (2,200 to 3,973 µg. per ml.) was bound by the A-2 to A-6 segments. The binding of the radiovitamin by these anodal segments was

equal to 48 to 78 per cent of the total serum binding of B<sub>12</sub>\*. The per cent saturation of the B<sub>12</sub>BP located at A-2 to A-6 segments ranged from 35 to 75 per cent, as calculated by the method outlined above.

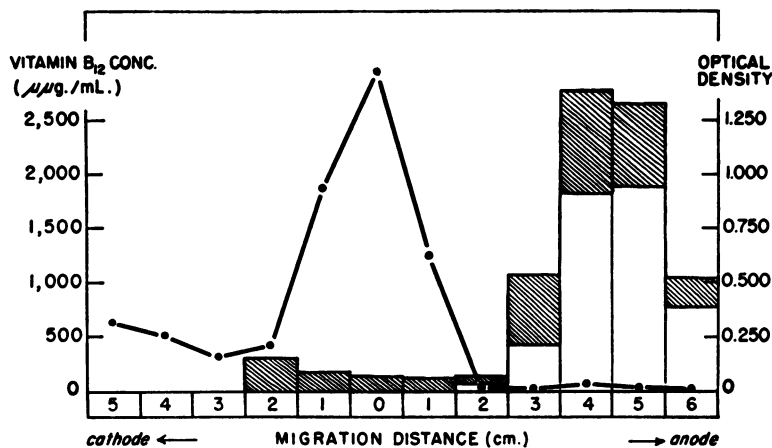


FIG. 8. DISTRIBUTION OF NATIVE VITAMIN B<sub>12</sub> AND OF BOUND (NON-DIALYZABLE) B<sub>12</sub>\* AMONG THE PROTEINS OF CHRONIC MYELOGENOUS LEUKEMIC (CML) SERUM SEPARATED BY ELECTROPHORESIS ON STARCH GEL AT PH 4.5

CML serum and an aliquot of the same serum containing bound (non-dialyzable) B<sub>12</sub>\* were electrophoresed on the same starch block. The native vitamin shown by the open boxes was measured by *Euglena gracilis* assay while the bound radiovitamin shown by the ruled boxes was measured by counting the radioactivity in each segment. Note the change in scale of vitamin B<sub>12</sub> concentration plotted on the left ordinate from that of Figure 6. The solid dots ●—● refer to protein concentration.

## DISCUSSION

A large fraction of the native vitamin B<sub>12</sub> of normal serum was found in the seromuroid. Cohn Fraction VI, thought to be similar to the seromuroid in composition (19), contained little of the vitamin. In Method VI of Cohn and associates, the major protein fractions of plasma are isolated by a series of precipitations which depend on changes in pH, ionic strength and ethanol concentration (14). Some coprecipitation of the seromuroid routinely occurs when it is prepared from serum by precipitation of the serum proteins with sulfosalicylic or perchloric acid (10), and similarly it may be presumed that coprecipitation of the B<sub>12</sub>BP with each Cohn plasma fraction may account for the absence of selective concentration of the vitamin in Fraction VI.

Since the great bulk of serum proteins have isoelectric points of 4.5 or higher, they either remained at the origin or migrated cathodally at this pH. However, due to the more acidic isoelectric point of the B<sub>12</sub>BP, it migrated anodally with the other proteins of the seromuroid. The native vitamin B<sub>12</sub> in serum or seromuroid preparations was usually distributed over a relatively wide anodal area of the starch block, with peak concentrations found in a 2 to 3 cm. width. Trailing of the protein and/or diffusion of the B<sub>12</sub>BP during the electrophoretic run may account for the wide distribution of the vitamin and does not necessarily indicate the presence of more than one binding protein. The anodal distribution of vitamin B<sub>12</sub> was similar to, but not identical with, that of the main protein and sialic acid peaks of the seromuroid or its chief constituent, the acidic  $\alpha_1$ -glycoprotein (19, 21). Despite their somewhat similar mobility, the B<sub>12</sub>BP is probably not the acidic  $\alpha_1$ -glycoprotein, since 1) the B<sub>12</sub>BP was not selectively precipitated with Cohn Fraction VI, whereas the acidic  $\alpha_1$ -glycoprotein has been found only in this fraction (19); and 2) Fahey, McCoy and Goulian fractionated serum on a diethylaminoethyl cellulose ion-exchange column and found that the B<sub>12</sub>BP was eluted off before the acidic  $\alpha_1$ -glycoprotein (22). A small amount of the native vitamin B<sub>12</sub> of normal serum (10 per cent) was found scattered among cathodally moving proteins. It is impossible to know whether this

represents technical error in the measuring of small amounts of the vitamin or another B<sub>12</sub>BP.

The B<sub>12</sub>BP of CML serum had properties similar to the normal B<sub>12</sub>BP since 1) it was found in the seromuroid, and 2) had anodal mobility when electrophoresed on starch gel at pH 4.5. Mendelsohn, Watkin, Horbett and Fahey found that both the normal and CML B<sub>12</sub>BP behaved similarly when serum was chromatographed on a diethylaminoethyl cellulose ion-exchange column (23). These chemical similarities between the normal and the CML B<sub>12</sub>BP strongly suggest that the increased concentrations of vitamin B<sub>12</sub> uniformly found in CML serum in relapse is due to an increase in concentration of the normal B<sub>12</sub>BP rather than to an abnormal protein. Despite the tremendous increases in concentration of the B<sub>12</sub>BP in CML sera (amounting to as much as 100-fold in one patient we have studied), only slight increases in the concentration of the total seromuroid fraction have been reported in this disease (24). However, the concentration of B<sub>12</sub>BP in normal serum is probably minute as compared to that of the total seromuroid. Thus, the B<sub>12</sub>BP of a typical normal serum, when completely saturated, can bind approximately 0.65  $\mu$ g. per ml. If the molecular weight of the B<sub>12</sub>BP is similar to that of the orosomuroid, *i.e.*, 44,000 (25), and if one molecule of the B<sub>12</sub>BP can bind one molecule of vitamin B<sub>12</sub> (molecular weight, 1,343), the concentration of normal B<sub>12</sub>BP would be equal to 2.20  $\mu$ g. per 100 ml. serum. Even a 100-fold increase in concentration of B<sub>12</sub>BP, to 0.22 mg. per 100 ml., would represent only an increase of 0.2 per cent over the normal concentration of the seromuroid, an increase well within the error of the chemical determination. It would appear that the elevations in seromuroid concentration found in CML are contributed by constituents of the seromuroid other than the B<sub>12</sub>BP.

When additional B<sub>12</sub>\* was added to normal serum, the bound (nondialyzable) radiovitamin was almost all associated with proteins found at the origin or cathodal segments, whereas a very small amount of the added B<sub>12</sub>\* was bound by the anodally migrating B<sub>12</sub>BP. This suggests that at the levels of vitamin B<sub>12</sub> found in normal serum, the B<sub>12</sub>BP approaches saturation and, therefore, the addition of further vitamin results



in the binding of the vitamin to nonacidic proteins. In CML serum on the other hand, there appears to be a commensurate increase of the B<sub>12</sub>BP in association with the increased level of vitamin B<sub>12</sub>, so that although the relative saturation of the former remains approximately normal, the absolute binding capacity for added vitamin B<sub>12</sub> is greatly increased.

The B<sub>12</sub>BP has chemical properties similar to those of the acidic  $\alpha_1$ -glycoprotein and the  $\alpha_2$ -glycoprotein isolated from Cohn Fraction VI (19, 26). Thus, they are all relatively heat resistant, are all soluble in sulfosalicylic or perchloric acid, are all precipitated by phosphotungstate in 2 N HCl and all have acidic isoelectric points as compared with other serum proteins (11, 12, 19, 21, 26, 27). The acidic  $\alpha_1$ -glycoprotein (19) and the  $\alpha_2$ -glycoprotein (26) are mucoid glycoproteins, *i.e.*, proteins containing significant amounts of firmly bound hexosamine (28). This would suggest that the B<sub>12</sub>BP may be a mucoid glycoprotein, although direct confirmation of such a structure must await its isolation in pure form. It is of interest that intrinsic factor (29, 30) and erythropoietin (31) are probably also mucoid glycoproteins. The available evidence suggests that the B<sub>12</sub>BP is not, however, identical with either of these substances. Thus, intrinsic factor has a differing electrophoretic mobility at pH 8.6 (32) and its binding sites for vitamin B<sub>12</sub> are relatively heat labile (33). Recently, we have found a vitamin B<sub>12</sub>-binding substance in CML urine which resembles that found in CML serum (34). This urinary material, when given orally to patients with pernicious anemia in amounts sufficient to bind an oral dose of B<sub>12</sub>\*, had no intrinsic factor activity. Also, increased erythropoietin levels have been reported in such diseases as chronic lymphatic leukemia (35), carcinoma of the cervix (35), secondary polycythemia (36) and hypoplastic anemias (37), whereas vitamin B<sub>12</sub> levels are not particularly increased in these conditions.

It seems reasonable to suppose that the B<sub>12</sub>BP functions as a transport protein for vitamin B<sub>12</sub>. The seromuroid fraction, representing a readily accessible, easily prepared, and easily concentrated source of the B<sub>12</sub>BP, should aid in experiments designed to elucidate the role played by this protein in vitamin B<sub>12</sub> metabolism.

#### SUMMARY

1. A method for the separation of the vitamin B<sub>12</sub>-binding protein (B<sub>12</sub>BP) of normal and chronic myelogenous leukemia serum by electrophoresis on starch gel at pH 4.5 has been described.

2. The B<sub>12</sub>BP of normal serum has been identified as a constituent of the seromuroid fraction of serum. It is a protein with an electrophoretic mobility at pH 4.5 similar to that of the acidic  $\alpha_1$ -glycoprotein, the most acidic protein found in serum. However, it was not selectively precipitated with Cohn Fraction VI.

3. The B<sub>12</sub>BP of chronic myelogenous leukemia serum had properties similar to the normal B<sub>12</sub>BP, suggesting that the great increase in vitamin B<sub>12</sub> levels found in chronic myelogenous leukemia sera results from an increased concentration of the normal B<sub>12</sub>BP rather than from an abnormal protein.

4. Only a very small fraction of the cobalt<sup>60</sup>-labeled vitamin B<sub>12</sub> bound by normal serum as determined by dialysis was bound to the B<sub>12</sub>BP. In contrast to the normal, the largest fraction of the cobalt<sup>60</sup>-labeled vitamin B<sub>12</sub> bound by chronic myelogenous leukemia serum was bound by the B<sub>12</sub>BP.

#### REFERENCES

1. Beard, M. F., Pitney, W. R., and Sanneman, E. H. Serum concentrations of vitamin B<sub>12</sub> in patients suffering from leukemia. *Blood* 1954, 9, 789.
2. Mollin, D. L., and Ross, G. I. M. Serum vitamin B<sub>12</sub> concentrations in leukaemia and in some other haematological conditions. *Brit. J. Haemat.* 1955, 1, 155.
3. Mollin, D. L., Pitney, W. R., Baker, S. J., and Bradley, J. E. The plasma clearance and urinary excretion of parenterally administered <sup>60</sup>Co B<sub>12</sub>. *Blood* 1956, 11, 31.
4. Heinrich, H. C., and Erdmann-Oehlecker, S. Der Vitamin B<sub>12</sub>-Stoffwechsel bei Hämoblastosen. III. Resorption, Blutverteilung, Serumproteinbindung, Retention und Exkretion der B<sub>12</sub>-Vitamine bei Hämoblastosen nach oraler und parenteraler B<sub>12</sub>-Applikation. *Clin. chim. Acta* 1956, 1, 326.
5. Miller, A., Corbus, H. F., and Sullivan, J. F. The plasma disappearance, excretion, and tissue distribution of cobalt<sup>60</sup> labelled vitamin B<sub>12</sub> in normal subjects and patients with chronic myelogenous leukemia. *J. clin. Invest.* 1957, 36, 18.
6. Raccuglia, G., and Sacks, M. S. Vitamin B<sub>12</sub> binding capacity of normal and leukemic sera. *J. Lab. clin. Med.* 1957, 50, 69.

7. Miller, A. The *in vitro* binding of cobalt<sup>60</sup> labeled vitamin B<sub>12</sub> by normal and leukemic sera. *J. clin. Invest.* 1958, **37**, 556.
8. Pitney, W. R., Beard, M. F., and Van Loon, E. J. Observations on the bound form of vitamin B<sub>12</sub> in human serum. *J. biol. Chem.* 1954, **207**, 143.
9. Heinrich, H. C., and Erdmann-Oehlecker, S. Der Vitamin-B<sub>12</sub>-Stoffwechsel bei Hämoblastosen. II. Die intravitale Bindung (Transport) der B<sub>12</sub>-Vitamine an die Serumproteinfraktionen bei Hämoblastosen. *Clin. chim. Acta* 1956, **1**, 311.
10. Winzler, R. J., Devor, A. W., Mehl, J. W., and Smyth, I. M. Studies on the mucoproteins of human plasma. I. Determination and isolation. *J. clin. Invest.* 1948, **27**, 609.
11. Mehl, J. W., Humphrey, J., and Winzler, R. J. Mucoproteins of human plasma; III. Electrophoretic studies of mucoproteins from perchloric acid filtrates of plasma. *Proc. Soc. exp. Biol. (N. Y.)* 1949, **72**, 106.
12. Miller, A., and Sullivan, J. F. Some physicochemical properties of the vitamin B<sub>12</sub> binding substances of normal and chronic myelogenous leukemic sera. *J. Lab. clin. Med.* 1959, **53**, 607.
13. Ross, G. I. M. Vitamin B<sub>12</sub> assay in body fluids using *Euglena gracilis*. *J. clin. Path.* 1952, **5**, 250.
14. Cohn, E. J., Strong, L. E., Hughes, W. L., Jr., Mulford, D. J., Ashworth, J. N., Melin, M., and Taylor, H. L. Preparation and properties of serum and plasma proteins. IV. A system for the separation into fractions of the protein and lipoprotein components of biological tissues and fluids. *J. Amer. chem. Soc.* 1946, **68**, 459.
15. Kunkel, H. G. Zone electrophoresis in *Methods of Biochemical Analysis*, D. Glick, Ed. New York, Interscience Publishers, 1954, vol. 1, p. 141.
16. Kunkel, H. G., and Tiselius, A. Electrophoresis of proteins on filter paper. *J. gen. Physiol.* 1951, **35**, 89.
17. Folin, O., and Ciocalteu, V. On tyrosine and tryptophane determinations in proteins. *J. biol. Chem.* 1927, **73**, 627.
18. Werner, I., and Odin, L. On the presence of sialic acid in certain glycoproteins and in gangliosides. *Acta Soc. Med. upsalien.* 1952, **57**, 230.
19. Schmid, K. Preparation and properties of serum and plasma proteins. XXIX. Separation from human plasma of polysaccharides, peptides and proteins of low molecular weight. Crystallization of an acid glycoprotein. *J. Amer. chem. Soc.* 1953, **75**, 60.
20. Hine, G. J., and Miller, A. Large plastic well makes efficient gamma counter. *Nucleonics* 1956, **14**, 78.
21. Weimer, H. E., Mehl, J. W., and Winzler, R. J. Studies on the mucoproteins of human plasma. V. Isolation and characterization of a homogeneous mucoprotein. *J. biol. Chem.* 1950, **185**, 561.
22. Fahey, J. L., McCoy, P. F., and Goulian, M. Chromatography of serum proteins in normal and pathologic sera: The distribution of protein-bound carbohydrate and cholesterol, siderophilin, thyroxin-binding protein, B<sub>12</sub>-binding protein, alkaline and acid phosphatases, radioiodinated albumin and myeloma proteins. *J. clin. Invest.* 1958, **37**, 272.
23. Mendelsohn, R. S., Watkin, D. M., Horbett, A. P., and Fahey, J. L. Identification of the vitamin B<sub>12</sub>-binding protein in the serum of normals and of patients with chronic myelocytic leukemia. *Blood* 1958, **13**, 740.
24. Moschides, E., Stefanini, M., Magalini, S. I., and Kistner, S. A. Content and composition of the mucoprotein fraction of human serum (seromucoid) in disease with special reference to hematologic disorders. *J. clin. Invest.* 1958, **37**, 127.
25. Smith, E. L., Brown, D. M., Weimer, H. E., and Winzler, R. J. Sedimentation, diffusion, and molecular weight of a mucoprotein from human plasma. *J. biol. Chem.* 1950, **185**, 569.
26. Schmid, K. Purification and properties of an  $\alpha$ 2-glycoprotein derived from normal human plasma. *Biochim. biophys. Acta* 1956, **21**, 399.
27. Schmid, K. Isolation of a group of  $\alpha$ 2-glycoproteins from human plasma. *J. Amer. chem. Soc.* 1955, **77**, 742.
28. Winzler, R. J. Glycoproteins of plasma in Ciba Foundation Symposium on the Chemistry and Biology of Mucopolysaccharides, G. E. W. Wolstenholme and M. O'Conner, Eds. Boston, Little, Brown and Co., 1958, p. 245.
29. Glass, G. B., Boyd, L. J., Rubinstein, M. A., and Svirgals, C. S. Relationship of glandular mucoprotein from human gastric juice to Castle's intrinsic antianemic factor. *Science* 1952, **115**, 101.
30. Latner, A. L., and Ungley, C. C. Intrinsic factor activity in solid material. *Brit. med. J.* 1953, **1**, 1448.
31. Rambach, W. A., Cooper, J. A. D., and Alt, H. L. Purification of erythropoietin by ion-exchange chromatography. *Proc. Soc. exp. Biol. (N. Y.)* 1958, **98**, 602.
32. Gräsbeck, R. Studies on the vitamin B<sub>12</sub>-binding principle and other biocolloids of human gastric juice. *Acta med. scand.* 154, 1956, Suppl. 314.
33. Bunge, M. B., and Schilling, R. F. Intrinsic factor studies. VI. Competition for vitamin B<sub>12</sub> binding sites offered by analogues of the vitamin. *Proc. Soc. exp. Biol. (N. Y.)* 1957, **96**, 587.
34. Miller, A., and Sullivan, J. F. Excretion of a vitamin B<sub>12</sub>-binding substance in chronic myelogenous leukemic urine. *Clin. Res.* 1959, **7**, 209.
35. Prentice, T. C., and Mirand, E. A. Studies of plasma erythropoietic factor in anemic human patients. *Blood* 1957, **12**, 993.
36. Linman, J. W., and Bethell, F. H. The plasma erythropoietic-stimulating factor in man. Observations on patients with polycythemia vera and secondary polycythemia. *J. Lab. clin. Med.* 1957, **49**, 113.
37. Gurney, C. W., Goldwasser, E., and Pan, C. Studies on erythropoiesis. VI. Erythropoietin in human plasma. *J. Lab. clin. Med.* 1957, **50**, 534.