Large-scale genome-wide analysis identifies genetic variants associated with cardiac structure and function


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BACKGROUND. Understanding the genetic architecture of cardiac structure and function may help to prevent and treat heart disease. This investigation sought to identify common genetic variations associated with inter-individual variability in cardiac structure and function.

METHODS. A GWAS meta-analysis of echocardiographic traits was performed, including 46,533 individuals from 30 studies (EchoGen consortium). The analysis included 16 traits of left ventricular (LV) structure, and systolic and diastolic function.

RESULTS. The discovery analysis included 21 cohorts for structural and systolic function traits (n = 32,212) and 17 cohorts for diastolic function traits (n = 21,852). Replication was performed in 5 cohorts (n = 14,321) and 6 cohorts (n = 16,308), respectively. Besides 5 previously reported loci, the combined meta-analysis identified 10 additional genome-wide significant SNPs: rs12541595 near MTSS1 and rs10774625 in ATXN2 for LV end-diastolic internal dimension; rs806322 near KCNNG, rs4765663 in CACNA1C, rs6702619 near PALMD, rs7127129 in TMEM16A, rs11207426 near FGGY, rs17603876 in GOSR2, and rs17696696 in CFDP1 for aortic root diameter; and rs12440869 in IQCH for Doppler transmural A-wave peak velocity. Findings were in part validated in other cohorts and in GWAS of related disease traits. The genetic loci showed associations with putative signaling pathways, and with gene expression in whole blood, monocytes, and myocardial tissue.

CONCLUSION. The additional genetic loci identified in this large meta-analysis of cardiac structure and function provide insights into the underlying genetic architecture of cardiac structure and warrant follow-up in future functional studies.

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Introduction

Heart failure (HF) is associated with substantial morbidity, mortality, and health care costs, and is increasing in prevalence with the aging of the global population (1). Hence, prevention and treatment of HF by identifying its genetic and environmental determinants is a public health priority. The identification of the

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genetic architecture of HF may be facilitated by evaluating echocardiographic traits of left ventricular (LV) structure and function. These heritable, quantitative traits can antedate HF and are more amenable to genetic analysis than more “distal” heart disease traits (2). Initial studies that related common genetic variants to echocardiographic traits and incident HF (2–5) were limited by modest sample size, analysis of only a few echocardiographic phenotypes, or evaluation of “all HF,” a phenotypically heterogeneous group (6–9).

We conducted a meta-analysis of genome-wide association studies (GWAS) on a comprehensive set of echocardiographic traits in carefully phenotyped individuals primarily of European ancestry within the EchoGen consortium (2) comprising 30 studies. We associated our identified genetic loci with echocardiographic traits in other ethnicities, in populations with related disease traits. Additionally, we further characterized loci by evaluating putative signaling pathways and examining their association with gene expression in whole blood, monocytes, and cardiac tissue.

**Results**

Cohort descriptions and the echocardiographic characteristics are presented in Supplemental Tables 1–5; supplemental material available online with this article; https://doi.org/10.1172/JCI84840DS1.

Individual study genomics inflation factors are shown in Supplemental Table 6. The meta-analytic genomic inflation factor (λ) was 1.09 or less for all traits evaluated. The genomic inflation factors for the traits with significant results (see below) were 1.09 (for aortic root diameter [AoD]) and 1.08 (for LV diastolic internal dimension [LVDD]). To address to what extent the genomic inflation might be due to unaccounted population stratification versus truly associated genetic markers, we applied the recently developed linkage disequilibrium (LD) score regression method to these two traits (10). The genomic inflation factor due to potential confounding bias reduced to 1.05 for AoD and to 1.03 for LVDD, suggesting that our meta-analytic approach accounted for population stratification reasonably well. Quantile-quantile (Q-Q) plots are shown in Supplemental Figures 1-16.
Table 1. Genetic loci associated with echocardiographic traits of LV structure and systolic function with genome-wide significance at $P < 5.0 \times 10^{-8}$ in the discovery dataset, replication results, and a meta-analysis combining discovery and replication data

| SNP          | Chr | Position | Nearest gene | Distance to nearest gene (kb) | SNP annotation | Effect/non-effect allele | EAFA | Discovery $P$ | Replication $P$ | Combined meta-analysis $P$ | Heterogeneity I | Heterogeneity $P$ |
|--------------|-----|----------|--------------|-------------------------------|----------------|--------------------------|------|---------------|----------------|--------------------------|----------------|----------------|}
| rs806322     | 13  | 49739445 | KCNRG        | 246.4                         | Unknown        | A/G                      | 0.61 | $6.70 \times 10^{-15}$ | 0.035          | $-0.021 (0.003)$          | 2.22 $\times 10^{-15}$ | 0              | 0.620          |
| rs6702619     | 1   | 99818834 | PALMD        | 65.4                          | Unknown        | G/T                      | 0.50 | $6.89 \times 10^{-15}$ | $3.84 \times 10^{-3}$ | 0.021 (0.003)           | $<1.10 \times 10^{-16}$ | 0              | 0.409          |
| rs10770612    | 12  | 20121906 | PDE3A        | 291.6                         | Unknown        | A/G                      | 0.80 | $3.20 \times 10^{-12}$ | –             | –                        | –               | –              | –              |
| rs17469907    | 5   | 12556319 | CCDC100      | 152.1                         | Unknown        | G/T                      | 0.72 | $1.02 \times 10^{-11}$ | –             | –                        | –               | –              | –              |
| rs1532952     | 17  | 204423   | SMG6         | 0                             | Intron         | T/G                      | 0.61 | $1.29 \times 10^{-11}$ | –             | –                        | –               | –              | –              |
| rs10878359    | 12  | 64690891 | HMGA2        | 44.6                          | Unknown        | T/C                      | 0.36 | $1.62 \times 10^{-11}$ | –             | –                        | –               | –              | –              |
| rs17696936    | 16  | 73950853 | CFDP1        | 0                             | Intron         | G/T                      | 0.59 | $1.96 \times 10^{-9}$  | 0.079          | $-0.016 (0.003)$         | 2.68 $\times 10^{-10}$ | 0              | 0.578          |
| rs7127129     | 11  | 69705561 | TMEM16A      | 0                             | Intron         | G/A                      | 0.41 | $2.45 \times 10^{-9}$  | 0.303          | $-0.015 (0.003)$         | 2.44 $\times 10^{-9}$ | 0.20           | 0.292          |
| rs17608765    | 17  | 42382730 | GOSR2        | 0                             | Intron         | C/T                      | 0.14 | $4.28 \times 10^{-9}$  | 0.020          | 0.0244 (0.0038)          | 2.25 $\times 10^{-10}$ | 0.66           | 0.032          |
| rs2649        | 15  | 61673646 | USP3         | 2.9                           | Untranslated-3' | T/C                      | 0.13 | $1.01 \times 10^{-8}$  | 0.535          | $-0.021 (0.004)$         | 5.37 $\times 10^{-8}$ | 0.67           | 0.029          |
| rs4765668     | 12  | 2049021  | CACNA1C      | 0                             | Intron         | C/G                      | 0.16 | $1.39 \times 10^{-8}$  | 0.068          | $-0.020 (0.003)$         | 4.00 $\times 10^{-9}$ | 0              | 0.925          |
| rs11207426    | 1   | 59458507 | FGGY         | 76.8                          | Unknown        | A/G                      | 0.37 | $2.93 \times 10^{-8}$  | 0.021          | 0.017 (0.003)           | 2.76 $\times 10^{-9}$ | 0              | 0.518          |

LVDD (cm)

| SNP          | Chr | Position | Nearest gene | Distance to nearest gene (kb) | SNP annotation | Effect/non-effect allele | EAFA | Discovery $P$ | Replication $P$ | Combined meta-analysis $P$ | Heterogeneity I | Heterogeneity $P$ |
|--------------|-----|----------|--------------|-------------------------------|----------------|--------------------------|------|---------------|----------------|--------------------------|----------------|----------------|}
| rs1115370    | 6   | 11874215 | SLC35F1      | 28.7                          | Unknown        | T/C                      | 0.51 | $6.40 \times 10^{-6}$ | –             | –                        | –               | –              | –              |
| rs12541595    | 8   | 12592640 | MTSSL1       | 116.7                         | Unknown        | T/G                      | 0.30 | $3.02 \times 10^{-12}$ | $4.03 \times 10^{-3}$ | $-0.023 (0.003)$       | $1.65 \times 10^{-10}$ | 0              | 0.513          |
| rs10746252    | 12  | 11039462 | ATXN2        | 0                             | Intron         | G/A                      | 0.50 | $1.90 \times 10^{-8}$ | 0.066          | 0.016 (0.003)           | $1.28 \times 10^{-8}$ | 0.67           | 0.011          |

LV (g)

| SNP          | Chr | Position | Nearest gene | Distance to nearest gene (kb) | SNP annotation | Effect/non-effect allele | EAFA | Discovery $P$ | Replication $P$ | Combined meta-analysis $P$ | Heterogeneity I | Heterogeneity $P$ |
|--------------|-----|----------|--------------|-------------------------------|----------------|--------------------------|------|---------------|----------------|--------------------------|----------------|----------------|}
| rs954157     | 4   | 17759792 | SPES3        | 108.4                         | Unknown        | C/T                      | 0.73 | $4.41 \times 10^{-9}$ | 0.301          | 1.384 (0.260)           | 9.68 $\times 10^{-8}$ | 0.52           | 0.066          |
| PS (%)       | 4   | 17759792 | SPES3        | 108.4                         | Unknown        | C/T                      | 0.73 | $4.41 \times 10^{-9}$ | 0.301          | 1.384 (0.260)           | 9.68 $\times 10^{-8}$ | 0.52           | 0.066          |

From combined meta-analysis. As a proxy for rs2762049, $R^2 = 1.0$, $D' = 1.0$. Locus found in discovery phase but not replicated in the previously published meta-analysis (2). Located within enhancer histone marks in ENCODE (17). Located within DNase-hypersensitive sites in ENCODE (17). Locus colocalizes with DEPICT prioritized gene (Supplemental Table 15). Known locus (2), not taken forward for replication. Significantly associated with transcripts in cis (see text for details). Chr, chromosome; EAFA, effect allele frequency; LVDD, LV diastolic internal dimension; AoD, diameter of the aortic root; FS, fractional shortening. Boldface indicates novel replicated findings. Effects are $\beta$ coefficients, which represent the change in echocardiographic measure in the units shown in the subheads (i.e., cm, g, or %) per unit difference in effect allele dose.
Single nucleotide polymorphisms related to cardiac structure and function (stage 1). We applied a two-stage design proposed by Skol et al. (11), including an additional stage for assessing the generalizability of the find, with details on samples and single nucleotide polymorphisms (SNPs) for each stage given in Figure 1. The meta-analysis of LV cardiac structure and systolic function traits included data from 21 cohorts with up to 30,201 individuals. For LV diastolic function, data were available from 17 cohorts with up to 21,852 individuals. We identified genome-wide significant associations (all \( P < 5 \times 10^{-8} \)) of: 1 locus with LV mass (LVM), 3 with LVDD, 12 with AoD, 1 with LV fractional shortening (LVFS). Additionally, the following associations were observed at a higher \( P \) value threshold (all \( P < 1 \times 10^{-5} \)): 2 with the peak velocity of the transmitral E-wave (Mv-E), 5 with the peak velocity of the transmitral A-wave (Mv-A), 5 with the ratio of Mv-E to Mv-A (E/A), 2 with deceleration time of Mv-E (DecTime), 4 with isovolumetric relaxation time (IVRT), 1 with the peak velocity of the excursion of the lateral mitral annulus in the early diastolic phase (E'), 3 with the ratio of Mv-E to E' (E/E'), 1 with asymptotic LV diastolic dysfunction with preserved ejection fraction (DDpEF), and 2 with HF with preserved ejection fraction (HFP EF). Using pre-defined selection criteria (Figure 1) and excluding known loci from our previous report (2), 12 SNPs for traits of cardiac structure and LV systolic function (Table 1) and 24 SNPs for traits of LV diastolic function (Table 2) were considered for additional analysis detailed in stage 2 below. Full results for all SNPs with \( P < 1 \times 10^{-4} \) are shown in Supplemental Table 7.

Replication and combined meta-analysis (stage 2). SNPs taken forward for stage 2 replication were analyzed in 5 cohorts (\( n = 14,002 \); 2 with in silico GWAS data, 3 with de novo genotyping) for cardiac structure and LV systolic function; and in 6 cohorts (\( n = 14,787 \); 3 with in silico GWAS data, 3 with de novo genotyping) for LV diastolic function (Figure 1). A final combined meta-analysis of discovery and replication data from overall 30 cohort samples included 44,203 individuals with data on cardiac structure and systolic function, and 36,639 individuals with data on LV diastolic function. The investigation revealed 10 SNPs with genome-wide significance: rs10774625 and rs12541595 for LVDD; rs806322, rs4765663, rs6702619, rs7127129, rs11207426, rs17608766, and rs17696696 for AoD; and rs12440869 for Mv-A (Tables 1 and 2). Manhattan plots for these 3 traits are presented in Figure 2. Forest plots for the most significantly associated SNPs for AoD (rs6702619), LVDD (rs12541595), and Mv-A (rs12440869) with the corresponding regional plots including functional annotation are presented in Figures 3, 4, and 5. The plots for the other genome-wide significant loci are shown in Supplemental Figures 17 and 18. Funnel plots for the significantly associated SNPs are shown in Supplemental Figure 19. All known and novel loci combined explained 1.7%, 0.5%, and 0.2% of the phenotypic variance in AoD, LVDD, and Mv-A, respectively, in a combined analysis of 3 of the larger cohorts.

Findings in children, other ethnicities, and related cardiovascular phenotypes (stage 3). In stage 3, the genome-wide significant SNPs were investigated for generalizability of the observed associations; small sample sizes of available cohorts partly limited the statistical power to replicate findings. In this exploratory analysis, we only found one weak association with AoD in white children of Europe-an ancestry in the Generation R study (12), and none in Hispanics (Northern Manhattan Study [NOMAS] study) or African Americans (Jackson Heart Study [JHS] and NOMAS study; Supplemental Table 8). When evaluating associations of the newly discovered SNPs with related disease traits, rs17696696, which was found to be associated with AoD, was also associated with pulse wave velocity in the AortaGen consortium (Supplemental Table 9 and ref. 13). There were no statistically significant associations with incident HF or mortality in HF patients of the CHARGE-Heart Failure (CHARGE-HF) consortium (Supplemental Table 10), or with all-cause mortality, HF, or cardiovascular mortality in the Ludwigshafen Risk and Cardiovascular Health (LURIC) cohort of patients with suspected coronary artery disease (CAD) (Supplemental Table 11). In the CARDIOGRAMplusC4D consortium data, rs17696696, rs17608766, and rs10774625 were significantly associated with CAD; rs10774625 was also strongly associated with the narrower phenotype myocardial infarction (MI; \( P = 5.09 \times 10^{-4} \), Supplemental Table 12).

Biological pathways related to echocardiographic traits. In pathway analysis, the observed genetic variants were significantly enriched for canonical pathways that might be involved in the biological regulation of echocardiographic traits: protein kinase A signaling (\( P = 5.8 \times 10^{-5} \)), death receptor signaling (\( P = 6.9 \times 10^{-5} \)), the Wnt/Ca\(^{2+} \) pathway (\( P = 2.2 \times 10^{-5} \)), and P2Y purinergic receptor signaling (\( P = 4.1 \times 10^{-5} \), Supplemental Tables 13 and 14, Supplemental Figure 20, and refs. 14–16).

When investigating the potential regulatory effect of the top loci using Encyclopedia of DNA Elements (ENCODE) data (17), 4 SNPs (rs10774625, rs6702619, rs17608766, and rs11207426) were located within enhancer histone marks and 4 (rs806322, rs6702619, rs7127129, and rs17608766) within DNase-hypersensitive sites. The literature search tool Snipper revealed no additional information, and no significant direct or indirect protein-protein interactions were found between loci using DAPPLE software (18). No significantly reconstituted gene sets were identified by the DEPICT tool (ref. 19 and Supplemental Table 15). DEPICT prioritized (false discovery rate [FDR] < 0.05) 10 genes across associated (\( P < 1 \times 10^{-4} \)) loci, including 4 colocalizing with genome-wide significant loci (Tables 1 and 2, and Supplemental Table 15).

Analyses of expression quantitative trait loci and gene expression in whole blood, monocytes, and myocardial tissue. Our data showed 4 SNPs that were significantly associated with cis transcripts in both datasets (whole blood and monocytes, Supplemental Table 16): rs10774625 with SH2B adaptor protein 3 (SH2B3, \( P = 8.15 \times 10^{-20} \) and \( P = 1.83 \times 10^{-3} \)), rs17696696 with craniofacial development protein 1 (CFDPI, \( P = 6.21 \times 10^{-10} \) and \( P = 7.59 \times 10^{-8} \)), rs7127129 with Fas-associated death domain–containing protein (FADD, \( P = 1.61 \times 10^{-9} \) and \( P = 2.71 \times 10^{-8} \)), and rs1532292 with serine racemase (SRR, \( P = 3.40 \times 10^{-20} \) and \( P = 4.63 \times 10^{-10} \)).

We also examined the associations of our top loci with gene expression in human LV tissue provided by the Myocardial Applied Genomics Network consortium (MAGNet consortium; unpublished data). Two SNPs were significantly associated with LV gene expression: rs12541595 showed cis-association with metastasis suppressor 1 (MTSSI, \( P = 1.25 \times 10^{-8} \)), with the effect allele T associated with lower MTSSI expression; rs1532292 showed again a cis-association with SRR (\( P = 2.62 \times 10^{-4} \), with the effect allele T...
Table 2. Genetic loci associated with echocardiographic traits of diastolic function with $P < 1.0 \times 10^{-6}$ in the discovery dataset, replication results, and a meta-analysis combining discovery and replication data

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<td>$1.60 \times 10^{-7}$</td>
<td>$1.57 \times 10^{-7}$</td>
<td>$1.67 \times 10^{-7}$</td>
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<td>63258362</td>
<td>FAM105B</td>
<td>0.01</td>
<td>$1.50 \times 10^{-7}$</td>
<td>$1.48 \times 10^{-7}$</td>
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<td>62040753</td>
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</table>

From combined meta-analysis. *P* for SNP was 1.0 × 10⁻⁶. Effects are β coefficients, which represent the change in echocardiographic measure in the units shown in the subheads (i.e., cm, g, or %) per unit difference in effect allele dose.
associated with lower SRR expression. Both expression quantitative trait locus (eQTL) associations from the LV tissue were also supported by the GTEx database (http://gtexportal.org/home/). The association with SRR expression for rs1532292 had the same direction of effect in different tissues, with the T allele generally associated with lower gene expression levels, e.g., in the aorta and in blood cells. Additionally, the following eQTLs with genes from the reference sequence database (RefSeq; https://www.ncbi.nlm.nih.gov/refseq/) in the aorta or heart tissue were found for the replicated SNPs in the GTEx database: rs17696696 (BCAR1), rs12541595 (LINC00964), and rs11153730 (SSX9P10). Detailed GTEx results are given in Supplemental Table 17.

**Discussion**

In the present investigation, we identified 7 genetic loci associated with aortic root size and confirmed the associations of 4 other loci previously reported (2). These 11 variants explained 1.7% of the inter-individual variation in aortic root size (Supplemental Table 18). However, use of genome-wide complex trait analysis (GCTA) software in one of the larger cohorts (Study of Health in Pomerania [SHIP]) as an illustrative example demonstrated that common genetic variation explains about 30% of the variation in AoD (Supplemental Table 19), underscoring the potential for more, as-yet-undiscovered, loci. Additionally, we observed three genetic loci that were associated with LV diastolic dimensions (including one previously reported; see below) and one locus that was associated with the transmitral A-wave velocity.

Among the SNPs identified in our study as being associated with LVDD, one was rs12541595 close to MTSS1, which interacts with cytoplasmic actin near the cell surface and modulates intercellular connections in the kidney and metastatic potential in tumors (20, 21). When investigating our top loci for cis-associations with gene expression in human LV myocardial tissue (MAGNet consortium, unpublished data) and the GTEx database, rs12541595 showed a significant association with MTSS1 expression, with the LVDD-lowering allele (T) associated with lower MTSS1 expression in this tissue (Supplemental Table 9). We speculate that a reduction in MTSS1 may promote favorable LV remodeling, perhaps by affecting cell junctions. The other novel variant associated with LVDD, rs10774625, was associated with expression of SH2B3 in eQTL analysis and lies in ATXN2 (ataxin 2),
which is adjacent to SH2B3, previously associated with retinal venular diameter, CAD, and arterial hypertension in separate reports (22–26). For LVDD, we also replicated the previously identified SLC35F1 locus (soluble transporter membrane protein) adjacent to the phospholamban (PLN) locus (protein inhibiting cardiac muscle sarcoplasmic reticulum Ca++-ATPase) (2).

Three loci associated with AoD have been linked previously to blood pressure as well as MI (GOSR2, Golgi SNAP receptor complex member 2; refs. 24, 27), blood pressure response to treatment (CACNA1C, calcium channel, voltage-dependent, L type, alpha 1C subunit; ref. 28), and carotid intimal-medial thickness, as well as with CAD (CFDP1; refs. 29, 30). The other novel AoD-associated genetic loci were in or close to PALMD (palmdelphin, a paralemmin-related cytosolic protein; ref. 31), KCNRG (soluble protein with regulatory function in voltage-gated potassium channels; ref. 32), FGGY (carbohydrate kinase domain–containing protein, phosphorylates carbohydrates; ref. 33), and in TMEM16A (transmembrane member 16A, protein involved in transepithelial anion transport and smooth muscle contraction; ref. 34). We also replicated in our discovery sample 4 loci associated with aortic diameter from our previous report (2): SMG6 (Smg-6 homolog, nonsense-mediated mRNA decay factor), CCDC100 (centrosomal protein 120kDa), HMGA2 (high-mobility group AT-hook 2), and PDE3A (phosphodiesterase 3A, cGMP-inhibited). The effect allele of rs1532292 was associated with lower SRR expression in human LV myocardial tissue (unpublished data from the MAGNet consortium; GTEx database, see Supplemental Table 9).

One of the SNPs associated with AoD in our meta-analysis was also associated with AoD in children in the Generation R Study. Additionally, one SNP was associated with pulse wave velocity. Two SNPs associated with AoD and one SNP associated with LVDD were also significantly associated with CAD, the LVDD SNP also with MI in the CARDIOGRAMplusC4D consortium. These associations strengthen the evidence of involvement of these loci in echocardiographic traits. However, given the sample sizes of cohorts with different ethnicities as well as the SNP allele frequencies, and taking the effect sizes into account, the power was not more than 35% to reveal a statistically significant association of select SNPs with traits in “look-up” exercises. Therefore, some of the null results for the assessment of the generalizability of observed associations to non-European samples should be interpreted with care.

Pathway analysis suggested enrichment of the Wnt/Ca++ canonical pathway among the genetic variants associated with echocardiographic traits. However, given the sample sizes of cohorts with different ethnicities as well as the SNP allele frequencies, and taking the effect sizes into account, the power was not more than 35% to reveal a statistically significant association of select SNPs with traits in “look-up” exercises. Therefore, some of the null results for the assessment of the generalizability of observed associations to non-European samples should be interpreted with care.

Pathway analysis suggested enrichment of the Wnt/Ca++ canonical pathway among the genetic variants associated with echocardiographic traits. These observations are consistent with the known effects of this pathway on myocardial biology (35). The Wnt/Ca++ pathway connects to the nuclear factor of activated T cells (NFAT) transcription factor (14, 15) and gene expression via calcineurin. Interestingly, both calcineurin and its target NFAT are involved in cardiac hypertrophy (16).

The association of our findings with expression data from human blood revealed 4 genes with potential functional signif-
likely have further improved diagnosis and classification of LV diastolic dysfunction in our study if this method had been available in more cohorts. Likewise, as noted above, several of the LV diastolic filling measures are notoriously susceptible to variation in ventricular loading conditions (38). The genetic variants identified in our study have small effect sizes and explain a relatively small percentage of the variance in the echocardiographic phenotypes. Larger studies with more detailed reference panels, as well as more detailed functional studies and studies into the interactions of the variants found with factors such as hypertension, will likely shed further light on the molecular mechanisms underlying these complex traits. Furthermore, alterations of the transmitral A wave velocity are challenging to interpret alone, without consideration of other measures of LV diastolic function and filling patterns. The transmitral A wave velocity reflects the late diastolic phase of the LV filling, i.e., the phase of atrial contraction. Thus, in theory this single measure provides important information about active atrial function. Yet in practice, this measure changes variably and in a complex manner with the progression of LV diastolic dysfunction: Increasing impaired ventricular relaxation is at first accompanied by a decrease in E-wave with a compensatory increase in A-wave, resulting in a “relaxation abnormality” pattern; it results in the further, continuous decrease in A-wave velocity, reflecting a progressive deterioration of the contractility of the left atrium, and also changes in LV compliance (39, 40). These pathophysiological considerations underline the importance of the active contraction.

Figure 4. Forest plot for the meta-analysis of the association between rs12541595 and LVDD, with the corresponding regional plot including functional annotation. P values were obtained by calculating Wald test statistics using a sample size of n = 30,201. Total sample size in the forest plot is n = 43,623.

The association between rs12541595 and LVDD is shown in Figure 4. The forest plot displays the results of the meta-analysis, with the corresponding regional plot including functional annotation. P values were obtained by calculating Wald test statistics using a sample size of n = 30,201. Total sample size in the forest plot is n = 43,623.
For analysis of LV diastolic dysfunction, we excluded individuals with reduced ejection fraction (EF) (defined as <50%, LVFS <29% or poor/impaired LV systolic function by visual estimation).

To conclude, we report the largest genetic association study to our knowledge of a comprehensive set of LV echocardiographic traits. The large number of interesting genetic loci identified for AoD and LV diastolic dimensions, and the biological pathways enriched within our association results provide new insights into the biology of cardiac remodeling. Additional studies are warranted to further evaluate experimentally the functional significance of the reported genetic variants and loci.

Methods

EchoGen consortium

The EchoGen consortium was initiated in 2007 and has grown to a consortium of 30 studies with population-based and hospital-based cohorts primarily of European ancestry, and additionally including two cohorts of African American and one of Hispanic individuals. For the present investigation, we applied harmonized phenotype definitions, covariate selection, and genotyping protocols and the same statistical analysis plan across all cohorts. For traits of cardiac structure and systolic function, individuals with a history of MI, clinical diagnoses of HF, or valve disease were excluded if this information was known or recorded during the echocardiographic examination.

For analysis of LV diastolic dysfunction, we excluded individuals with reduced ejection fraction (EF) (defined as <50%, LVFS <29% or poor/impaired LV systolic function by visual estimation).

Strategy for analysis

For the identification of genetic variants associated with cardiac structure and function, we followed a 3-stage analysis plan (Figure 1). First, a discovery meta-analysis of up to 21 population- and hospital-based GWAS was performed (stage 1). Second, replication of the findings from stage 1 was performed in up to 6 independent cohort studies (3 with in silico data and 3 with de novo genotyping), and a combined meta-analysis of discovery and replication data was carried out (stage 2). In stage 3, SNPs that were genome-wide significant in the combined meta-analysis were investigated for the generalizability of the observed associations in a cohort of white children of European ancestry (the Generation R study), in two cohorts of other ethnicities (Hispanic in the NOMAS Study and African American in the JHS and in the NOMAS study), and in associations with related disease traits (data from the AortaGen and CHARGE-HF consortia, and the LURIC study).

Echocardiographic methods

Detailed echocardiographic methods used and distributions of traits in each cohort study are reported in Supplemental Methods and Supplemental Tables 3 and 4.

The present investigation focused on 5 traits of cardiac structure: LVM, LVDD, LVWT, AoD, and left atrial size (LA). Additionally,
we evaluated 2 traits of systolic cardiac function (LVFS and LVSD) and 9 traits of LV diastolic function: Mv-A, Mv-E, E/A, E’, the ratio E/E’ as a surrogate for LV end-diastolic pressure, DecTime, and IVRT, as well as DDPpEF and HFpEF (41). Measurements were based on the European and American guidelines for the echocardiographic assessment of the LV (42).

Genotyping methods and imputation
Details on genotyping, imputation, and quality control are presented in Supplemental Table 5. Population stratification as well as family structure, if applicable, was accounted for in each individual cohort’s analysis. For replication, 3 of the 6 cohorts (Gutenberg Health Study III [GHS-III]; Cardiovascular Risk Factors, Living and Ageing in Halle [CARLA] study; and Malmö Preventive Project [MPP] study) underwent de novo genotyping using 5’ nuclease assays on 384-well plates. For quality control, genotypes were validated in 10% of the samples for all SNPs.

Definition of traits and statistical methods
Discovery (stage 1). All traits were analyzed as continuous traits, with the exception of LVSD, DDPpEF, and HFpEF. LVSD was defined as an EF <50%, fractional shortening (FS) <29% or reduced (poor or impaired) EF by visual estimation. Aggregate binary phenotypes were defined for asymptomatic participants with echocardiographic evidence of LV DDPpEF and for those with HFpEF based on information on classes of HF according to the New York Heart Association (NYHA) and medication for HF in addition to echocardiography. Stage 1 analyses were first performed separately at the individual cohort level for each trait (Figure 1). Continuous echocardiographic traits were related to genotype dosage (0–2 copies of the effect allele) for each autosomal SNP using linear regression assuming additive genetic models adjusted for age, sex, height, weight, and study site (only applicable for the Cardiovascular Health Study [CHS] and Anglo-Scandinavian Cardiac Outcomes Trial [ASCOT]). For binary traits, we used logistic regression models with the same adjustments. In the Framingham Heart Study (FHS), linear-mixed-effects models were applied to account for familial correlations. The associations of genotypes with echocardiographic traits were quantified by beta estimates, SEM, and P values. After verifying strand alignment across studies and applying genomic control to each study, we performed inverse variance–weighted fixed-effects meta-analysis across the discovery cohorts with METAL (43) for the structural and the systolic function traits and with the R package MetABEL (http://www.r-project.org) for the diastolic traits. After the meta-analysis, we excluded SNPs with a minor allele frequency (MAF) below 0.5% for diastolic function traits and below 1% for structural traits, and FS and below 3% for LVSD.

We used an a priori P value threshold of <5 x 10\(^{-8}\) to indicate genome-wide statistical significance in the discovery meta-analysis for the selection of SNPs taken forward to the next stage. As no SNP reached genome-wide significance in the analysis of diastolic function traits, SNPs with P < 1 x 10\(^{-6}\) were taken forward for replication as “suggestive” findings. This threshold was chosen because there was approximately 80% power to achieve a genome-wide significant P value in the combined discovery and replication analysis for most of the traits given the effect sizes observed in the discovery stage. The association results were grouped based on the LD structure from the Hap-
cis eQTL analysis
To evaluate the potential functional significance of our findings, we related each replicated SNP to the expression levels of genes in three sets of tissues: human whole blood samples from \( n = 5,311 \) individuals evaluated by Westra et al. (49), human monocytes from \( n = 1,372 \) participants in the GHS (50), and LV free-wall tissue from \( n = 313 \) patients with HF undergoing transplantation and from unused donor hearts from the MAGNet consortium (http://www.med.upenn.edu/magnet). Further details are presented in Supplemental Methods. To evaluate possible cis eQTLs across multiple tissues, an additional look-up was performed in the GTEx database for the new findings.

Pathway analysis
The collective effects of multiple genetic variants on biological systems were investigated by pathway analysis, first for the 7 structural and systolic traits combined, and then for the 9 combined diastolic traits and for all 16 echocardiographic traits combined (for details, see Supplemental Methods).

To identify whether any of the associated SNPs fall within regulatory regions of the genome, we evaluated data from ENCODE (17). We compared the expected overlap of the putative SNPs with functional domains due to chance with the actual observed overlap by creating a permuted set of non-associated SNPs that were evaluated for overlap with the functional domains. We also used the DEPICT tool to further explore functionality of the identified SNPs (19). In addition, variants with a 2-tailed \( P < 5 \times 10^{-8} \) were used as the input for the DAPPLE software (18), which then built both direct and indirect interaction networks from seed genes near the top loci.

Statistics
If not specified otherwise, a Wald test statistic was calculated by dividing the estimated effect size by its standard error and comparing them with a normal distribution (2-tailed) with mean zero. In the GWAS, \( P < 5 \times 10^{-8} \) for the combined stage 1 and 2 analysis was deemed significant (11), which corresponds to a significance level of 0.05 after correcting for 1 million independent SNPs (51). For pathway analyses, a FDR was applied as multiple testing correction with a cutoff-value correcting for 16 hypothesized pathways.

Study approval
All study protocols of participating cohorts were reviewed and approved by a local ethics committee and followed the recommendations of the Declaration of Helsinki. All subjects in the cohorts provided informed written consent prior to their participation in the study. Therefore, no specific approval was required for this meta-analysis of human data. The institutional review boards of all study protocols approved the study-specific Acknowledgments, funding, and ethics statements.

Author contributions

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Study-specific Acknowledgments, funding, and ethics statements
The following list provides the study-specific names and funding sources with grant numbers only. Details on the study acronyms, study-specific Acknowledgments, funding, support for researchers, and ethics statements are provided in the supplemental material.

AortaGen
AGES. NIH N01-AG-1-2100, National Institute on Aging (NIA) Intramural Research Program, Hjartavérsd (the Icelandic Heart Association), Althingi (the Icelandic Parliament).

ASCOT. Pfizer, New York; Servier Research Group, Paris, and Leo Laboratories, Copenhagen; partial funding: Barts and the London School of Medicine and Dentistry, Centre Nationale de Genotypage Paris, and Irish Research Council GREP award.
ASPS. Austrian Science Fund Project P20545-P05 Genetics of Cerebral Small Vessel Disease.

CARDIA. National Heart, Lung, and Blood Institute in collaboration with the University of Alabama at Birmingham (HHSN268201300025C and HHSN268201300026C), Northwestern University (HHSN268201300027C), University of Minnesota (HHSN268201300028C), Kaiser Foundation Research Institute (HHSN268201300029C), and Johns Hopkins University School of Medicine (HHSN26820000004IC); partial funding: Intramural Research Program of the NIA, Gene Environment Association Studies (GENEVA) through grants U01-HG004729, U01-HG04424, and U01-HG004446 from the National Human Genome Research Institute.

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CHARGE-HF

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Generation R. Erasmus Medical Center, Rotterdam, Erasmus University Rotterdam, Netherlands Organization for Health Research and Development (ZonMw), Netherlands Organisation for Scientific Research (NWO), Ministry of Health, Welfare and Sport and the Ministry of Youth and Families.


HyperGEN. HyperGEN echocardiography ancillary study: NIH (R01 HL 55673). HyperGEN parent study: Cooperative agreements (U10) with NHLBI: HL54471, HL54472, HL54473, HL54495, HL54496, HL54497, HL54509, and HL54515.


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