Kallmann syndrome is an inherited deficiency of gonadotropin-releasing hormone (GnRH) that is characterized by hypogonadism with delayed or absent puberty and dysfunctional olfaction. While Kallmann syndrome–associated mutations have been identified in some sets of patients, for many of these individuals, the underlying cause remains unknown. In this issue of the JCI, Cariboni and colleagues identify mutations in semaphorin 3E (SEMA3E) in two brothers with Kallmann syndrome. In animal models, loss of SEMA3E signaling recapitulated phenotypes of the probands and resulted in enhanced GnRH neuron death during development. The results of this study offer important insight into the development of Kallmann syndrome and provide tools for elucidating mutations that underlie complex hormonal phenotypes.

The hypothalamic/pituitary/gonadal axis

Function of the hypothalamic/pituitary/target gland axis is tightly regulated by three interacting tiers of control that integrate brain, anterior pituitary, and peripheral hormone signaling. The anterior pituitary gland comprises at least six highly differentiated cell types that each express a specific trophic hormone. Corticotroph cells express adrenocorticotropin (ACTH), which regulates adrenal function; somatotrophs express growth hormone (GH), which regulates growth and body composition; thyrotrophs express thyrotrphin (TSH), which regulates thyroid function; lactotrophs express prolactin (PRL), which regulates mammary gland development and lactation; and gonadotrophs express both lutropin and gonadotropin, insulin-like growth factor (IGF-I), and thyroid hormon, also participate in feedback loops to regulate their respective pitutary trophic hormone secretion (1).

Gonadal function is mediated by the maturation and function of germ cells as well as sex steroid-dependent regulation of secondary sexual characteristics (2). During development, central GnRH neurons arise mainly from the olfactory placode and migrate to the hypothalamus, as determined by regulated neuronal and glial scaffolding as well as chemotactic and adhesion molecules (refs. 3, 4, and Figure 1).

The hypothalamic gene GNRH1 encodes a 92-aa precursor protein that yields a 10-aa mature GnRH molecule, which binds the pituitary GnRH receptor (GnRHR) (5). Generation of hypothalamic GnRH pulses is coordinated by a hypothalamic neural network, which consists of neurons projecting to the median eminence and terminating in the hypophysial portal-capillary network. Coordinated and repetitive GnRH secretion patterns determine gonadal functions through an exquisite regulation of pitutary FSH and LH release, pulsatility, and hormone glycosylation (6).

Hypogonadotrophic hypogonadism

Disruption of the hypothalamic/pituitary gonadal axis at any level proximal to the gonads may lead to the reproductive and developmental phenotype of hypogonadotropic hypogonadism, which is characterized by gonadal failure accompanied by attenuated levels of FSH and LH (7). Kallmann syndrome (8), which is now known to be caused by defective olfactory bulb and GnRH neuron development (9), is associated with varying degrees of anosmia/hyposmia. In contrast, hypogonadotropic hypogonadism, caused by disruption of GnRH secretion or action, is usually associated with normosmia and hypogonadism (2). Although acquired causes of the disorder are well documented, molecular elucidation of a genetic etiology of hypogonadotropic hypogonadism is now possible for a large number of these patients (10). In this issue, Cariboni et al. provide insight into how a developmental gene mutation disrupts the development or migration of central GnRH neurons through their identification and characterization of a mutation in semaphorin 3E (SEMA3E) (11). Discovery of this gene mutation elucidates a developmental cause of hypogonadotropic hypogonadism with anosmia. Previous studies have identified mutations in genes encoding several hypothalamic regulatory factors, including hypothalamic kisspeptin and neurenikins, that disrupt GnRH synthesis.
or secretion (12). GnRH secretion is also regulated by nutrition-related hormones, including leptin and ghrelin (13). GnRH action may be abrogated by mutations in the GnRHR receptor, leading to trafficking or signaling defects. As GnRHR signaling specificity is strongly determined by conformational integrity, mutations that lead to receptor misfolding may disrupt hypothalamic signal transduction to the gonadotroph cell (14). Reinforcing this observation, defective GnRHR stability may be rescued by small-molecule chaperones (15). Within the gonadotroph, the distal component of the hypothalamic/pituitary cascade, LH and FSH synthesis, may be disrupted as the result of mutations in genes encoding pituitary hormones or gonadotroph-specific transcription factors. Acquired hypogonadotrophism may also be caused by sellar masses compressing gonadotrophs, or by structural damage after surgery, radiation, or pituitary inflammation (1).

**Mechanisms underlying Kallmann syndrome**

Kallmann syndrome is an X-linked disorder that results from disruption of hypothalamic GnRH neuron development and presents with associated olfactory nerve agenesis or hypoplasia, leading to hypomia/anosmia (16). The syndrome comprises varying degrees of hypogonadism with attenuated pituitary FSH and LH secretion, as well as heterogenous features that are linked to specific mutations but are not associated with the reproductive system. For example, synkinesia is associated with Kallmann syndrome 1 (KAL1, which encodes anosmin) mutations; hearing loss is linked to chromodomain helicase DNA-binding protein 7 (CHD7) mutations; and dental agenesis and syndactyly are associated with mutations in FGF (10, 16).

Other clinical features of both autosomal dominant and recessive forms of Kallmann syndrome include color blindness, renal agenesis, and cryptorchidism (16, 17). MR imaging of individuals with Kallmann syndrome may also reveal hypoplasia in the olfactory tract. As hypogonadotropism is present throughout the lifespan of those with Kallmann syndrome, developing and mature tissues are deprived of gonadal steroid exposure, leading to hypogonadism with delayed puberty and attenuated secondary sexual development, including continued bone growth and primary amenorrhea in females, and micropenis in males (18). Moreover, these patients are devoid of hypothalamic GnRH and exhibit attenuated synthesis and secretion of LH and FSH by pituitary gonadotrophs, resulting in low or undetectable estradiol or testosterone secretion and atrophic gonads.

**Figure 1. The cascade of reproductive axis signal amplification initiated by GnRH and culminating in gonadal function confers fertility and secondary sex characteristics.** Central molecules, including KAL1, FGF8/FGFR1, PROK2/PROK2R, NELF, SEMA3A/PLXND1, HS6ST1, CHD7, and WDR11, determine GnRH neuron development in the olfactory region (27) and migration to the hypothalamus. Multiple intrahypothalamic, nutritional, and hormonal signals, such as KISS1/KISS1R, TAC3/TAC3R, and PCSK1, determine the generation of age- and time-determined GnRH pulses, which traverse the hypophyseal portal circulation to signal through the pituitary gonadotroph GnRHR. Transcription factors, including NR5A1, NR0B5, SF1, DAX1, and PROP1, also determine cell-specific gonadotropin hormone (FSH and LH) expression. Unique pulsatile FSH and LH release patterns induce gonadal hormones and determine germ cell function. Hypogonadotrophic hypogonadism can be caused by disruptions at each of the depicted CNS/hypothalamic/pituitary levels. Heterogenous oligogenic patterns of inheritance may be associated with a similar clinical phenotype of gonadal failure in the presence of inappropriately low gonadotropin hormone levels. Acquired causes of hypogonadotrophic hypogonadism include neoplastic, inflammatory, or iatrogenic (surgery, radiation) hypothalamic-pituitary damage.
Several factors have been identified that determine the development of olfactory placode cells in the developing brain and their migration toward the hypothalamus. Complex signaling enables GnRH precursor cells to navigate the frontal lobe, where they ultimately reside in the hypothalamic median eminence. Kallmann syndrome–linked mutations have been ascribed to dysfunctional GnRH neuron development, migration, survival, and differentiation. Anosmin is required for olfactory cell migration and adhesion (9), and loss of anosmin disrupts GnRH cell migration from the olfactory placode; however, this X-linked defect accounts for a only small cohort of these patients (2). Affected kindreds have also been shown to include individuals who manifest with autosomal dominant or recessive traits with variable penetrance. Heterogeneous combinations of mutations result in an oligogenic inheritance pattern and often include the gene encoding FGFR1 ligand, which regulates axonal development (19). Several gene products interact with the FGF network, including the chromatin-remodeling factor CHD7 (20). Mutations in CHD7 are a component of CHARGE syndrome, which includes coloboma, choanal atresia, and heart defects, along with growth and development dysfunction (21). Mutations of the gene encoding SEMA3A, which functions as an axonal pathway regulator of GnRH neurons, result in ectopic GnRH nasal neuron migration (22). Other factors, including neuronal migration factor NELF, WD repeat domain 11 (WDR11), and heparan sulfate 6-O-sulfotransferase 1 (HS6ST1), also participate in GnRH development or migration (18), and olfactory bulb development is regulated by prokineticin 2 (PROK2) (23).

Despite remarkable advances in identifying Kallmann syndrome–associated gene mutations, it is apparent that hitherto unidentified gene mutations underlie this syndrome in many patients with idiopathic phenotypes (24). In this issue, Cariboni et al. demonstrate that SEMA3E and the SEMA3E receptor PLXND1 are important factors for GnRH neuron survival in the developing brain after these cells have traversed the forebrain during hypothalamic migration. Using whole-exome sequencing and computational modeling, Cariboni and colleagues identify a SEMA3E mutation in two brothers with features of Kallmann syndrome (11). This particular missense mutation (R619C) enables selective death and apoptosis of central GnRH-producing neurons. SEMA3E, in conjunction with PLXND1, acts as a cell repellent and angiogenesis regulator (25) and is now shown by Cariboni et al. to function as a prosurvival factor for mature hypothalamic GnRH neurons by signaling through PLXND1 and its coreceptor KDR, a kinase known to mediate neuronal growth. Whole-exome sequencing also revealed the presence of a CHD7 mutation in the two probands, indicating that the combination of heterozygous mutations underlies the disease, as fertility was enabled in both parents, each of whom harbored a heterozygous mutation. The presence of combined heterozygous CHD7 and SEMA3E mutations in the offspring afflicted with Kallmann syndrome is consistent with the proposed oligogenic pattern of Kallmann syndrome inheritance (26). Cariboni and colleagues also recapitulated the human phenotype in compound heterozygous mice that were null for both Plxnd1 and Chd7. GnRH neuron innervation in the median eminence was severely disrupted in these animals, further demonstrating the cooperative functions of SEMA3E and CHD7 in regulating GnRH neuron development (11).

SEMA3E was also shown to induce PLXND1-dependent PI3K-mediated GnRH neuron survival, as GnRH neuron apoptosis was increased and GnRH neuron innervation was decreased in the developing brains of KO mice lacking either SEMA3E or PLXND1. This reduced adult median eminence population of GnRH-positive neurons recapitulated clinical features of hypogonadism, such as testicular atrophy. Accordingly, PLXND1 and CHD7 were proposed to synergistically regulate GnRH neuron development. As mechanisms underlying these observations likely do not involve indirect forebrain vascular effects nor reflect defective SEMA3E dimerization, Cariboni and colleagues propose that introduction of an extracellular cysteine may lead to disrupted protein structure (11).

Conclusions and future directions
The results of the study by Cariboni et al. (11) support adding SEMA3E and PLXND1 to the list of candidate genes to be considered when evaluating patients with hypogonadotropic hypogonadism associated with hyposmia/anosmia. For patients with other as-yet idiopathic endocrine disorders, the approach of Cariboni and colleagues, which involved a combination of whole-exome sequencing, computational modeling, functional cell biology, and clinical recapitulation in KO transgenic mice, should lead to the discovery of additional genetic causes of complex, rare familial endocrine syndromes. Thus, the elegant discovery of Cariboni et al. sheds light on an investigational approach for further detection of as-yet cryptic causes of other central pituitary hormone deficiency syndromes. For example, rare syndromes of isolated ACTH, TSH, or GH deficiency with unclear genetic causes could, in fact, be ascribed to defective hypothalamic CRH, GHRH, or TRH neuron development or function, respectively. The identification of these defects is a daunting task, as these syndromes are usually rare, making sporadic mutations difficult to identify and convincingly demonstrate to be the underlying cause of dysfunction. Moreover, these syndromes could likely turn out to be oligogenic disorders, as was the case for the individuals with Kallmann syndrome characterized by Cariboni and colleagues. Nevertheless, the successful application of novel techniques to unravel complex hormonal phenotypes exemplified by Cariboni et al. is encouraging for the elucidation of heretofore enigmatic molecular causes of central reproductive, adrenal, thyroid, and growth failure.

Acknowledgments
The author is grateful to Anat Ben-Shlomo, Ning-Ai Liu, and P. Michael Conn for critical reading of the manuscript and to Grace Labrado for manuscript preparation. Work in the author’s laboratory is supported by NIH grant DK 103198 and the Doris Factor Molecular Endocrinology Laboratory.

Address correspondence to: Shlomo Melmed, Cedars-Sinai Medical Center, 8700 Beverly Blvd., Room 2015, Los Angeles, California 90048, USA. Phone: 310.423.4691; E-mail: melmed@csmc.edu.


