Protein tyrosine phosphatase-σ regulates hematopoietic stem cell-repopulating capacity

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Introduction

RTKs regulate the maintenance, differentiation, and malignant transformation of hematopoietic stem cells (HSCs) (1–5). The activity of RTKs is counterbalanced through the action of receptor protein tyrosine phosphatases (PTPs), which dephosphorylate receptor and intracellular kinases (6, 7). The functions of certain intracellular PTPs, such as SHP2, in hematopoiesis are well characterized. SHP2 is required for the maintenance of HSCs and progenitor cells (8). Gain-of-function mutations in SHP2 cause a myeloproliferative disorder, and SHP2 is essential for oncogenic c-KIT transformation to myeloproliferative disease (9, 10). Recently, the intracytoplasmic phosphatase of regenerating liver PRL2 was found to be important for SCF-mediated HSC self renewal (11). In addition to the intracytoplasmic PTPs, there are 21 distinct receptor PTPs. However, the functions of receptor PTPs in hematopoiesis are not well understood (7).

We recently discovered the function of a heparin-binding growth factor, pleiotrophin (PTN), which is secreted by BM endothelial cells (ECs) and promotes the in vitro expansion of murine and human HSCs (12). PTN mediates HSC expansion via binding and inhibition of a receptor PTP, PTPζ (encoded by PTPRZ), on HSCs (12, 13). Deletion of Ptn caused a 10-fold reduction in HSC content in vivo, whereas deletion of Ptpz caused a significant expansion of HSCs in vivo (13). Based on these findings, we sought to determine whether other receptor PTPs might also be expressed by HSCs. We found that PTPs (encoded by PTPRS) is highly expressed in murine and human HSCs. Interestingly, BM cells from Ptpsr−/− mice displayed markedly increased competitive repopulating capacity compared with Ptpsr−/− BM cells. The increased functional capacity of Ptpsr−/− HSCs was associated with increased activation of the RhoGTPase RAC1 (14, 15), and inhibition of RAC1 blocked the augmented migration capacity of Ptpsr−/− cells. Furthermore, negative selection of human cord blood (CB) HSCs for PTPσ− cells caused a 15-fold increase in repopulating capacity compared with human PTPσ− HSCs. These data reveal a role for PTPσ in regulating HSC function and suggest that PTPσ inhibition or negative selection for PTPσ can increase HSC repopulation in vivo.

Results and Discussion

We sought to determine the relative expression of receptor PTPs in murine HSCs. Cd45, Ptprs, and Ptpre were expressed at more than 100-fold higher levels in BM ckit−sca-1−lin− (KSL) stem/progenitor cells compared with other receptor PTPs, including Ptpre (Fig-
Since PTPσ has been implicated in regulating the regeneration of neural stem cells (16, 17), we hypothesized that PTPσ might also regulate HSC function. Ptprs expression was increased significantly in HSCs compared with more mature hematopoietic cell populations (Figure 1A). In order to determine whether PTPσ had a functional role in regulating HSC fate, we compared the hematopoietic phenotype and function of Ptprs–/- mice and Ptprs+/- mice (18). Ptpsr–/- mice were viable, and we confirmed decreased PTPσ expression in BM lin- cells from Ptpsr–/- mice (Supplemental Figure 1; supplemental material available online with this article; doi:10.1172/JCI77866DS1). Adult Ptpsr–/- mice had normal peripheral blood (PB) counts and no alterations in total BM cells, KSL cells, SLAM+KSL HSCs, HSC cell-cycle status, or apoptosis compared with Ptpsr+/- mice (Supplemental Figure 1). However, Ptpsr+/- mice contained significantly increased myeloid colony-forming cells (CFCs) compared with Ptpsr–/- mice (Figure 1B). Furthermore, mice that were competitively transplanted with limiting doses of BM cells from Ptpsr–/- mice had 8-fold increased donor CD45.2+ hematopoietic cell engraftment at 16 weeks compared with mice transplanted with the identical cell dose from Ptpsr+/+ mice (Figure 1C). Restoration of myeloid, B cell, and T cell lineages was also significantly increased in mice transplanted with Ptpsr–/- BM cells compared with recipients of Ptpsr+/+ cells (Figure 1, C and D). Secondary competitive transplantation assays demonstrated that Ptpsr–/- donor BM cells contained significantly increased long-term HSC function compared with BM cells from Ptpsr+/- mice (Figure 1E). Of note, we observed no differences in the homing capacity of donor BM cells from Ptpsr–/- mice versus Ptpsr+/- mice (Supplemental Figure 1).

Figure 1. Deletion of Ptpsr augments HSC-repopulating capacity. (A) Mean expression of receptor PTPs in BM KSL cells by quantitative reverse-transcriptase PCR (qRT-PCR) (left) and expression of Ptpsr within hematopoietic cell subsets (right) are shown. n = 3–9/group. *P < 0.0001 for each of the 3 comparisons. (B) Mean (± SEM) numbers of CFCs are shown for 12-week-old Ptpsr–/- and Ptpsr+/- mice. *P = 0.002 (n = 6, Mann-Whitney U test). CFU-GEMM, CFU–granulocyte erythroid monocyte megakaryocyte; BFU-E, burst-forming unit–erythroid; CFU-GM, CFU–granulocyte macrophage. (C) Mean levels of donor CD45.2+ hematopoietic cell engraftment are shown in the PB of CD45.1+ mice at 16 weeks following competitive transplantation of 3 × 10⁴ BM cells from Ptpsr–/- or Ptpsr+/- mice. *P < 0.0001 (n = 15–18/group, Mann-Whitney U test). Multilineage engraftment of Mac-1/Gr-1+, B220+, and CD3+ donor cells is shown at right. **P = 0.008; †P = 0.0001; ‡P = 0.04 (Mann-Whitney U test). (D) Multilineage flow cytometric analysis of donor hematopoietic cell engraftment in the PB is shown from mice competitively transplanted with BM cells from Ptpsr–/- or Ptpsr+/- mice at 16 weeks after transplant. Quadrant numbers represent the percentages of donor lineage cells. (E) In the upper panel, mean donor CD45.2+ cell engraftment in the PB is shown over time following transplantation of BM cells from Ptpsr–/- or Ptpsr+/- mice in primary recipient mice. *P < 0.0001; **P < 0.0001; †P = 0.001; and ‡P < 0.0001 for engraftment at 4, 8, 12, and 16 weeks, respectively. In the lower panel, mean donor CD45.2+ cell engraftment in secondary transplanted mice is shown over time. *P = 0.004; **P = 0.01; †P = 0.005; and ‡P = 0.002 for engraftment at 4, 8, 12, and 16 weeks, respectively (n = 7–8/group, Mann-Whitney U test).
lial migration capacity and cobblestone area–forming cell (CAFC) content of BM cells compared with control BM cells (15). We found that Ptprs–/– BM cells had 4-fold increased numbers of 5-week CAFCs compared with Ptprs+/+ BM cells (Figure 2C). Furthermore, Ptprs–/– BM cells displayed significantly increased transendothelial cell migration capacity compared with Ptprs+/+ BM cells (Figure 2D). Treatment of Ptprs–/– BM cells with EHT1864, a Rac inhibitor, completely abrogated the enhanced transendothelial migration capacity of Ptprs–/– cells (Figure 2D). These data suggest that PTPσ inhibits Rac1 activation in BM HSCs and that the increased HSC engraftment capacity of Ptprs–/– BM cells is dependent, at least in part, on Rac1 activation.

Since deletion of Ptprs increased murine HSC-repopulating capacity, we sought to determine whether the negative selection of human HSCs for PTPσ expression could enrich for HSCs with enhanced repopulating capacity. PTPRS was expressed by a

Figure 2. PTPσ regulates Rac1 activation in HSCs, and Rac1 inhibition abrogates the Ptprs–/– BM cell migration capacity. (A) At left, flow cytometric analysis of Rac1-GTP levels in BM KSL cells from Ptprs+/+ and Ptprs–/– mice is shown. Numbers represent the percentages of Rac1-GTP+ cells. At right, mean percentages of Rac1-GTP+ KSL cells are shown in Ptprs+/+ and Ptprs–/– mice. *P = 0.008 (n = 3, t test). (B) At left, flow cytometric analysis of Rac1-GTP levels in wild-type BM KSL cells treated with scramble shRNA or PTPσ shRNA is shown. Numbers represent the percentages of Rac1-GTP+ cells. At right, scatter plot of percentage of Rac1-GTP+ KSL cells is shown in each group. Horizontal bars represent mean values. *P = 0.01 (n = 6, t test). (C) Poisson statistical analysis of a limiting dilution assay of 5-week CAFCs from Ptprs–/– versus Ptprs+/+ BM cells. The CAFC frequency for Ptprs–/– BM cells was 1 in 839 cells versus 1 in 3,801 cells for Ptprs+/+ BM cells (n = 10/group, P = 0.0001). (D) Mean numbers of CFUs are shown from the lower chambers of transendothelial migration assays containing Ptprs+/+ BM cells and Ptprs–/– BM cells, treated with and without EHT1864. *P < 0.0001 (n = 12, t test) for total CFUs; **P < 0.0001 for total CFUs (n = 6, t test).
mean of 49.9% of human CB CD34+CD38−lin− stem/progenitor cells (n = 6, Figure 3, A and B). We then performed transplantation assays into NOD/SCID−IL-2 receptor γ chain-null (NSG) mice to assess the repopulating capacity of CB HSCs selected for PTPσ expression. At 16 weeks after transplant, NSG mice transplanted with CD34+CD38−CD45RA−lin−PTPσ− cells displayed 15-fold higher engraftment compared with mice transplanted with parent CD34+CD38−CD45RA−lin− cells and more than 15-fold higher compared with mice transplanted with CD34+CD38−CD45RA−lin−PTPσ+ cells (Figure 3C). NSG mice transplanted with CD34+CD38−CD45RA−lin−PTPσ− cells had significantly increased engraftment of donor myeloid cells, B cells, and T cells compared with mice transplanted with CD34+CD38−CD45RA−lin−PTPσ+ cells (Figure 3C, D and E). Temporally, the engraftment of PTPσ− CB cells significantly increased between 8 and 16 weeks compared with that of parent CB cells or PTPσ+ CB cells (Figure 3E). Of note, CD34+CD38−CD45RA−lin− PTPσ− cells displayed no difference in cell-cycle status compared with CD34+CD38−CD45RA−lin− cells or CD34+CD38−CD45RA−lin−PTPσ+ cells (Supplemental Figure 3). Surface expression of CXC chemokine receptor type 4 (CXCR4), which regulates HSC homing and retention in the BM microenvironment (21, 22), was not different between CD34+CD38−CD45RA−lin− cells and CD34+CD38−CD45RA−lin−PTPσ− cells, but both populations had higher CXCR4 expression compared with CD34+CD38−CD45RA−lin−PTPσ+ cells (Supplemental Figure 3). We found no differences in CXCR expression between BM KSL cells from Ptprs−/− mice and Ptprs+/+ mice (mean 2.7% CXCR4+ vs. 3.2%, respectively, n = 6).

Our findings reveal several interesting aspects of PTPσ function in hematopoiesis. First, PTPσ negatively regulates HSC engraftment and self renewal in vivo following competitive transplantation. Our findings in hematopoiesis are analogous to the putative role of PTPσ in nerve regeneration, in which PTPσ mediates chondroitin sulfate proteoglycan–driven (CSPG–driven) inhibition of nerve regeneration following spinal cord injury (23–27).
ervative challenge to HSCs. Our results suggest that PTPσ inhibits HSC regeneration in vivo, perhaps via interaction with CSPGs, which are abundant in the extracellular matrix of the BM (28).

Going forward, it will be important to dissect the precise cellular mechanism through which PTPσ regulates HSC repopulation. We have established that PTPσ regulates RAC1 activation and that RAC activation is responsible for at least some of the augmented function of Ptprs+/− HSCs. RAC proteins regulate several HSC functions, including chemotraction, homing, proliferation, survival, and endosteal localization (29, 30). Since we found no alterations in HSC apoptosis, cell-cycle status, or homing capacity of PTPσ-deficient HSCs, we propose that the PTPσ-RAC1 axis may regulate HSC localization or “lodgment” in the niche (15, 31). We plan to directly visualize the localization of transplanted Ptprs+/− and Ptprs−/− progenitor cells in BM niches in vivo utilizing cell-labeling techniques (13) and will interrogate the effect of RAC inhibition on the HSC lodgment process. We will also investigate the role of CXCR4 in this process, since CXCR4 mediates signals via RhoGTPases and RAC1 regulates CXCR4 conformation and function in hematopoietic cells (29, 32).

Translationally, we have shown that the negative selection of human CB CD34+CD38−CD45RA+lin− cells for PTPσ surface expression enriches for human long-term HSCs by approximately 15-fold. This observation has fundamental implications, since the molecular characterization of human HSCs may be improved by utilization of PTPσ to isolate more purified HSCs (33). Surface expression of CD90 (Thyl) has also been utilized to enrich for human HSCs (33, 34). The most effective purification strategy for human HSCs described to date utilized the expression of CD49f (integrin α6) such that a subset of NSG mice transplanted with single CD34+CD38−CD45RA+lin−RholoCD49f+ cells demonstrated multilineage hematopoietic engraftment (33). It is noteworthy that CD49f+ and PTPσ+ are both receptors for the extracellular matrix glycoproteins laminin and chondroitin sulfate/heparin sulfate proteoglycans, respectively. This shared feature suggests that proteoglycan-mediated signaling in the BM microenvironment regulates HSC repopulation in a context-specific manner under the control of integrin- and PTPσ-mediated signaling. Practically, we have provided a method to isolate human HSCs for therapeutic objectives such as gene therapy and allogeneic transplantation. Our results also provide the mechanistic basis for the systemic administration of PTPσ inhibitors (35) as a means to accelerate hematopoietic reconstitution in settings such as adult CB transplantation, in which delayed hematopoietic engraftment remains a major clinical problem (36).

Methods

For more detailed information, see the Supplemental Methods.

Animals. Mice bearing constitutive deletion of Ptprs in a Balb/c background were provided by Michel Tremblay (McGill University, Montreal, Quebec, Canada). Cby.SJL(B6)-Ptprc−/−/J (CD45.1 Balb/c) and NOD.Cg-Ptkr1<sup>cre</sup>/Il2rg<sup>tm10</sup>/Szj (NSG) mice (Jackson Laboratory) were also utilized.

Statistics. All data are shown as mean ± SEM. We used the Mann-Whitney U test (2-tailed nonparametric analysis) and the 2-tailed Student’s t test for the comparisons shown. P < 0.05 was considered significant.

Study approval. Animal procedures were performed under protocols approved by Duke University and UCLA animal care and use committees.

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