Spinal muscular atrophy (SMA) remains one of the most common and lethal autosomal recessive diseases. Homozygous deletion of survival of motor neuron 1 (SMN1) and resulting SMN protein deficiency manifests predominantly with motor neuron degeneration; however, a wealth of emerging data supports a broader influence of SMN deficiency in disease pathogenesis. In this issue of the JCI, Kariya and colleagues demonstrate the relatively selective impact of SMN depletion on the distal motor unit using a series of SMN2-expressing transgenic mice in which constitutive SMN knockdown results in a truncated, less stable protein. Historically relating to obvious defects in neuromuscular junction (NMJ) maturation, decreased synapse formation to obvious defects in neuromuscular junction (NMJ) maturation, decreased synapse formation.

SMN-targeted therapeutics: the hope and the hype

Spinal muscular atrophy (SMA) is an autosomal recessive disorder characterized by motor neuron loss in the spinal cord and brainstem. Progressive muscular weakness and atrophy typically emerge in infancy or early childhood after a variable period of normal development. Deficiency in survival motor neuron (SMN) is associated with SMA. In humans and bonobos, a unique inverted duplication on chromosome 5 encompasses SMN1 and its nearly identical homolog, SMN2; however, lower vertebrates, including mice, lack SMN2. Deletion and/or mutation of SMN1 is disease causing, and a translationally silent nucleotide substitution in SMN2 results in a truncated, less stable protein. Historically, low SMN was thought to selectively target motor neurons; however, emerging data from SMA mouse models indicate that low levels of SMN affect many tissues, including components of the neuromuscular circuitry, and skeletal muscle (1). In addition to obvious defects in neuromuscular junction (NMJ) maturation, decreased SMN results in early abnormalities in synaptic input from muscle fibers to motor neurons within the spinal cord (2, 3). Both SMA mouse models and severely affected human infants exhibit abnormalities in myofiber maturation, muscle size, and muscle function (4–6). Furthermore, there is evidence that SMN-intrinsic defects in Schwann cells promote SMA pathogenesis, and peripheral nerve myelination is abnormal in severely affected infants (7). In humans lacking SMN production from SMN1, SMN2 phenotypically modifies...
SMA severity, with SMN2 dosage inversely correlating with disease severity. In mice, Smn deletion results in embryonic lethality; however, introduction of multiple copies of SMN2 into Smn−/− mice ameliorates disease phenotypes (8). Because SMN2 is universally present in SMA patients, it is a compelling therapeutic target for SMA-associated neurodegenerative disease. SMN-targeted therapies are already being evaluated in clinical trials, increasing the urgency for understanding when and where SMN is required and the best delivery mode for effective SMN-mediated prevention, stabilization, or reversal of motor neuron degeneration. Accumulating evidence suggests that abnormalities of the motor neuron cell body and its central synapses, the axon, the NMJ, and skeletal muscle contribute synergistically to SMA pathogenesis and that SMN repletion within both central and peripheral nervous systems may be required for beneficial therapeutic outcomes (9, 10).

**The ubiquitous SMN protein: here, there, and everywhere**

All cells require SMN for biogenesis of snRNP particles and pre-mRNA splicing. If there is a universal requirement for SMN, why are motor neurons selectively affected by its loss, and where does SMA first manifest? Does SMN dysfunction begin at the level of the motor neuron cell body, the axon, or the NMJ (11, 12), and what effect does reduced SMN have on other components of the neuromuscular circuitry (13)? In zebrafish, SMN is required in motor neurons for normal growth and connectivity (14); however, SMA mice, which die within two to three weeks postnatally, display normal motor neuron connectivity at birth and only minimal abnormalities in distal NMJs during prenatal and neonatal development (15, 16). Several groups have also demonstrated that early postnatal induction of SMN is critical for prolonged survival in transgenic SMA mouse models (17, 18). In order to address the precise temporal requirements for SMN in postnatal development, Kariya and colleagues developed mouse models in which SMN knockdown could be temporally induced (19). By generating a series of transgenic mice with variable SMN2 copy number and inducible SMN knockdown, Kariya et al. were able to evaluate the impact of SMN depletion both constitutively and at multiple time points in postnatal development. Kariya and colleagues determined that prior to P15, motor unit development is vulnerable to abrupt SMN knockdown in the presence of two SMN2 copies; however, mature (after P50) mice with two copies of SMN2 were not markedly affected by SMN knockdown (19). Interestingly, as noted by
Kariya et al., the emergence of this relative resistance to SMN depletion at P15 in the mouse correlates morphologically with maturation of the distal NMJs (19).

Back to reality
Unfortunately, the most frequently inherited genotype in humans, homozygous SMN1 deletion with two SMN2 copies, is associated with the severe infantile phenotype (SMA type I), which accounts for about half of all newly diagnosed SMA patients each year. There are limited data regarding reversibility of SMA phenotypes in these often severely weak infants, who are currently diagnosed at a mean age of approximately 4.9 months (Families of Spinal Muscular Atrophy [FSMA] registry data from 2012; Jill Jarecki, personal communication). Compared with neonatal SMA mice, infants with SMA type I have more precipitous and severe denervation, and autopsies of infants that have died soon after their diagnosis demonstrate severe motor neuron loss within the spinal cord (C.J. Sumner and K.J. Swoboda, unpublished data).

While the critical SMN-sensitive period identified by Kariya et al. encompasses the period of maturation of the distal motor unit, it also spans a period of tremendous growth and developmental change (19). The translation of observations in mice to human disease remains a major challenge for scientists and clinicians trying to reach conclusions about the relevance of any preclinical observations, even those obtained under the most rigorously controlled experimental conditions. Despite these challenges, interesting parallels between mouse SMA models and human infants with SMA should be taken under consideration. The critical period in mice for distal NMJ maturation corresponds to the onset of weaning, as animals transition from suckling to mastication. Mice demonstrate tremendous growth during the first few weeks after birth, increasing their body weight seven- to eight-fold. Such rapid growth requires the motor unit to accommodate growth of the primary peripheral target, skeletal muscle, and to meet the needs of the motor neuron to manage axonal extension and distal NMJ remodeling. Like their murine counterparts, presymptomatic infants with SMA appear to have normal distal motor connectivity at birth.

Previously, we demonstrated the relationship among SMA type, SMN2 copy number, motor function, and denervation status in 89 SMA children enrolled in a natural history study at the University of Utah (20). Denervation status in a distally innervated hand muscle was studied using two electrophysiologic techniques: the maximum ulnar compound muscle action potential (CMAP) and a technique for motor unit number estimation (MUNE). Evaluation of CMAP and MUNE data from SMA type I infants presenting from birth to two years of age revealed a drastic denervation in these infants by two months of age (Figure 1, A and B, and ref. 20). Furthermore, we documented precipitous denervation within weeks to months after birth in a small subset of presymptomatic type I infants within this cohort (see Figure 4 of ref. 20). Since this publication, we have extended these observations to more than 80 type I and 150 type II SMA infants and children (K.J. Swoboda, unpublished data). SMA type I infants demonstrate an almost logarithmic decline in CMAP and MUNE values with age, reaching a nadir by four to six months, and associated with the development of generalized hypotonia and weakness. In contrast with infants with type I SMA, we observed that denervation in infants and children with SMA type II is much less acute and more variable, with the most active denervation occurring between six and 36 months of age (ref. 20 and Figure 1, C and D). Together, these observations support that human infants with a given SMN genotype demonstrate a vulnerability to disease progression during a relatively specific and critical period of development, similar to the observations in a murine model of SMA by Kariya and colleagues. This window of disease progression in SMA infants occurs during a period of rapid growth, peripheral neuromuscular development, and central nervous system development, as well as peripheral and central nervous system myelination. Additionally, in type I infants, this time period correlates with the transition from central pattern–generated reflex suckling to a more modulated pattern of nutritional intake, as infants advance from a liquid diet to a diet including solids.

What does this mean for clinical trials?
For years, clinicians have been stymied by the distinctive bimodal pattern of SMA–associated progression of neurodegeneration, which is perhaps most aptly characterized by Tom Crawford as “a slowing of the rate of degeneration with the passage of time” (21). Bimodal disease progression is uncommon in neurodegenerative disorders and makes an accurate prognosis following SMA diagnosis challenging. There is some overlap in the age of SMA onset in infants the severe form (type I) and those with the intermediate form (type II). While most SMA infants who present with weakness within the first year but are able sit unsupported will manifest an intermediate phenotype (type II), some will go on to ambulate independently (type IIIa; ref. 22).

Patterns of disease progression that vary by age and SMA subtype make clinical trial design challenging. Most children with SMA type II or III also present with an acute phase of denervation and clinical disease progression, albeit somewhat less obviously than those with SMA type I. Because the initial presentation of symptoms is followed by a period of stabilization of motor function that can last for months or even years, it is a challenge to accurately diagnose SMA subtype and provide meaningful prognosis. Additionally, SMA subtype, SMN genotype, modifying genetic factors, and environmental factors such as nutritional state or superimposed illness can all contribute to phenotypic heterogeneity. While an SMN2 copy number of two or less in the setting of symptom onset in early infancy remains a strong predictive determinant for developing SMA type I, identification of intrinsic and extrinsic factors that influence disease outcome has increased the urgency for better predictive and prognostic SMA biomarkers. Development of additional SMA biomarkers would allow for better characterization of SMA patient cohorts for early clinical trial enrollment, allowing for better assessment of SMA therapies.

Conclusions and future directions
Building upon previous studies, Kariya and colleagues demonstrate that SMN requirements are higher during early development through the completion of maturation of the distal NMJ and that acute depletion in adulthood results in minimal motor neuron dysfunction in mice with at least two SMN2 copies. Finally, the study by Kariya et al. demonstrates that SMN reduction after mice have reached maturity impairs reinnervation following peripheral nerve injury (19).

In their discussion, Kariya and colleagues boldly translate observations from SMA mouse models to human subjects, speculating that once effective SMN-targeted therapeutics are identified, they may be tapered
at a future time point. Given the major differences in time to maturation and life span between mice and humans, translating SMA-sensitive periods from mouse models to humans remains a major challenge. If SMA mouse models prove to accurately represent what might be expected in human clinical trials, then early or even presymptomatic intervention may be required to meet FDA efficacy standards. Because SMA type I infants present with generalized muscle weakness within six months of birth, newborn screening may be the only realistic means for preserving motor function in these patients. Patients with milder forms of SMA account for a substantial proportion of surviving patients and demonstrate a slower rate of denervation, providing a longer window of opportunity for therapeutic intervention. Due to the high stakes of therapeutics development, setting unrealistic efficacy goals could doom promising therapies that could benefit SMA patients with milder disease forms. For therapeutic trials targeting the most fragile patients, symptomatic type I infants, we must remain conscious that in some cases, a modest benefit simply is not enough.

While caution is warranted, the future is bright for SMN-targeted therapeutics, and the opportunity to dramatically improve or even cure SMA remains tantalizingly within reach. Given the continued emergence of data in both animal models and humans that SMN has the greatest impact on motor neurons during a critical developmental window, early intervention has never been more appealing. Heeding the lessons provided to us by the increasing wealth of data from a variety of experiments across diverse animal models will undoubtedly permit us to realize the greatest impact for our human patients.

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