MerTK receptor cleavage promotes plaque necrosis and defective resolution in atherosclerosis

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Introduction

Atherothrombotic vascular disease is often triggered by a distinct type of atherosclerotic lesion that displays features of impaired inflammation resolution, notably a necrotic core and thinning of a protective fibrous cap that overlies the core. A key cause of plaque necrosis is defective clearance of apoptotic cells, or efferocytosis, by lesional macrophages, but the mechanisms underlying defective efferocytosis and its possible links to impaired resolution in atherosclerosis are incompletely understood. Here, we provide evidence that proteolytic cleavage of the macrophage efferocytosis receptor c-Mer tyrosine kinase (MerTK) reduces efferocytosis and promotes plaque necrosis and defective resolution. In human carotid plaques, MerTK cleavage correlated with plaque necrosis and the presence of ischemic symptoms. Moreover, in fat-fed LDL receptor–deficient (Ldlr−/−) mice whose myeloid cells expressed a cleavage-resistant variant of MerTK, atherosclerotic lesions exhibited higher macrophage MerTK, lower levels of the cleavage product soluble Mer, improved efferocytosis, smaller necrotic cores, thicker fibrous caps, and increased ratio of proresolving versus proinflammatory lipid mediators. These findings provide a plausible molecular-cellular mechanism that contributes to defective efferocytosis, plaque necrosis, and impaired resolution during the progression of atherosclerosis.

Results and Discussion

We analyzed cross sections of carotid artery endarterectomy specimens from 14 patients for soluble Mer (sol-Mer), a marker of MerTK cleavage (11, 12), and plaque necrosis and found that lesional sol-Mer was positively correlated with necrosis (Figure 1A and Supplemental Figure 1A; supplemental material available online with this article; doi:10.1172/JCI90520DS1). Sol-Mer was not detected in nonatherosclerotic human arteries. We next compared carotid plaques from additional subjects who either had suffered a transient ischemic attack or stroke (symptomatic) or were asymptomatic. Plaques of symptomatic patients had higher levels of sol-Mer compared with those of asymptomatic patients (Figure 1B), and plasma sol-Mer was also increased in symptomatic patients (Supplemental Figure 1B).

MerTK cleavage causes defective MerTK-mediated efferocytosis (8, 11, 12). A previous study showed that recombinant sol-Mer can inhibit efferocytosis in vitro, presumably through competition for molecules that bridge apoptotic cells (ACs) to MerTK, like GAS6 (11). However, we found that recombinant sol-Mer inhibited efferocytosis only at doses that were more than 1,000-fold those in plasma (Supplemental Figure 1C). Thus, unless sol-Mer concentrations in lesions reach extremely high levels, sol-Mer is most likely a marker rather than a cause of defective MerTK efferocytosis. In this context, we found that cell-surface MerTK in macrophage-rich areas in plaques from symptomatic patients was less than that in asymptomatic patients (Figure 1C) despite sim-
To study advanced atherosclerosis, WT or MertkCR bone marrow was transplanted into Ldlr−/− mice, and atherosclerotic lesions and plasma were analyzed after 16 weeks of WD feeding. These 2 groups had similar body weight, blood glucose, plasma lipids, and circulating leukocytes (Supplemental Figures 3 and 4). We confirmed that aortic sol-Mer was reduced and that MerTK was higher in the CR cohort (Figure 2, A and B). Most importantly, MertkCR lesions had substantially less plaque necrosis but no change in total lesion area or lesional cell composition (Figure 2C and Supplemental Figure 5), indicating a true decrease in percentage necrosis.

Further, MertkCR lesions had a markedly lower ratio of free to macrophage-associated TUNEL+ (apoptotic) cells (Figure 2D), which reflects improved efferocytosis.

MertkCR lesions also had thicker fibrous cap thickness and increased collagen gene expression (Figure 3, A and B), which are associated with plaque stability and inflammation resolution (1, 15). These lesions also had more Tregs (Figure 3C), which can promote resolution (16, 17), are decreased in human and mouse advanced atheroma (18), and have been causatively linked to plaque progression in mice (19).

The inflammation resolution program has a number of positive-feedback processes. First, SPMs can increase production of proteins that dampen inflammation and promote repair (22, 23).

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Two such proteins, TGF-β1 and IL-10, were significantly increased in MertkCR plasma (Supplemental Figure 6A). Second, SPMs can upregulate their own receptors (24), and we found that the expression of the SPM receptor ALX/FPR2 was markedly induced in the lesions of MertkCR mice (Supplemental Figure 6B). Third, engagement of MerTK by apoptotic cells and other ligands in cultured macrophages and in sterile inflammation in vivo enhances the synthesis of 5-LOX–derived SPMs at the expense of inflammatory leukotrienes (8, 25). We therefore analyzed the ratio of 5-LOX–derived SPMs to leukotrienes and found that this ratio was markedly increased in MertkCR extracts (Figure 3E). Moreover, this ratio was negatively correlated with necrotic core area (Figure 3F).

Fourth, using bone marrow–derived macrophages, we found that resolvin D1 (RvD1) blocks LPS-induced MerTK cleavage (Supplemental Figure 7, A and B). Importantly, when WD-fed Ldlr−/− mice were treated with RvD1, which suppresses plaque necrosis and improves lesional efferocytosis (20), lesional MerTK expression was increased (Supplemental Figure 7C). Thus, MerTK cleavage during plaque progression suppresses the integrated processes of efferocytosis and inflammation resolution and disrupts several components of positive-feedback signaling. These findings likely explain the marked improvement in plaque morphology in the atherosclerotic lesions of MertkCR→Ldlr−/− mice. Understanding processes involved in the formation of the unique types of atherosclerotic plaques that cause acute cardiovascular events is a critical goal in cardiovascular research. Progression to plaque necrosis is a key process because of the highly inflammatory and plaque-destabilizing nature of the necrotic core (26). Data from human and mouse studies suggest that plaque necrosis results from 2 complementary processes, postapoptotic necrosis secondary to defective efferocytosis as examined here (2, 3) and a primary cell necrosis process called necroptosis (27, 28). While the current study is focused on an efferocytosis defect in phagocytes themselves, a recent study showed that some apoptotic cells in human and mouse lesions inappropriately retain CD47, which blocks their engulfment by normal phagocytes (29). Interestingly, oxLDL, which was shown here to promote MerTK cleavage and defective efferocytosis, can induce necroptosis in macrophages, and these cells are also poorly

Figure 2. Suppression of MerTK cleavage in myeloid cells improves efferocytosis and decreases plaque necrosis in WD-fed Ldlr−/− mice. (A) WT→Ldlr−/− or MertkCR→Ldlr−/− bone marrow–transplanted mice were fed WD for 16 weeks. Aortas were harvested and analyzed for sol-Mer as described in Figure 1D (n = 3 per group). *P < 0.05. (B) Confocal microscopy of MerTK in aortic root cross sections (scale bar: 20 μm). Images were quantified as MerTK MFI relative to the WT→Ldlr−/− cohort (n = 8 per group). *P < 0.01. (C) Representative H&E images of aortic root sections, with necrotic core (NC) regions indicated by broken lines (scale bar: 40 μm), and quantification of necrotic core area (n = 10 per group, 2 independent experiments). *P < 0.01. (D) Representative images of aortic root sections in which apoptotic cells were labeled by TUNEL (red), macrophages by anti-F4/80 (green), and nuclei by Hoechst (blue) (scale bar: 10 μm). The white arrows depict apoptotic cells that were either free (top image) or associated with macrophages (bottom image). The graph shows quantification of the ratio of free to macrophage-associated apoptotic cells (n = 10 for each group, 2 independent experiments). *P < 0.05. A 2-tailed Student’s t test was used for all panels.
internalized by normal phagocytes (28). Defects in multiple steps of efferocytosis, together with activation of necroptotic pathways in advanced plaques, would create a “perfect storm” for the development of plaque necrosis.

Our data support the concept that MerTK plays an integral part in a program of feedback signaling that amplifies resolution. This concept aligns with previous studies showing that (a) engagement of MerTK by apoptotic cells or other activators can activate both proinflammatory and proresolving pathways (6–8, 30–32); and (b) proresolving mediators can promote MerTK expression and efferocytosis (7, 33, 34). Thus, when MerTK is rendered inactive by proteolytic cleavage, not only is there a defect in efferocytosis, but positive-feedback signaling in resolution is impaired, leading to loss of resolving mediators and further defects in efferocytosis. In the setting of advanced atherosclerosis, this profound deficit promotes key features of clinically dangerous plaques, including necrosis, imbalance of proresolving versus proinflammatory lipid mediators, fibrous cap thinning, and deficiency of Tregs. The finding that cleavage of a single molecule in the setting of a chronic inflammatory condition could have such robust pathophysiologic effects not only suggests its potential as a therapeutic target but also raises the question of the possible role of MerTK cleavage in other chronic inflammatory conditions previously linked to MerTK and sol-Mer, e.g., systemic lupus erythematosus and Sjögren’s syndrome (35–38).

Methods
A complete description of methods is provided in the Supplemental Methods.

Statistics. Data are displayed as mean ± SEM. Because all data in this study fit a normal distribution, a 2-tailed Student's t test was used to determine statistical significance. P values less than 0.05 were considered significant. For the data in Figure 1A, correlation coefficient (R) and P value were calculated using Pearson product-moment correlation analysis.

Study approval. For human studies, carotid endarterectomy samples were obtained from M.J.A.P. Daemen, Academic Medical Center, Amsterdam, the Netherlands, and Bernhard Dorweiler, University Medical Center, Johannes-Gutenberg University, Mainz, Germany. Use of all material conformed with the declaration of Helsinki and was approved by the appropriate university ethics review board. Complete details are available in the Supplemental Methods. All animals were cared for according to NIH and IACUC guidelines in a barrier facility at Columbia University Medical Center, New York, New York, USA.

Author contributions
BC, EBT, GF, and IT conceived and designed the research. MJAPD and BD donated patient samples and were instrumental in the interpretation of the human data. BC, EBT, BES, MS, and GF conducted the experiments. BC, EBT, ACD, BES, GF, MS, and IT analyzed the data. BC, ACD, and IT wrote the paper.
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