RIP140 increases APC expression and controls intestinal homeostasis and tumorigenesis

Marion Lapierre,1 Sandrine Bonnet,1 Caroline Bascoul-Mollevi,2 Imade Ait-Arsa,1 Stéphan Jalaguier,1 Maguy Del Rio,1 Michela Plateroti,3 Paul Roepman,4 Marc Ychou,5 Julie Pannequin,6 Frédéric Hollande,6 Malcolm Parker,7 and Vincent Cavailles1

1IRCM, Institut de Recherche en Cancérologie de Montpellier, INSERM U896, Université Montpellier1, Montpellier, France. 2Institut régional du Cancer Montpellier, Unité de Biotéchnologie, Montpellier, France. 3Centre de Génétique et de Physiologie Moléculaire et Cellulaire, Université Lyon 1, CNRS, Villeurbanne, France. 4Agendia NV, Amsterdam, The Netherlands, and Agenda Inc., Irvine, California, USA. 5Institut régional du Cancer Montpellier, Service d’Oncologie Digestive, Montpellier, France. 6Centre National de la Recherche Scientifique (CNRS), Unité Mixte de Recherche (UMR) S203, Institut de Génomique Fonctionnelle, Montpellier, France. 7Institute of Reproductive and Developmental Biology, Faculty of Medicine, Imperial College, London, United Kingdom.

Deregulation of the Wnt/APC/β-catenin signaling pathway is an important consequence of tumor suppressor APC dysfunction. Genetic and molecular data have established that disruption of this pathway contributes to the development of colorectal cancer. Here, we demonstrate that the transcriptional coregulator RIP140 regulates intestinal homeostasis and tumorigenesis. Using Rip140-null mice and mice overexpressing human RIP140, we found that RIP140 inhibited intestinal epithelial cell proliferation and apoptosis. Interestingly, following whole-body irradiation, mice lacking RIP140 exhibited improved regenerative capacity in the intestine, while mice overexpressing RIP140 displayed reduced recovery. Enhanced RIP140 expression strongly repressed human colon cancer cell proliferation in vitro and after grafting onto nude mice. Moreover, in murine tissues and human cancer cells, RIP140 stimulated APC transcription and inhibited β-catenin activation and target gene expression. Finally, RIP140 mRNA and RIP140 protein levels were decreased in human colon cancers compared with those in normal mucosal tissue, and low levels of RIP140 expression in adenocarcinomas from patients correlated with poor prognosis. Together, these results support a tumor suppressor role for RIP140 in colon cancer.

Introduction

The Wnt pathway is one of the major pathways deregulated in colorectal cancer. In the physiologically normal gut, activation of this pathway ensures proliferation of precursor cells and renewal of the intestinal epithelium by activating the transcriptional properties of the T cell factor/lymphoid enhancer factor-1 (TCF/LEF1) family (1). Stimulation by Wnt ligands leads to stabilization of the transcription coactivator β-catenin, which becomes associated with TCF/LEF1 in the nucleus, leading to the expression of specific target genes. Canonical Wnt signaling operates by regulating the phosphorylation and degradation of β-catenin (2). Without stimulation by Wnt ligands, the levels of β-catenin in the cytoplasm are normally regulated by a multiprotein destruction complex that targets for degradation. This complex is assembled over the scaffold component axin, which contains binding domains for β-catenin, the tumor suppressor adenomatous polyposis coli (APC), and glycogen synthase kinase-3 (GSK3) and casein kinase 1 (CSNK1). Within the axin complex, β-catenin is sequentially phosphorylated by CSNK1 and GSK3 and then degraded by the proteasome (3). This complex thereby controls the proliferation of intestinal epithelial cells by maintaining the pool of active β-catenin. However, mutations of the APC gene, which were first identified in patients suffering from familial adenomatous polyposis (FAP), occur in a high proportion of sporadic colorectal carcinomas (up to 80%) (4). Stimulation of the Wnt pathway due to a mutation in the negative regulator APC provokes the hyperproliferation of the epithelium. Several mouse models have been generated, such as the ApcΔ14/+ mouse, in which exon 14 has been deleted (frameshift mutation at codon 580) and which presents a FAP phenotype with tumors in the distal colon and rectum, showing that heterozygous disruption of the Apc gene is associated with accumulation of β-catenin and overexpression of the β-catenin target genes cyclin D1 and c-Myc (5).

The transcription cofactor RIP140 (receptor-interacting protein of 140 kDa), also known as NRIIP1 (nuclear receptor-interacting protein 1), was first identified in human cancer cells through its interaction with estrogen receptor α (6). RIP140 was also shown to interact with many other nuclear receptors (NRs) and transcription factors (for a review see ref. 7). More recently, we demonstrated that RIP140 behaves as an Rb-like regulator of the E2F pathway by directly binding to E2Fs and repressing their transactivation potentials (8). RIP140 mainly acts as a transcriptional repressor by means of four inhibitory domains that recruit histone deacetylases or C-terminal binding proteins (9). Several post-translational modifications, such as sumoylation and acetylation, also play important roles in controlling the subcellular location and repressive activity of RIP140 (for a review see ref. 10). RIP140 is a ubiquitously expressed gene whose transcription is finely regulated at the transcriptional levels by both NRs and E2Fs (11). The physiological importance of RIP140 has been evaluated using mice...
Figure 1
RIP140 expression in mouse intestinal epithelium. (A) Immunofluorescence showing a gradient of RIP140 in the nuclei of epithelial cells along the villus/crypt axis of wild-type mice. Original magnification, ×20. (B) Real-time qPCR analysis of Rip140 mRNA in wild-type intestinal epithelial fractions. mRNAs encoded by the Lyz and Muc2 genes were used to verify the enrichment in villus- or crypt-associated cells. Data were normalized to RS9 mRNA. mRNA quantification for each gene is indicated in AU as the mean ± SD; n = 4 mice. (C) Left panel: H&E-stained transverse sections of small intestine from RIPKO, wild-type, and RIPTg mice. Right panel: The length of villi and crypts was measured on at least six fields selected from whole intestine sections of each mouse of the three different genotypes. Values represent the means ± SD; n = 6 mice for each genotype. Original magnification, ×10. Mann-Whitney U test. **P < 0.01; ***P < 0.001.
that lack the Rip140 gene (RIPKO mice). These animals are viable, but display a wide range of phenotypic alterations in various tissues and organs such as infertility of female mice (12) or reduced body fat content (13), and, more recently, severe cognitive impairments (14) and mammary gland morphogenesis (15).

Our present results demonstrate the role of RIP140 in homeostasis and tumorigenesis of the intestinal epithelium. We used mice with a loss or gain of RIP140 function to show that RIP140 inhibits cell proliferation and apoptosis in the intestinal epithelium. At the molecular level, RIP140 positively controls APC gene expression and, consequently, reduces β-catenin activation and Wnt target gene expression. Overexpression of RIP140 inhibits the proliferation of human colon cancer cells in vitro and in vivo after grafting onto nude mice. Finally, RIP140 mRNA and protein levels are reduced in colon cancer biopsies as compared with those in normal tissue, and patients whose tumors exhibit high RIP140 gene expression have the best survival rates. Altogether, this work identifies RIP140 as a key factor regulating intestinal tumorigenesis and as a potential new oncology biomarker.

Results

RIP140 expression in the intestinal epithelium. Previous data indicated that RIP140 is a ubiquitously expressed transcription factor (16). By quantitative real-time quantitative PCR (qPCR) analysis, Rip140 mRNA was detected in all the mouse tissues tested and particularly in the intestine and colon (Supplemental Figure 1A; supplemental material available online with this article; doi:10.1172/JCI65178DS1). First, we used immunofluorescence to analyze the distribution of RIP140 in the intestinal epithelium of wild-type mice and found that RIP140 was expressed in the nucleus of all epithelial intestinal cells, with a clearly increasing gradient along the crypt/villus axis (Figure 1A). To confirm this observation, we applied sequential isolation of wild-type mouse small intestine epithelial cells. To verify the enrichment of the different fractions in villus-associated or crypt-associated cells, we quantified the mRNA levels of the lysozyme (Lyz) and mucin 2 (Muc2) genes, which are, respectively, differentiation markers of Paneth cells located in the crypts and goblet cells in the villi (17). As expected, the Lyz gene was exclusively expressed in cells from fraction 4, whereas the Muc2 gene was mainly expressed in cells from fractions 1 and 2 (Figure 1B). Using the same enriched fractions, we found that the expression of Rip140 mRNA was significantly higher in differentiated cells of the villi than in cells from the crypts (Figure 1B).

RIP140 alters small intestine homeostasis. To study the role of RIP140 in the intestinal epithelium, we used Rip140-null (RIPKO) or Rip140 transgenic (RIPTg) mice. In these two mouse models, the respective deletion and overexpression of the Rip140 gene in intestinal sections were validated by immunofluorescence (Supplemental Figure 1B). As expected, RIP140 protein was undetectable in the RIPKO mice and overexpressed in the nuclei of all intestinal epithelial cells in the RIPTg mice as compared with their wild-type littermates. Real-time qPCR analysis confirmed that RIPKO mice expressed β-galactosidase as a marker of murine Rip140 gene disruption (12) and that the transgenic human RIP140 gene was specifically expressed in RIPTg mice (Supplemental Figure 1C).

Qualitative histopathological analysis of paraffin sections stained with hematoxylin and eosin (H&E) demonstrated no gross morphological changes in the structures of the intestinal epithelia of the two mouse models (Figure 1C, left panel). However, precise measurements of villus and crypt lengths demonstrated statistically significant differences between transgenic and wild-type animals (Figure 1C, right panel). Indeed, when compared with wild-type animals, the length of the crypt/villus axis increased in Rip140-null mice and decreased in mice overexpressing RIP140. This was associated with an altered total length of the small intestine, which was shorter in RIPKO mice and longer in RIPTg animals when compared with their respective wild-type littermates (Supplemental Figure 1D, top panel). As a consequence, invalidation or overexpression of the Rip140 gene did not affect the total surface area of the intestinal epithelium (Supplemental Figure 1D, bottom panel).

RIP140 inhibits cell proliferation and apoptosis. To determine whether cell proliferation was regulated by RIP140 expression, mice were injected with 5-ethyl-2′-deoxyuridine (EdU) 2 hours prior to sacrifice. As shown in Figure 2A, analysis of EdU labeling demonstrated that the number of cells synthesizing DNA was significantly increased (2.6-fold) in RIPKO mice as compared with that in wild-type animals. By contrast, the level of EdU staining decreased (3.7-fold) in RIPTg mice as compared with that seen in control mice. These results were strengthened by quantifying the mRNAs corresponding to Myc and proliferating cell nuclear antigen (PcnA) genes (Supplemental Figure 2A). The corresponding mRNA levels were increased up to 2-fold in RIPKO mice and were decreased by up to 3-fold in the RIPTg strain as compared with mRNA levels in the wild-type animals. The differences in PcnA gene expression among the three genotypes were confirmed by immunofluorescence labeling (Supplemental Figure 2B). We next assessed the effect of RIP140 expression on apoptosis by TUNEL assay on intestinal sections. The top of the villi showed a significant 3-fold increase in TUNEL-positive cells in Rip140-null mice as compared with that observed in their wild-type littermates, whereas villi overexpressing RIP140 (RIPTg) showed a clear 2-fold decrease in apoptosis (Figure 2B). Altogether, these observations suggest that RIP140 regulates both cell proliferation and apoptosis and may therefore inhibit the renewal of intestinal epithelium.

RIP140 inhibits the response of intestinal epithelium to irradiation. To highlight the role of RIP140 in the homeostasis of intestinal mucosa, RIPKO and RIPTg mice, together with their wild-type littermates, were exposed to a single 12-Gy dose of whole-body irradiation (WBI). On day 2.5, a time when recovery begins, we assessed the number and length of villi together with the length of crypts after H&E staining of intestinal paraffin sections (Figure 2C). Following WBI, we observed that Rip140-null mice exhibited a significantly increased number of regenerating villi as compared with that seen in wild-type animals. The length of villi and crypts was also significantly higher in RIPKO animals. As shown in Supplemental Figure 2C, this effect was associated with a strong rise in PCNA staining in the crypts, where progenitors and stem cells reside, and suggested a role for RIP140 in ionizing radiation-induced cell proliferation. As expected, this process was much slower in RIPTg mice (Figure 2C), which exhibited only a few buds of villi and weak PCNA staining of crypt cells 2.5 days after WBI (Supplemental Figure 2C). Altogether, these results strongly indicate that RIP140 inhibits renewal of the intestinal epithelium.

RIP140 controls β-catenin activation. Wnt/APC/β-catenin signaling is the major pathway regulating cellular homeostasis of the intestinal epithelium (18). To decipher the underlying mechanisms involved in the effect of RIP140 in intestinal epithelium homeostasis, we sought to establish whether its expression impacted the Wnt/APC/β-catenin signaling pathway. We first investigated the pattern of β-catenin expression and activity in the intestinal epithelial cells (19). We next proceeded to evaluate the role of RIP140 in β-catenin signaling. We first investigated the pattern of β-catenin expression and activity in the intestinal epithelial cells (19).
Figure 2
RIP140 inhibits intestinal cell proliferation and apoptosis. (A) EdU labeling of proliferating cells or (B) TUNEL staining in paraffin sections of small intestine from RIPKO, wild-type, and RIPTg mice. Images show merged EdU or TUNEL detection (white) and nuclear staining (blue) in small intestine. Staining is expressed as the means of signal quantification ± SD; n = 6 mice for each genotype. Original magnification, ×20 (A) and ×40 (B). (C) Representative H&E-stained transverse sections of small intestines showing crypt survival in irradiated mice (2.5 days after 12-Gy WBI). The number of surviving villi (indicated by black arrows) and the length of villi and crypts were counted in at least six fields of small intestine cross sections from RIPKO, wild-type, and RIPTg mice. Values are the means ± SD; n = 6 mice for each genotype. Original magnification, ×10. Mann-Whitney U test. **P < 0.01; ***P < 0.001.
epithelia of the different mutant mice (Figure 3A). Immunolabeling of paraffin sections of small intestine from Rip140-null and transgenic mice revealed no significant differences in the levels of total β-catenin (Figure 3A, left panel). We then measured the level of active β-catenin (which represents only a small fraction of total β-catenin; ref. 19) using an antibody specific to the protein dephosphorylated on Ser37 or Thr41. Quantification of active β-catenin (ABC) staining revealed clear regulation by RIP140, with a 2-fold increase in RIPKO mice and about a 3-fold decrease in RIPTg mice as compared with controls (Figure 3A, right panel). In order to validate the regulation of β-catenin activation by RIP140, we evaluated β-catenin transcriptional activity in intestinal epithelium by quantifying the mRNA levels of several target genes. As shown in Figure 3B, we observed a significant regulation of most of these genes (e.g., c-Myc [Myc]; claudin 1 [Cldn1], ref. 20; endothelin 1 [Edn1], ref. 21; jagged 1 [Jag1], ref. 22; and c-Jun [Jun], ref. 23), whose mRNA levels were increased in Rip140-null mice and reduced in RIPTg mice as compared with levels in wild-type animals. For endothelin 1 and jagged 1, the same deregulation was observed at the protein level by immunofluorescence (data not shown). Finally, since activation of the Wnt pathway drives the differentiation program of Paneth cells (24), we investigated the effect of RIP140 on this lineage. Using an antibody specifically directed against lysozyme, we observed that the number of Paneth...
cells per crypt increased in RIPKO mice and decreased in RIPCtg mice (Supplemental Figure 3), thus strengthening the negative regulation of the Wnt signaling pathway by RIP140.

RIP140 positively regulates Apc gene expression. ABC levels are normally controlled by a multiprotein degradation complex composed of APC, AxIN1 and 2, GSK3β, CSNK1, and PP2A (18). We therefore postulated that RIP140 could control the level of β-catenin activation by influencing the expression of members of this degradation complex. To test this hypothesis, we quantified the mRNAs encoded by the main genes implicated in β-catenin degradation. Interestingly, we found a correlation between expression of the Rip140 gene and the mRNA levels of Apc, Axin2, and Csk1E genes, which were decreased and increased in Rip140-null and transgenic mice, respectively (Figure 3C). This suggests that RIP140 inhibits β-catenin activation in the intestinal epithelium by positively regulating the expression of several members of the β-catenin degradation complex. Altogether, this inhibition of the β-catenin signaling pathway could explain, at least in part, the above-described effects of RIP140 on intestinal homeostasis.

RIP140 inhibits proliferation of human colon cancer cells. Given the critical role played by the Wnt/ APC/β-catenin pathway in promoting colon tumorigenesis, we then investigated the role of RIP140 in human colon cancer cells. First, we measured RIP140 gene expression in different colorectal cancer cell lines by real-time qPCR and observed that the gene was much more weakly expressed in colon cancer cells than in breast cancer cell lines (data not shown). To analyze the effects of RIP140 on the malignant phenotype of human colon adenocarcinoma cells, we established a stably transfected HCT116 cell line overexpressing a GFP-tagged version of RIP140 (HCT-RIP cells) and used the same cells transfected with the pEGFP empty vector (HCT-GFP cells) as a control.

Real-time qPCR analysis confirmed a much greater abundance of RIP140 mRNA in the HCT-RIP cells compared with that in control HCT-GFP cells (Figure 4A, left panel). Similarly, Western blot analysis confirmed that GFP-RIP140 protein was overexpressed in the nuclear fraction of HCT-RIP cell lysates (Figure 4A, right panel). Fluorescence analysis of HCT-GFP and HCT-RIP cells revealed a cytoplasmic labeling corresponding to GFP-expressing cells and...
as expected, a punctuate signal corresponding to the previously described localization of GFP-RIP140 protein in speckles (ref. 25 and Figure 4A, right panel).

We first investigated the effect of RIP140 expression on cell proliferation using xCELLigence technology (Roche Diagnostics), which allows real-time monitoring of cell proliferation through measurement of impedance-based signals. Data shown in Figure 4B clearly demonstrate that RIP140 inhibited human colon cancer cell proliferation in vitro. To determine whether RIP140 affected cell cycle progression, we transiently transfected the two stable HCT116 cell lines with a cdt1-RFP Premo FUCCI Cell Cycle Sensor, allowing the detection of cells in G1. In parallel, we performed EdU labeling in order to monitor cells synthesizing DNA (Figure 4C). When compared with control cells, HCT-RIP cells showed a strong and significant decrease in the S-phase cell population (from 43.8% to 17.8%), with a concomitant increase in the number of cells in the G1 phase (from 24.1% to 34.6%). These data indicate that RIP140 expression results in a significant and specific blockade of cell cycle progression with cell arrest in G1.

We then assessed the impact of RIP140 on tumor growth in vivo by testing the ability of HCT-GFP and HCT-RIP cells grafted s.c. onto immunoedeficient mice to form tumors (Figure 4D). In line

**Figure 5**

Regulation of β-catenin activation by RIP140 in HCT116 colon cancer cells. (A) Total (left panel) and active (right panel) β-catenin in HCT-GFP and HCT-RIP colon cancer cells (β-catenin immunolabeling in green and nuclear staining in blue). Quantifications are the means ± SD; n = 6 independent experiments for each cell line. Original magnification, x63. (B) Levels of total and ABC proteins were detected by Western blot analysis using specific antibodies in HCT-GFP and HCT-RIP cells. (C) Expression of β-catenin–regulated genes was measured by real-time qPCR. Values represent fold changes ± SD corrected by 28S mRNA and normalized to HCT-GFP cells; n = 6 independent experiments for each cell line. (D) TOP/FOP experiments were performed by transiently transfecting HCT-GFP and HCT-RIP cells (left panel) or MEF WT and MEF RIPKO cells (right panel) with the pTOPflash or pFOPflash reporter plasmids (200 ng). Cells were treated with or without Wnt3a-conditioned medium. Relative luciferase activity was expressed as the mean ratio of TOP/FOP luciferase activities ± SD; n = 3 independent experiments. Mann-Whitney U test. *P < 0.05; **P < 0.001; ***P < 0.001.
with the inhibition of cell cycle progression by RIP140, we observed that tumors from RIP140-overexpressing HCT116 cells exhibited a strong and significant decrease in volume (2.5-fold) as compared with that of controls (Figure 4D). Sections of HCT-RIP xenografts stained with H&E showed large areas with faint eosin staining and lysed nuclei, which are usually associated with necrosis (Figure 4E). Such areas with karyolysis were significantly smaller in HCT-GFP tumors than in those in the HCT-RIP tumors (14.4% ± 8.1% and 45.6% ± 19.1% of the whole tumor surface, respectively). Altogether, these data demonstrate that RIP140 inhibits colon cancer cell proliferation in vitro and tumor growth in vivo.

**Effect of RIP140 on Wnt signaling in human colon cancer cells.** To investigate whether the effect of RIP140 on human colon cancer cell proliferation was associated with an inhibition of β-catenin activation, we performed immunofluorescence staining to detect total and active (ABC) β-catenin in the two HCT116 cell lines, with some of the cells overexpressing RIP140. As noticed in mouse intestine, we observed no differences in the quantification of total β-catenin and ABC staining (Supplemental Figure 4B) and resulted in an increase in its target gene expression (Supplemental Figure 4C). Altogether, these results demonstrate a clear inhibition of the Wnt/β-catenin signaling pathway by RIP140. As shown in Supplemental Figure 4D, we assessed the impact of RIP140 on twelve Wnt target genes in both mouse gut and human colon cancer cells and found that a majority of these genes (more than 60% and 80%, respectively) were regulated by RIP140.

**RIP140 increases transcription of the APC gene in human colon cancer cells.** We then quantified the expression of members of the β-catenin degradation complex in the stable HCT116 clones with or without RIP140 overexpression. As shown in Figure 6A, the APC tumor suppressor gene was significantly upregulated by about 2-fold in HCT116 cells overexpressing RIP140. We also observed the specific deregulation of APC mRNA levels (2-fold decrease) after siRNA-mediated depletion of RIP140 expression in RKO cells (Figure 6B). Very few studies have examined the regulation of APC gene transcription. However, characterization of the proximal promoter region of the human APC gene identified functional consensus binding sites for several transcription factors including p53, USF1/2, and SP1 (refs. 26, 27, and Figure 6C). In order to demonstrate transcriptional regulation of the APC gene by RIP140, we transiently transfected HCT116 colon cancer cells with two reporter constructs containing the APC gene’s proximal promoter region (encompassing sequences of 972 bp and 186 bp, respectively) fused to the luciferase coding sequence. As shown in Figure 6D, we observed a clear induction of luciferase activity upon cotransfection with a RIP140 expression vector, indicating that RIP140 was able to transactivate the APC gene promoter. This effect was specific, since control reporter vectors used in parallel were not affected by RIP140 overexpression (Figure 6C and ref. 28).

ChIP experiments were performed to demonstrate the presence of RIP140 on the proximal region of the APC promoter. PCR was performed in parallel on the proximal human APC promoter corresponding to the sequence that conferred regulation by RIP140 in the luciferase assays, on an upstream region (control), and on the CCNE2 promoter (both used as negative controls). As shown in Figure 6D, we observed significant amplification of all three regions after anti-histone H3 ChIP. By contrast, we detected specific recruitment of RIP140 (as compared with the use of an irrelevant antibody only) on the APC promoter, thus identifying the APC gene as a direct transcriptional target of RIP140.

In order to demonstrate that regulation of APC was indeed required for regulation of β activation by RIP140, we used HCT116 cells with APC overexpression or depletion, as assessed by quantification of APC mRNA levels (Figure 6E), and observed regulation of ABC staining mainly after ectopic expression of APC (Figure 6F). Very interestingly, inhibition of ABC staining by RIP140 was lost when APC expression was modulated (Figure 6G), indicating that RIP140 activity was dependent on APC gene regulation.

To further support the role of APC in RIP140 activity in colon cancer cells, we used SW480 cells, which expressed a mutated form of APC (29) and stably engineered these cells to overexpress GFP-
RIP140 (SW-RIP cells) or GFP alone (SW-GFP cells) (Supplemental Figure 5A). The effect of RIP140 overexpression on in vitro cell proliferation (Supplemental Figure 5B) and on tumor growth after xenografting (Supplemental Figure 5C) was significantly less intense in stably transfected SW480 cells as compared with its effect on HCT116 cells (Figure 4, B and C). The effect of RIP140 on β-catenin activation was also milder in SW480 cells than in HCT116 cells (Supplemental Figure 5D).

Decreased expression of the RIP140 gene in human colon cancers. To determine the biological relevance of our results in human pathology, we first quantified RIP140 gene expression in human colon cancer samples as compared with that in normal colon tissues. First, we reanalyzed the published Affymetrix DNA microarray data on seventeen normal tissues and twenty-two colon cancers (30). As shown in Figure 7A, the results clearly revealed a significant decrease in RIP140 mRNA levels in colon cancer biopsies. We confirmed these findings by real-time qPCR analysis of twenty-four pairs of normal colon and tumor colon tissue samples and found decreased RIP140 expression in almost 60% of the tumors (Figure 7B).

In order to highlight the decreased RIP140 gene expression in colon cancer, we performed immunohistochemical analysis using an anti-RIP140 antibody on a tissue microarray (TMA) containing twenty-four adenocarcinoma samples matched with the corresponding normal tissues. As observed at the mRNA level, we noticed a significant decrease in nuclear RIP140–positive cells in 60% of the colon adenocarcinomas as compared with those seen in normal tissues (Figure 7C).

We searched for a correlation between RIP140 and APC, MYC, or GAST gene expression in 396 colorectal adenocarcinomas. Statistical significance was assessed using Spearman’s correlation analysis.

Figure 7
RIP140 expression in human colon cancers. (A) RIP140 mRNA was quantified in seventeen normal tissues and twenty-two colorectal adenocarcinomas as previously described. Data are presented as box plots showing the median value, upper and lower quartiles, and minimum and maximum nonoutlier values. (B) RIP140 mRNA levels expressed in AU after normalization to actin mRNA levels in twenty-four matched normal and tumor colon samples (the means of the two populations are indicated as horizontal lines). (C) RIP140 immunohistochemistry of a tissue microarray containing normal mucosa from twenty-four patients together with the corresponding adenocarcinoma. Nuclear RIP140–positive cells were counted and are represented in a dot plot. A Mann-Whitney U test was used for statistical analysis. Original magnification, x10. (D) Correlation between RIP140 and APC, MYC, or GAST gene expression in 396 colorectal adenocarcinomas. Statistical significance was assessed using Spearman’s correlation analysis.
Figure 8
RIP140 is a prognosis marker in colon cancer. (A) Kaplan-Meier plots of cumulative survival of Apc<sup>+/+</sup>, Apc<sup>Δ14/+</sup>, and Apc<sup>Δ14/+</sup> Rip140<sup>+/–</sup> mice; n = 4 mice for the Apc<sup>Δ14/+</sup> Rip140<sup>+/–</sup> genotype and 6 mice for the Apc<sup>+/+</sup> and Apc<sup>Δ14/+</sup> genotypes. (B) Kaplan-Meier plots of OS of patients with tumors exhibiting low or high RIP140 mRNA expression. (C) RIP140 immunochemistry of a tissue microarray containing adenocarcinomas from fifty-nine patients. Patients were ranked according to RIP140 expression in the tumor and divided into low and high RIP140 expression groups, respectively. Kaplan-Meier plot of the cumulative survival of patients with low or high RIP140 gene expression. A log-rank test was used for statistical analysis. Original magnification, x10. (D) Schematic diagram depicting the role of RIP140 on β-catenin signaling in human colon cancer cells. RIP140 directly increases APC gene expression, which inhibits activation of β-catenin (RIP140 reduces levels of the unphosphorylated form of β-catenin). As a consequence of the drop in ABC levels, RIP140 inhibits Wnt target gene expression and decreases the ability of human colon cancer cells to proliferate and form tumors in nude mice.
would affect APC gene expression. As shown in Supplemental Figure 6, we found that the correlation between APC and RIP140 expression was indeed stronger in microsatellite-unstable tumors (rho = 0.659) than in microsatellite-stable tumors (rho = 0.439). Finally, and as expected, RIP140 expression was inversely correlated with β-catenin target genes such as c-Myc (MYC) or gastrin (GAST) in the whole cohort (Figure 7D). Altogether, these data clearly demonstrate that colon tumorigenesis is associated with a decrease in RIP140 expression at both the mRNA and protein levels, a finding that is inversely correlated with the activation of Wnt signaling.

**RIP140 expression as a prognostic marker in colon cancer.** The results obtained in colon cancer cells and in patients led us to look for a link between RIP140 expression and colon cancer prognosis. As a first approach to demonstrate that low RIP140 expression levels in colon cancer could be associated with shorter survival, we used the Rip140 Δ14/Δ14 mouse model bred with the tumor-prone ApcΔ14/Δ14 mouse strain (5). These mice bear a germline invalidation of the Apc gene due to deletion of exon 14, which mimics human FAP in the severity of colon tumorigenesis. In this Apc-inactivated background, the loss of one Rip140 allele strongly affected the lifespan of these mice. As shown in Figure 8A, Kaplan-Meier plots revealed that mice heterozygous for both the Apc and Rip140 genes died earlier than ApcΔ14/Δ14 mice bearing the wild-type Rip140 gene. These mouse experiments thus strongly supported the hypothesis that RIP140 expression could be associated with survival in colon cancer patients.

In support of this hypothesis, using the same set of tumors as in Figure 7A, we found that patients with high RIP140 mRNA levels in the tumor presented better rates of overall survival (OS) than patients with low RIP140 mRNA levels (Figure 8B). RIP140 expression was also found to be significantly correlated with progression-free survival (PFS) in two independent cohorts (Supplemental Figure 7, A and B). Moreover, as shown in Figure 8C, data obtained from labeling of a TMA containing fifty-nine adenocarcinoma samples indicated that OS rates were higher in patients bearing tumors with high RIP140 expression than in those with tumors expressing low levels of RIP140. Altogether, these results demonstrate that RIP140 expression decreases during colon tumorigenesis and is associated with poor patient survival.

**Discussion**

In the present study, we demonstrate that the transcriptional repressor RIP140 is involved in the regulation of intestinal epithelial homeostasis and tumorigenesis. Based on in vivo experiments using transgenic mice, our study first identified RIP140 as a new regulator of intestinal epithelial homeostasis through its significant impact on cell proliferation and apoptosis. Indeed, using both gain- and loss-of-function approaches, our data reveal that RIP140 strongly inhibits cell proliferation and programmed cell death in the intestinal epithelium. Our data also demonstrate that RIP140 plays an important role in colon cancer cells by inhibiting cell cycle progression and cell proliferation in vitro and in vivo after grafting onto nude mice.

Our work clearly demonstrates that RIP140 acts as a new inhibitor of the Wnt signaling pathway through the regulation of β-catenin activation (see Figure 8D). Indeed, RIP140 overexpression clearly inhibited the levels of ABC in both transgenic mice and in human colon cancer cells and tumor biopsies. This effect was observed by measuring the level of unphosphorylated ABC, by quantifying the expression of several β-catenin target genes, and by analyzing the regulation of a transiently transfected β-catenin–responsive reporter construct. Such regulation of the Wnt/APC/β-catenin signaling pathway by RIP140 could explain its effect on intestinal homeostasis and tumorigenesis, since many studies have demonstrated the importance of the Wnt pathway in controlling these processes. For instance, loss of APC function or altered β-catenin/TCF4 signaling in intestinal epithelium led to enhanced proliferation and apoptosis (33).

However, other nuclear signaling pathways targeted by RIP140, such as NRs (7) and E2Fs (8), could mediate some of the effects we observed. Indeed, several NRs exhibit a strongly deregulated expression in intestinal epithelium and have been shown to play an important physiopathological role in the intestine (34). For instance, receptors such as LRH-1 have promitotic activity, while others such as VDR, ER, and FXR play an antitumor role in mice (35). Some of these effects are mediated through regulation of the Wnt/APC/β-catenin signaling pathway by the corresponding NRs (36). By contrast, few data are available on the role of E2Fs in the intestine, although a recent study revealed that E2F4 determines cell cycle progression in normal intestinal epithelial crypt cells (37), while another study reported the role of Rb in the maintenance of enterocyte quiescence (38).

Interestingly, our work demonstrates that RIP140 directly induces transcription of the tumor suppressor APC gene. Indeed, RIP140 was recruited on the APC gene promoter and increased its transcription in transient transfection experiments. More importantly, RIP140 and APC expression was significantly correlated in mouse intestinal epithelium, in human colon cancer cell lines, and in human tumor biopsies. Although very few studies on the APC gene promoter have been published, several functional binding sites have been found in the human promoter sequence (26), and further work will be necessary to decipher the precise underlying molecular mechanisms.

Our data clearly indicate that the effects of RIP140 on β-catenin activation rely on the regulation of APC expression in human colon cancer cells. This observation could also be relevant in colon cancer cells bearing a mutation of the APC gene (see data on SW480 cells in Supplemental Figure 5) and is in line with a recent study demonstrating that truncated APC controls proliferation and β-catenin activity in colorectal cancer cell lines (39). However, more complex mechanisms could be implicated in physiological conditions. Indeed, in the mouse intestinal epithelium, we found that RIP140 regulated the expression of other members of the β-catenin destruction complex such as the Csk1E and Axin2 genes. Positive regulation of the Axin2 gene was surprising, since one would have expected the inhibition by RIP140 that occurs with other β-catenin target genes. This suggests that other transcription factors controlling expression of the Axin2 gene are positively regulated by RIP140. Interestingly, the Axin2 promoter encompasses a CG-rich region with Sp1 binding sites, which might support such a transcriptional increase in response to RIP140, as we previously reported for several promoters containing Sp1 binding sites (28).

Our results also demonstrate that RIP140 expression is finely regulated in both the normal and tumoral intestinal epithelia. The RIP140 gene is expressed in epithelial cells, with stronger expression in differentiated cells of the villus than in proliferating cells of the crypt. The same gradient of APC gene expression was observed along the crypt/villus axis, where it counteracts β-catenin signaling and allows differentiation (40). We also showed that RIP140 expression (at both the mRNA and protein levels) strongly decreased during tumorigenesis. An attractive hypothesis to explain all of this regulatory activity is that RIP140 expression is negatively controlled by...
the Wnt/β-catenin pathway itself. Several negative transcriptional responses to activation of the Wnt/β-catenin pathway have been reported (41), and a negative transcriptional element (sequence AGAWAW) has been identified in the promoter of the repressed genes (42). Bioinformatics analysis of the RIP140 gene promoter identified two AGAWAW sites in the proximal region. The existence of such a transcriptional regulatory loop would explain the low RIP140 gene expression in proliferating crypts and colon tumors, both of which exhibit constitutively high Wnt/β-catenin activity, and might indicate that RIP140 expression must be tightly controlled to avoid deleterious effects on cells.

Finally, our work demonstrates that patients whose tumors contained high levels of RIP140 had better PFS and OS rates than those whose tumors had low RIP140 levels. Further studies are now required to define the clinical relevance of RIP140 as a prognostic marker compared with the previously reported gene signatures (43). These data, linking RIP140 gene expression with patient survival, were supported by the decreased lifespan of mice bearing the ApcΔ14/+ deletion associated with loss of one allele of the Rip140 gene. It should be noted that during the crossing experiments, we did not obtain Rip140−null animals on an ApcΔ14/+ background, suggesting that such double-transgenic animals are not viable. Altogether, our results strongly suggest that RIP140 acts as a modifier gene of the APC-mutable phenotype in colon tumorigenesis. Indeed, it is now well known that although APC is considered the central gatekeeper gene in colon cancer, several genetic modifiers (such as the Mmm genes for modifier of Min 1) have been identified and connect myriad factors that determine the course of the disease (for a review, see ref. 4). In vivo experiments using chemically induced carcinogenesis would also confirm the relevance of RIP140 in the initiation of colon tumorigenesis. A first experiment using azoxymethane and dextran sulfate sodium in RIPKO mice led to a stronger inflammatory response associated with histological abnormalities as compared with that in their wild-type littermates (data not shown). In line with all of these observations, a recent study performed in colon cancer samples by the Cancer Genome Atlas Network reported a mutation in the RIP140 gene that encoded a truncated protein together with several nonsynonymous SNPs introducing amino acid changes in the RIP140 coding sequence (44). Interestingly, another frame-shift mutation was previously reported in patients with familial colon cancers (45).

In summary, this study uncovers the role of RIP140 in intestinal homeostasis and colorectal carcinogenesis. In addition, these results identify RIP140 as a new regulator of APC gene transcription and as an inhibitor of the Wnt/β-catenin pathway. Although several molecular events involved in tumor progression have been identified, the full sequence of steps leading to cancer is yet to be established. Our findings extend the knowledge of the role and mechanism of action of the RIP140 gene at a molecular level and highlight, at a clinical level, the potential value of RIP140 as a biomarker with diagnostic, prognostic, and therapeutic utility in colon cancer.

**Methods**

**Animals.** C57BL/6 Rip140−/− (RIPKO) mice (12) were provided by M.G. Parker (Imperial College London). C57BL/6 129 Rip140 transgenic (RIPtg) mice were generated using Speedy Mouse Technology (Nucleus) by insertion of a single copy of human RIP140 cDNA at the HPRT locus. The coding sequence of the human RIP140 gene (BclI/Clal fragment) was cloned into the BamHI site of the Gateway pENTR-1A vector (Invitrogen) previously modified to include the rabbit β-globin polyA sequence and the CAG promoter. The transgene was then transferred into the pDEST-HPRT vector (Nucleus), which was linearized using AgeI and electroporated into HPRT-deficient BPES ES cells by standard methods. This construct contained two regions of homology with the HPRT gene, which allowed insertion of the transgene at this locus. The ES cell clones with homologous recombination were selected on hypoxanthine-aminopterin-thymidine–supplemented (HAT-supplemented) medium, and genotyping of HAT-resistant ES cell clones was performed by PCR analysis of genomic DNA using primers in the RIP140 gene and in the poly(A) sequence (Supplemental Table 1). Targeted ES cells were injected into C57BL/6-derided blastocysts that were then transplanted into the uteri of recipient females. Resulting chimeric males were bred with C57BL/6 females, and the F1 offspring female offspring were backcrossed with C57BL/6 males. Animals were genotyped by qPCR using RIP140 primers. The tumor-prone ApcΔ14/+ mouse strain was obtained from C. Perret (Institut Cochin, Paris, France) (5). These mice bear a germline inactivation of the Apc gene due to deletion of exon 14 and develop intestinal lesions mainly in the colon. All animals were maintained under standard conditions on a 12-hour light/12-hour dark cycle and were fed a chow diet ad libitum, according to European Union guidelines for the use of laboratory animals. In vivo experiments were performed on male and female mice in compliance with the French guidelines for experimental animal studies (agreement B34-172-27).

**WBI.** Twelve-week-old mice were anesthetized (i.p. injection of ketamine and xylazine), placed into a ventilated Plexiglas pie container, and underwent 12-Gy WBI using a Shephard 137Cs-ray irradiator at a dose rate of 4.21 Gy/minute. This was done according to the biosafety guidelines of the Val d’Aurelle hospital. Mice were killed 2.5 days after WBI, and small intestine was removed for histological analyses. The number and length of regenerating villi, together with the length of surviving crypts, were quantified (24 fields per genotype using x10 magnification) in small intestine after H&E staining of paraffin sections (9 animals per group).

**EdU detection in mouse tissues.** Mice were injected i.p. with 100 to 200 μg of EdU (Invitrogen) in PBS. Small intestine was removed 2 hours after injection, paraformaldehyde fixed, paraffin embedded, and sectioned. After paraffin removal, sections were permeabilized with 0.5% Triton X100-PBS for 30 minutes at room temperature and then washed with 3% BSA-PBS. EdU incorporation was detected using the Click-iT EdU cell proliferation kit according to the manufacturer’s instructions (Invitrogen). Following two rinses with PBS, slides were counterstained with Hoechst and mounted for fluorescence microscopy.

**Real-time qPCR.** Total RNA was extracted from cells or mouse tissues using the High Pure RNA Isolation kit (Roche Applied Science) according to the manufacturer’s instructions. Segmentation of wild-type mouse small intestine epithelial cells was performed as described (46). Total RNA (1 μg) was subjected to reverse transcription using Superscript II reverse transcriptase (Invitrogen). Real-time qPCR was performed with the LightCycler 480 SYBR Green I Master (Roche Applied Science) and was carried out in a final volume of 10 μl using 0.25 μl of each primer (25 μM), 5 μl of the supplied enzyme mix, 2.5 μl of H2O, and 2 μl of the template at a 1:10 dilution. After preincubation at 95°C, runs proceeded to 35 cycles of 15 seconds each at 95°C, 5 seconds at 60°C, and 15 seconds at 72°C. Melting curves of the PCR products were analyzed using LightCycler software to exclude amplification of nonspecific products. Results were normalized to R59 or 28S housekeeping gene transcripts. See Supplemental Table 1 for the list of primer sequences.

**ChIP analysis.** For ChIP analysis, HCT116 cells (70% confluent) were cross-linked with 3.7% formaldehyde for 10 minutes at 37°C. The Chimp ChIP One-Day Kit (QUAGEN) was then used according to the manufacturer’s recommendations. Immunoprecipitations were performed using rabbit polyclonal antibodies against acetylated-H3 (06-599; Upstate), RIP140 (Ab42126, Abcam), or an irrelevant IgG antibody as a control. qPCR was then performed using the LightCycler 480 SYBR Green I Master with 2 μl...
of material per point. Primers flanking the RIP140 site of the APC promoter are listed in Supplemental Table 1. The input DNA fraction corresponded to 5% of the amount of immunoprecipitated chromatin.

**Histological and immunofluorescence analyses.** Mouse tissues were fixed with 4% paraformaldehyde, embedded in paraffin, and sectioned (5 μm). Histological analyses were performed by H&E staining of paraffin-embedded tissue sections. For immunofluorescence analysis, following incubation in citrate buffer solution, paraffin-embedded tissue sections were incubated with blocking serum for 3 hours to reduce nonspecific binding. Sections were then incubated with antibodies specific to RIP140 (Ab42126; Abcam), APC (05-665, clone BE7, Millipore), and total β-catenin (610153, clone 14; BD Transduction Laboratories). IF revelation was performed using an Alexa-conjugated secondary antibody (Invitrogen). After washing, sections were counterstained with Hoechst (Sigma-Aldrich) and mounted for fluorescence microscopy. Negative controls using rabbit or mouse IgGs were performed, and no staining was observed in these conditions. Staining quantification was performed at either ×20 or ×40 magnification (24 fields per genotype corresponding to 6 different animals) by determining the mean fluorescence intensity and staining area using the measurement and colocalization module available on AxioVision software, version 4.7.1 (Zeiss).

**Cell culture and transfections.** HCT116 human colon adenocarcinoma cells were stably transfected with the empty pEGFP vector (Clontech) or with the same vector containing full-length human RIP140 cDNA (25). Pools of G418-resistant cells (respectively named HCT-GFP and HCT-RIP) were selected and grown in McCoy medium supplemented with 10% FCS, 100 U/ml penicillin, 100 mg/ml streptomycin, 100 mg/ml sodium pyruvate, and 750 μg/ml G418. To determine the fraction of cells in G1 and S phases, HCT116 cells were transiently transfected with cdT-RFP using the Premo Fusco Cell Cycle Sensor (Invitrogen) and incubated with EdU (then detected as described above for mouse tissues). After fixation and mounting, Fusco and EdU fluorescent signals were quantified only in GFP- and GFP-RIP140–positive cells in order to determine the percentage of cells in G1 phase (Fusco-positive) and in S phase (EdU-positive). All transfections were carried out using Lipofectamine 2000 (Invitrogen) according to the manufacturer’s instructions.

**Luciferase assays.** Stably transfected HCT116 cells and MEFs derived from wild-type or RIPKO mice (12) were transfected with the pTOPflash or pFOPlflash reporter vectors and treated with or without Wnt3a-conditioned medium (47). HCT116 cells were transiently transfected with an APC gene promoter reporter vector or with the same reporter plasmid bearing the SV40 promoter together with a plasmid allowing RIP140 expression (pEF-c-myc-RIP140; ref. 48). Cells were plated in 96-well plates (3 x 10^4 cells per well) 24 hours prior to DNA transfection with Jet-PEI (200 ng of total DNA). The pRLCMV/Bis plasmid (Ozyme) was used to normalize transfection efficiency. Firefly luciferase values were measured and normalized to Renilla luciferase activity. For TOP/FOP assay, the values were expressed as the mean ratio of pTOPflash/pFOPlflash luciferase activity.

**Immunoblotting.** After transfection, nuclear and cytoplasmic protein extracts were prepared using the NE-PER kit (Thermo Scientific), and cell extracts were analyzed by Western blotting using a primary antibody against RIP140 (Ab42126; Abcam), β-catenin (active or total), or β-actin (A2066; Sigma-Aldrich). Signals were revealed using a horseradish peroxidase–conjugated secondary antibody (Jackson ImmunoResearch) and enhanced chemiluminescence (ECL-Plus; GE Healthcare) according to the manufacturer’s instructions.

**Tumorigenicity assay.** Six-week-old nu/nu female mice from Harlan, housed in a pathogen-free facility, were used to assess the effect of RIP140 on tumor growth. HCT-GFP and HCT-RIP cells (8 x 10^6 cells) were injected s.c., and tumor formation was monitored every 3 days using a caliper. Tumor volume was calculated as width x length x thickness. Mice were sacrificed 40 days after injection.

**Cell proliferation assay.** Stably transfected HCT116 cells (HCT-GFP and HCT RIP cells) were seeded at a density of 2,500 cells per well into E-Plate 16 (ACEA Biosciences, Inc.) containing 150 μl per well of medium supplemented with 10% FCS. Dynamic monitoring of cell growth was determined every 24 hours for 4 days using the impedance-based xCELLigence system (Roche Applied Science). The cell index was derived from measured cell-electrode impedance that correlates with the number of viable cells.

**DNA microarray analysis.** Published DNA microarray data (30, 31) were reanalyzed for RIP140, APC, MYC, and GAST expression. Optimal threshold values, which induced the best discrimination according to the disease progression status, were calculated to maximize the Youden’s index. This index is defined as the sum of sensitivity and specificity minus 1 and is frequently used to dichotomize continuous variables in receiver operating characteristic (ROC) curve methodology.

**TMAs.** TMAs contained samples from twenty-four matched paraffin blocks from identified patients with primary colon cancer and adjacent normal tissue (T8235790d; Biochain) or from fifty-nine patients with adenocarcinoma (CD4; SuperBiochip). Three-micrometer sections of the TMA were deparaffinized and rehydrated in graded alcohol. Following epitope retrieval treatment in citrate buffer and neutralization of endogenous peroxidase, TMA sections were incubated overnight at 4°C with anti-RIP140 (Ab42126; Abcam). Immunohistochemical labeling was revealed using peroxidase-conjugated anti-rabbit secondary antibodies (Jackson ImmunoResearch) and 3,3′-diaminobenzidine (DAKO cytomation) as a substrate. Sections were counterstained with hematoxylin. Images were taken using NanoZoomer (Hamamatsu Photonics), and the percentage of stained nuclei was quantified using TissueQuest software (TissueGnostics).

**Statistics.** Data are presented as the means ± SD. Statistical comparisons were performed with Mann-Whitney U or Spearman’s tests. Differences were considered statistically significant at a P value of less than 0.05. PFS was calculated from the start of palliative chemotherapy until disease progression or the last follow-up. Patients who died without progression were censored at the time of death. OS was calculated from the start of palliative chemotherapy until death. Patients lost to follow-up were censored at the time of last contact. The Kaplan-Meier method was used to estimate PFS and OS. The log-rank test was used to test the differences between groups that were considered statistically significant at a P value of less than 0.05. STATA statistical software (STATA) was used for all analyses. Additional methods are described in Supplemental Methods.

**Acknowledgments**

We thank the Réseau d’Histologie Expérimentale de Montpellier (RHEM) for histology facilities and F. Bernex for her expertise. We are grateful to J. Ziliacus (Karolinska Institutet Novum) for the gift of the pEGFP-C2-RIP140 vector, S. Narayan (University of Florida) for providing the APC reporter construct; P. Forzegg (Cnr Saint-Antoine) for the APC expression vector; and A. Coquelle (INSERM U896) for the Fucci/G1 plasmid. We also thank S. Fritsch for initial characterization of the RIP'Tg mice and C. Teysier for critical reading of the manuscript. This work was supported by INSERM, Université de Montpellier 1, Fondation Lejeune, INCa (2011-054), SIRIC Montpellier, and the Institut Régional du Cancer de Montpellier (ICM).

Received for publication June 6, 2012, and accepted in revised form January 23, 2014.

Address correspondence to: Vincent Cavaillès, IRCM, INSERM U896, 208 rue des Apothicaires, Montpellier F-34298, France. Phone: 33.4.67.61.24.05; Fax: 33.4.67.61.67.87; E-mail: vincent.cavaillès@insERM.fr.