Introduction
Exposed tissue surfaces, including skin and the mucosa, are under constant threat of attack by opportunistic and pathogenic microorganisms. Multilayers of epithelial cells lining this host-microbial interface form a defense barrier to prevent microbe penetration (1). Significant effort has been focused on understanding how epithelial cells participate in inflammatory responses when infectious disease occurs or during other forms of tissue challenge (2). It has been shown that Toll-like and Nod-like receptor-mediated innate defense systems allow epithelial detection of extracellular or intracellular microbes via conserved antigens, with resultant expression of cytokines and chemotactic signaling to recruit and activate cellular immune defenses (3–7). Less is known about how epithelia resist infection during health, i.e., under “constitutive” circumstances, except that they provide a formidable barrier.

As an example, the healthy human cornea resists epithelial penetration by virtually all microbes, and infection requires significant (deep-penetrating) damage or contact lens wear (8, 9). In animal models, even large inocula of potentially pathogenic bacteria are rapidly cleared from the ocular surface when it is healthy (10–12). Various antimicrobial peptides are expressed at ocular, and other, epithelia resist infection during health, i.e., under “constitutive” defense against infection. Here we report that epithelial cytokeratins have innate defense properties because they constitutively produce cytoprotective antimicrobial peptides. Glycine-rich C-terminal fragments derived from human cytokeratin 6A were identified in bactericidal lysate fractions of human corneal epithelial cells. Structural analysis revealed that these keratin-derived antimicrobial peptides (KDAMPs) exhibited coil structures with low ξ-helical content. Synthetic analogs of these KDAMPs showed rapid bactericidal activity against multiple pathogens and protected epithelial cells against bacterial virulence mechanisms, while a scrambled peptide showed no bactericidal activity. However, the bactericidal activity of a specific KDAMP was somewhat reduced by glycine-alanine substitutions. KDAMP activity involved bacterial binding and permeabilization, but the activity was unaffected by peptide charge or physiological salt concentration. Knockdown of cytokeratin 6A markedly reduced the bactericidal activity of epithelial cell lysates in vitro and increased the susceptibility of murine corneas to bacterial adherence in vivo. These data suggest that epithelial cytokeratins function as endogenous antimicrobial peptides in the host defense against infection and that keratin-derived antimicrobials may serve as effective therapeutic agents.

Results
K6A contributes to the endogenous antimicrobial activity of corneal epithelial cells. A telomerase-immortalized human corneal epithelial (hTCEpi) cell line, capable of differentiation, stratification, and desquamation similar to normal epithelium (15), was used to mine differentiated cells were lysed and fractionated by molecular weight, and fractions were screened for bactericidal activity against Pseudomonas aeruginosa, a common Gram-negative opportunistic pathogen. For the laboratory isolate PAC1R, lysate fractions containing molecules of less than 3 kDa were most effective in killing bacteria (Figure 1A). For clinical isolates 6294 and 6206, both the <3-kDa and the 3- to 10-kDa fractions were found to be bactericidal in a concentration-dependent manner (Figure 1, B and C).

To examine the functional significance of antimicrobial activity in epithelial cell lysates, lysate fractions were tested for their capacity to impact virulence mechanisms of P. aeruginosa. Both the <3-kDa and the 3- to 10-kDa small-molecule lysate fractions protected cultured human corneal epithelial cells against invasion by P. aeruginosa in vitro, again in a concentration-dependent manner.
The 3-kDa fraction also protected cells against the cytotoxic activity of *P. aeruginosa* strain 6206 (Figure 1E).

Since previously identified endogenously expressed antimicrobial peptides are mostly approximately 3 to 10 kDa or larger in size (16), the bactericidal and cytoprotective activity of the 3-kDa fraction suggested the presence of novel antimicrobials. Thus, lysate fractions were analyzed for their peptide content using LC MS/MS (Table 1). To identify native peptides in the 3-kDa fraction, a nontrypsin-digested fraction was initially analyzed. The majority of detected fragments were derivatives of human keratins, including K5, K6A, K15, and K19. Among these, a 17-amino acid K6A peptide fragment (AIGGGLSSVGGGSSTIK, aa 534–550) was consistently detected, with or without trypsin processing (Supplemental Table 1; supplemental material available online with this article; doi:10.1172/JCI64416DS1). Interestingly, both the N and C termini of this fragment (Ala and Lys, respectively) were flanked

**Figure 1**

Keratin peptide derivatives are major constituents of the antimicrobial fractions of human corneal epithelial cell lysates. (A) Crude lysates were serially fractionated by size to yield 4 fractions: >100 kDa, 10–100 kDa, 3–10 kDa, and <3 kDa (black). Crude lysates and the small-molecule <3-kDa lysate fraction showed bactericidal activity against *P. aeruginosa* (strain PAC1R). Culture media was fractionated in parallel to serve as controls for the corresponding lysate fractions in the bacterial killing assays (white). (B and C) Similar results were observed for *P. aeruginosa* clinical isolate (B) 6294 (invasive strain) and (C) 6206 (cytotoxic strain) except that 3- to 10-kDa lysate fractions were also found to be bactericidal. Dilution (1:3) (gray) reduced activity, demonstrating that antimicrobial activity was concentration dependent. (D) The <3-kDa and 3- to 10-kDa fractions both protected epithelial cells against *P. aeruginosa* internalization. (E) The <3-kDa fraction protected epithelia from bacterial-induced cell death (indicated by trypan blue–staining).
by a theoretical trypsin cleavage site (Arg-Ala and Lys-Tyr, respectively), suggesting that this fragment could be proteolytically released from full-length K6A, e.g., via endogenous trypsin-like serine proteases. The same 17-mer fragment was detected in the 3- to 10-kDa fraction, along with a series of overlapping peptides comprising its full or partial sequence (variants) derived from the C-terminal region (aa 515–559) of K6A. Together these peptides represented the majority of detected fragments in this fraction (Supplemental Table 2).

K6A contributes to ocular defense against P. aeruginosa in vitro. Consistent with a previous proteome study of the healthy human cornea (17), Western immunoblot confirmed the presence of full-length (~60 kDa) and fragmented K6A (<1.7 kDa) in the crude lysate of these immortalized human corneal epithelial cells (Figure 2A). Interestingly, fragments of K6A were not observed in lysates of cells grown in KGM-2 containing lower calcium (0.15 mM CaCl₂) (Figure 2A). The knockdown of hK6A expression using siRNA resulted in an increase in P. aeruginosa growth in the crude lysate, suggesting reduced bactericidal activity (Figure 2B). Controls showed that hK6A knockdown did not significantly impact cell density, morphology, or viability, as demonstrated by trypan blue exclusion assays (data not shown). In vivo knockdown of endogenous K6A in murine corneas (murine K6A) by approximately 25% (assessed by quantitative real-time PCR) using subconjunctival injection of siRNA was found to increase P. aeruginosa adherence to the surface of intact corneas ex vivo by approximately 5 fold (Figure 2, C and E). Controls revealed that this knockdown did not impact general barrier function, as indicated by the absence of penetration by fluorescein (Figure 2D). Collectively these data support the hypothesis that endogenous K6A and keratin-derived antimicrobial peptides (KDAMPs) serve an antimicrobial role in epithelial innate defense.

Structural analysis of KDAMPs. ExPASy proteomic tools were used to predict peptide secondary structures, transmembrane helices, amphipathicity (hydrophobic moment), and net charges (18–21) for 8 variants of K6A (Table 2). Notably, none of the peptides tested showed sequence homology with any known mammalian antimicrobial peptides (22), but they all exhibited a predominance of glycine repeats and a common sequence, GGLSSVGGGS. Variants of the originally identified 17-mer fragment with an additional N- and/or C-terminal residue(s) (19-mer and two 18-mer peptides) were predicted to be favorable for antimicrobial activity based on predicted coil structure, potential for transmembrane helices, a hydrophobic face, and cationic charge. For example, the 19-mer peptide (aa 533–551) was predicted to have features associated with other well-recognized antimicrobial peptides, including a positive net charge and a coil structure able to form transmembrane helices. In contrast, other active KDAMPs showed variability in each of these properties except predicted coil structure. A scrambled (inactive) control of the 19-mer peptide was predicted to retain similar properties to the 19-mer, e.g., positive net charge and coil structure, but not transmembrane helix formation. The scrambled 19-mer peptide also had a lower hydrophobic moment than the active 19-mer and other active KDAMPs (Table 2).

Antipseudomonal activity of synthetic KDAMPs. The bactericidal activity of synthetic K6A fragments was tested using a physiological salt (NaCl) concentration (150 mM) and a “lawn”/biofilm-like culture of bacteria. These conditions are known to reduce the activity of known cationic antimicrobial peptides, such as human β-defensins-1, -2, and -4 and cathelicidin LL-37 (23, 24). The 19-mer KDAMP was almost equally effective in killing the cytotoxic P. aeruginosa clinical isolate 6206 in 150 mM NaCl solution or in water, in both cases reducing bacterial viable counts by approximately 3 to 4 logs after 3 hours at 200 μg/ml (P < 0.005 in each instance) (Figure 3A). This 19-mer peptide also showed bactericidal activity against the invasive clinical isolate 6294 in physiological saline, a 97.5% reduction in bacterial viability after 3 hours at 200 μg/ml, and against a laboratory isolate PAO1, a 96.1% reduction in viability under the same conditions (P < 0.005 in each instance) (Figure 3B). A scrambled sequence control of the 19-mer KDAMP (IRGSVTSYSGGKGSAG) showed little or no bactericidal activity against all 5 P. aeruginosa isolates tested compared with that of the natural peptide sequence (P < 0.01) (Figure 3C). Real-time video phase-contrast microscopy revealed that 19-mer KDAMP-treated P. aeruginosa strain 6206 lost capacity for swimming motility and some appeared to aggregate, compared with untreated bacteria or bacteria treated with a scrambled peptide control (100 μg/μl, 15 minutes), in the presence of hTCEpi cells, which appeared healthy (Supplemental Videos 1–3).

Treatment of cultured human corneal epithelial cells with the 19-mer KDAMP (200 μg/ml) during challenge with P. aeruginosa protected the epithelial cells against bacterial invasion and bacteria-induced cytotoxicity as compared with controls. Bacterial invasion was reduced by 89.1%, while cytotoxicity was reduced by 85.1% (P < 0.001 in each instance) (Figure 3, D and E). Epithelial cells were not visibly impacted by peptide treatment alone, since they showed normal morphology and excluded trypan blue, which only enters epithelial cells if their membrane is permeabilized (data not shown). These data show that KDAMPs have potential to protect epithelial cells against bacterial virulence mechanisms at concentrations not toxic to host cells.

Synthetic analogs of other K6A-derived peptides detected in corneal epithelial cell lysates were also found to be active against P. aeruginosa strain 6206. The 18-mer N-terminal fragment (18–2667

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Table 1

<table>
<thead>
<tr>
<th>Lysate fraction</th>
<th>Tryptic digestion</th>
<th>Total no. of keratin proteins detected</th>
<th>No. of keratin proteins detected</th>
<th>Identity of keratin detected</th>
<th>Total no. of peptides detected</th>
<th>No. of K6A-derived peptides detected</th>
<th>No. of K6A (aa 515–559) peptides detected</th>
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<tbody>
<tr>
<td>&lt;3 kDa</td>
<td>No</td>
<td>8</td>
<td>4</td>
<td>5, 6A, 15, 19</td>
<td>9</td>
<td>2</td>
<td>2</td>
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<tr>
<td>&lt;3 kDa</td>
<td>Yes</td>
<td>1</td>
<td>1</td>
<td>6A</td>
<td>1</td>
<td>1</td>
<td>1</td>
</tr>
<tr>
<td>3–10 kDa</td>
<td>Yes</td>
<td>16</td>
<td>5</td>
<td>5, 6A, 14, 15, 19</td>
<td>82</td>
<td>24</td>
<td>20</td>
</tr>
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</table>

Mass spectrometric analysis of <3-kDa and 3- to 10-kDa lysate fractions revealed that the majority of detected peptides were keratin-derived, while K6A was common to all samples.
mer-N), 18-mer C-terminal fragment (18-mer-C), 17-mer, 14-mer, and 13-mer (fragments within the 19-mer) each showed concentration-dependent bactericidal activity (90% or greater killing at 200 μg/ml, \( P < 0.0005 \)) comparable to that of the 19-mer peptide (Figure 3F). Lower concentrations (100 μg/ml) reduced bactericidal efficacy for most KDAMPs except the 18-mer-N and the 14-mer. These results were somewhat surprising, since several of these peptides (e.g., 14-mer and 13-mer) had predicted features less favorable for antimicrobial function, including less or no positive charge, no transmembrane helix, or hydrophobic face. However, other examples exist of truncated antimicrobial peptides retaining activity, including some of the defensins and LL-37 (25, 26). Two other vari-

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**Figure 2**

K6A contributes to endogenous antimicrobial activity of corneal epithelial cells. (A) Western blot of whole cell lysates (from direct lysis with SDS) confirmed constitutive expression of K6A and its fragments (<2 kDa) in high-calcium–treated (1.15 mM CaCl₂) human corneal epithelial (HCE) cells. Fragments of K6A were not observed in lysates of cells grown in KGM-2 with lower calcium (0.15 mM CaCl₂). FL, full length. (B) Antimicrobial activity of crude cell lysates against *P. aeruginosa* strain 6206 was attenuated by siRNA knockdown of hK6A in vitro. Knockdown efficiency was confirmed by qRT-PCR. \( *P = 0.007 \). (C) Ex vivo bacterial adherence on the corneal epithelium of intact mouse eyeballs (3-hour incubation with \( 10^{11} \) CFU/ml of *P. aeruginosa* strain PAO1-GFP) was increased significantly by in vivo knockdown of mouse K6A (mK6A) in the corneas. Knockdown efficiency was confirmed by qRT-PCR. \( *P = 0.0005 \). (D) Slit-lamp examination of mouse eyes showed that integrity of the physical barrier was not impacted by K6A knockdown, as indicated by the absence of corneal staining with fluorescein. A superficially injured (by tissue paper blotting) mouse cornea served as the positive control of fluorescein staining. (E) Combined 2-photon confocal imaging showed increased adherence of *P. aeruginosa* (PAO1-GFP, green) to whole corneas previously treated with siRNA for murine K6A compared with those treated with scrambled control. Without *P. aeruginosa*, siRNA treatments alone did not visibly affect corneal morphology. Z stacks of 1.0-μm step size (100-μm total thickness) were taken with a \( \times 60 \) water-dipping lens to give an xy field of 210 μm × 210 μm.
ants of the 19-mer with longer or shorter peptide sequences were tested: a 36-mer, comprising all previously tested variants, and a 10-mer, a sequence contained within both the 14-mer and 13-mer KDAMPs (Table 2). Both of these peptides showed dose-dependent bactericidal activity against *P. aeruginosa* (Figure 3F). At 200 μg/ml, killing of *P. aeruginosa* strain 6206 by the 19-mer, 36-mer, and 10-mer was 99.1%, 96.9%, and 99.2% (*P < 0.0005*).

In other experiments, mutations in *P. aeruginosa* LPS, a negatively charged cell wall component of Gram-negative bacteria, had only a minor impact on susceptibility of *P. aeruginosa* strain PAC1R (27) to the 19-mer KDAMP (Figure 3G).

Together these data suggest that KDAMP antimicrobial activity is retained under physiological conditions and requires a specific amino acid sequence(s) and a coil secondary structure but is independent of potential for transmembrane α-helix formation, ability to form a hydrophobic face, and cationic charge.

**KDAMP structure and bactericidal activity.** Most known antimicrobial peptides require a specific secondary structure (e.g., α-helix, β-sheet) for antimicrobial and other functions (28). However, we tested the hypothesis that this was not the case for these short K6A-derived fragments, given their predicted structural characteristics and their common anti-*Pseudomonas* activity, combined with a high glycine composition (35% to 50%) that would be expected to confer minimal conformational constraints. Circular dichroism (CD) spectroscopy was used to analyze the secondary structure of the 19-mer KDAMP, its scrambled control, and the nonbactericidal scrambled 19-mer control exhibit a low to negligible helical content of regardless of environment hydrophobicity, suggesting that α-helicity is not required for KDAMP bactericidal actions.

Since glycine residues were a common feature of naturally occurring KDAMPs (Table 2), and dominance of glycine has been identified in some nonmammalian antimicrobial peptides (22, 30), the contribution of 2 glycine residues to the bactericidal activity of synthetic KDAMPs was investigated. Two glycine residues (G-2 and G-8) of 10-mer peptide (50% glycine content) were substituted with alanine to interrupt consecutive glycine stretches. Thus, GGLSSVGGGS became GALSSVGAGS, while maintaining similar amphipathicity and physiochemical properties. Two site-directed mutant (G2A/G8A) showed reduced bactericidal activity against *P. aeruginosa* (∼86% killing versus >98% killing of control after 3 hours at 200 μg/ml, *P < 0.0001*) (Figure 4C), showing that these glycine residues contribute to activity.

**KDAMP activity against other pathogens.** The activity of synthetic KDAMPs was evaluated against other clinically important bacteria (Figure 4D). As expected, all K6A-derived peptides were active against *P. aeruginosa*. Antimicrobial activity was also observed toward other bacterial pathogens, but levels of activity varied between bacteria and peptides. The group A Streptococcus (*Streptococcus pyogenes*) was particularly susceptible. The 14-mer, the 19-mer, the two 18-mers (the 18-mer-N and the 18-mer-C, lacking the N- or C-terminal residue of the 19-mer, respectively), and the 36-mer were each highly effective against this clinically important pathogen (e.g., >99.9% killing for the 14-mer, 18-mer-N, and 19-mer, after 3 hours at 200 μg/ml, *P < 0.0001*). The 14-mer (>99.9% killing, *P < 0.0001*) and the 18-mer-N (98.1%, *P < 0.0001*) peptide also showed bactericidal activity against *E. coli* but less against *Serratia marcescens* (e.g., 14-mer, 32.4% killing, *P < 0.0001*). However, the 14-mer was highly effective in killing *Staphylococcus aureus* (97.5%, *P < 0.0005*) and *Staphylococcus epidermidis* (>99.7%, *P < 0.0001*). These data show that KDAMP bactericidal activity extends beyond *P. aeruginosa* to other clinically important pathogens, suggesting that compositional flexibility may allow optimal activity against different pathogens. Indeed, it is possible that naturally occurring or synthetic KDAMPs may complement each other in situ to provide a broad-spectrum antimicrobial defense.

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**Table 2**

Sequence and predicted structural characteristics of K6A-derived peptides

<table>
<thead>
<tr>
<th>Peptide</th>
<th>Sequence</th>
<th>Predicted secondary structure</th>
<th>Possible transmembrane helix</th>
<th>Predicted hydrophobic face</th>
<th>pl</th>
<th>Net charge (pH 7.0)</th>
<th>Hydrophobic moment (μH)</th>
</tr>
</thead>
<tbody>
<tr>
<td>19-mer</td>
<td>RAIGGGLSSVGGGSSTIKY</td>
<td>Coil</td>
<td>Yes</td>
<td>Yes</td>
<td>9.99</td>
<td>+2</td>
<td>0.279</td>
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<tr>
<td>18-mer-N</td>
<td>AIGGGLSSVGGGSSTIKY</td>
<td>Coil</td>
<td>Yes</td>
<td>Yes</td>
<td>8.63</td>
<td>+1</td>
<td>0.238</td>
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<tr>
<td>18-mer-C</td>
<td>RAIGGGLSSVGGGSSTIK</td>
<td>Coil</td>
<td>Yes</td>
<td>Yes</td>
<td>11.0</td>
<td>+2</td>
<td>0.348</td>
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<tr>
<td>17-mer</td>
<td>AIGGGLSSVGGGSSTIK</td>
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<td>No</td>
<td>8.80</td>
<td>+1</td>
<td>0.309</td>
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<tr>
<td>14-mer</td>
<td>GGLSSVGGGSSTIK</td>
<td>No</td>
<td>Yes</td>
<td>No</td>
<td>8.75</td>
<td>+1</td>
<td>0.252</td>
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<tr>
<td>13-mer</td>
<td>AIGGGLSSVGGGS</td>
<td>No</td>
<td>No</td>
<td>No</td>
<td>5.57</td>
<td>0</td>
<td>0.325</td>
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<tr>
<td>36-mer</td>
<td>YGSGLGSGFSSGSSGRAIGGGLSSVGGGSSSTIKY</td>
<td>No</td>
<td>No</td>
<td>No</td>
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<td>+2</td>
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<td>10-mer</td>
<td>GGLSSVGGGS</td>
<td>No</td>
<td>No</td>
<td>5.52</td>
<td>0</td>
<td>0.255</td>
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<tr>
<td>Scrambled 19-mer</td>
<td>IRGVSITISGYSGGKGSAG</td>
<td>No</td>
<td>No</td>
<td>Yes</td>
<td>9.99</td>
<td>+2</td>
<td>0.138</td>
</tr>
</tbody>
</table>

A 19–amino acid peptide (aa 533–551) encompassing all 5 fragments (i.e., 18-mer-N, aa 534–551; 18-mer-C, aa 533–550; 17-mer, aa 534–550; 14-mer, aa 537–550; 13-mer, aa 534–546) was predicted to be a cationic peptide. A 36–amino acid variant representing a combined sequence of 2 detected fragments (aa 517–533 and aa 534–552) parted by a tryptic cut site, a 10–amino acid variant representing the overlapping region between 14-mer and 13-mer, and an inactive scrambled 19-mer were also assessed. The shared glycine-rich sequence is underlined. pI, isoelectric point.
KDAMP bactericidal activity involves bacterial cell wall/membrane disruption. To begin to understand how KDAMPs kill bacteria, *P. aeruginosa* were treated with a 19-mer peptide tagged with the fluorochrome TAMRA (5-(and-6)-carboxytetramethylrhodamine) (Invitrogen), and treated bacteria were stained with SYTOX Blue, a cell-impermeant dye. Fluorescence microscopy showed that untreated control bacteria and those treated with a scrambled 19-mer or TAMRA alone did not label with SYTOX, indicating that their cell walls/membranes remained intact, i.e., impermeable to the dye (Figure 5A). Those bacteria also remained motile (Supplemental Videos 1 and 2). In contrast, 19-mer–treated bacteria labeled with SYTOX (blue) and colabeled with TAMRA when the peptide was tagged (red) (Figure 5B) and were nonmotile (Supplemental Video 3). Bactericidal assays (viable counts) confirmed that TAMRA-tagged, or non-tagged, 19-mer–treated bacteria were dead, that killing activity of tagged-19-mer was similar to that of...
the untagged peptide, and that TAMRA alone was inactive (Figure 5C). Interestingly, while TAMRA tagging did not affect activity of 19-mer against *P. aeruginosa*, it did impact 19-mer activity against other bacteria. For example, the TAMRA-tag enhanced 19-mer activity against *S. marcescens* and *E. coli* relative to unlabeled peptide (*P* < 0.005) but reduced activity against *S. pyogenes* (*P* < 0.005) (Supplemental Figure 1). Both of those results may reflect steric effects of the TAMRA-tag on the N-terminal residue, but further studies will be needed to understand their significance. At present, the data show that the K6A-derived 19-mer peptide binds bacterial cells and that it causes bacterial cell wall/membrane permeabilization, loss of swimming motility, and bacterial cell death.

KDAMP-induced bacterial cell permeabilization could directly cause cell death, as has been shown for cationic antimicrobial peptides (31). Alternatively, permeabilization may be secondary to peptide-induced cell death via another mechanism. Thus, the temporal profile of killing by the 19-mer peptide was examined. This was done using log-phase bacterial cultures and a concent-
research article

A  
Untreated  
P. aeruginosa  
BF  
SYTOX Blue  

B  
TAMRA-19-mer  
BF  
SYTOX Blue  

C  
% Survival  
P. aeruginosa strain 6206  
Scrambled 19-mer  
19-mer  
TAMRA-19-mer  

D  
P. aeruginosa strain 6294  
P. aeruginosa strain 6206  
No peptide  
50 µg/ml  
75 µg/ml  
25 µg/ml  

E  
5 min  
19-mer  
Scrambled 19-mer  
No peptide  
DIC  
SYTOX Blue  

15 min  
DIC  
SYTOX Blue
The rapid action of the 19-mer suggested that direct membrane for maximal impact on bacterial viability (>90% killing) (32, 33). and/or protein, generally require at least 30 minutes of exposure effects via inhibition of macromolecular synthesis, i.e., DNA, RNA, (Figure 5D). Antimicrobial proteins or peptides that exert killing 15 minutes, respectively, and 98% of strain 6294 after 15 minutes SYTOX-labeled α-fragments and P. aeruginosa provided initial insights into KDAMP epithelia from bacterial infection. [that they induce cell wall/membrane permeabilization, and that they inhibit bacterial motility. Together, we believe these data establish a new role for cytotkeratin 6 in protecting mammalian epithelia from bacterial infection. Structure-activity and imaging studies with K6A-derived peptide fragments and P. aeruginosa provided initial insights into KDAMP mechanism(s) of action. Firstly, antimicrobial activity of K6A-derived peptides was unaffected by changes in charge or predicted hydrophobic moment, and all but one (36-mer) lacked an α-helical secondary structure in conditions mimicking the environment of bacterial cell membranes. This contrasts with many other previously identified types of antimicrobial peptide for which cationic charge, hydrophobicity, and α-helix formation are important criteria in their antimicrobial function (25, 34). Proposed mechanisms for these requirements include binding to anionic bacterial cell membranes with disruption involving pore formation. However, amphipathicity of antimicrobial peptides can have a more significant role in determining bactericidal activity than hydrophobicity or α-helical content, as shown against E. coli (35). Moreover, rapid action of some antimicrobial peptides involves membrane disintegration via the “carpet” mechanism, in which peptides accumulate to critical thresholds on bacterial membrane surfaces (36). Selectivity between host and microbial cells can also be based on electrochemical potential across the bacterial cytoplasmic membrane due to the proton flux (transmembrane potential) (37). Studies have also suggested that more specific interactions within bacterial cell walls or membranes help to define antimicrobial peptide activity (38). For example, binding of the antimicrobial peptide minis can be mediated by peptidoglycan-associated lipid II (39), and salivary histatins bind a 67-kDa surface protein of Candida albicans (40). Specific cell wall/membrane targeting may explain some of the differences in the spectrum of K6A-derived peptide activity, e.g., the 19-mer is effective against S. pyogenes but not S. aureus or E. coli, both of which are highly susceptible to the 14-mer. Scrambling the sequence of the 19-mer all but eliminated activity against P. aeruginosa, even though predicted charge and coil structure were similar to those of the active peptide. Although the scrambled 19-mer lost ability to form a transmembrane helix, that predicted property was not evident for other smaller KDAMPs (e.g., 13-mer, 10-mer) that showed similar activity against P. aeruginosa as the 19-mer. The scrambled 19-mer did show a reduced hydrophobic moment (μH) of 0.138 μH compared with that of the active 19-mer (0.279 μH) and other active KDAMPs (0.219 μH or greater). Studies with fallaxin-based antimicrobial peptides have shown that active and inactive peptides could be grouped according to hydrophobic moment data, although these data alone were not helpful in predicting activity (41). However, for citropin-based antimicrobial peptides, increased antimicrobial activity and improved bacterial membrane binding (using lipid extracts) were associated with increased hydrophobic moment (42). Thus, differences in the hydrophobic moment of KDAMPs may be helpful in further analysis of their structure-activity relationships. Use of TAMRA-labeled K6A-derived peptides and SYTOX showed these that peptides bind P. aeruginosa and that they cause cell wall/membrane permeabilization. The specific mechanism for cell wall/membrane disruption by K6A-derived peptides is not yet known. Differences between predicted KDAMP structural features (e.g., charge, hydrophobicity, secondary structure, etc.), compared with those of other known antimicrobial peptides, suggest the possibility that the mechanism is novel. Glycine predominates the amino acid composition of the K6A-derived peptides (up to 50% of the 10-mer), and substitutions of glycine residues with alanine at positions 2 and 8 reduced bactericidal activity against P. aeruginosa (38). For example, binding of the antimicrobial peptide nisin can walls or membranes help to define antimicrobial peptide activity (38). For example, binding of the antimicrobial peptide nisin can...
Keratins are an ancient and extremely diverse set of proteins (44). In humans, keratins comprise a large family (54 functional genes) distributed intracellularly and extracellularly among multiple tissues, including skin, teeth, hair, nails, and mucosal surfaces (45, 46). Within epithelial cells, basic (type II) and acidic (type I) cytokera-tins work as complementary pairs to form heteropolymeric intermediate filaments of the cytoskeleton and are key structural components of human cells and tissues. Cytokeratins also play roles in wound healing and tissue regeneration (47) in intracellular signaling and apoptosis (46). Discovery of antimicrobial and cytoproteective properties for K6A-derived peptides could advance our understanding of the functional significance of cytkeratin 6, and other keratins, in general. For example, it might explain the universal presence of keratins in cells and tissues of virtually all species and their diversity within species. Interestingly, a previous study reported that TR65, an antibacterial, pore-forming glycoprotein found in the skin mucus of the rainbow trout, shows strong homology to the trout type II epidermal cytokeratin E1 and is rich in glycine and serine residues (48). Moreover, the authors of that paper suggested that there may be a role for human cytokeratins in innate immunity, given the previously reported increased expression of several cytokeratins in respiratory epithelial cells in response to P. aeruginosa strain PA01 (49). Another study showed that PBP-5, a 5.7-kDa glycerine-, serine-, and threonine-rich antibacterial peptide derived from porcine bladder mucosa, has strong homology to human cytokeratin 7 (50). The abundance of cytokeratin cytoskeletal proteins, and the constant and rapid manner in which full-length cytokeratin monomers are released from their filamentous assembly (51), could support a cellular demand for antimicrobial peptides to help defend human mucosal epithelia against microbial pathogens. It is possible that antimicrobial activity contributes to the demonstrated role of cytokeratin 6 in wound healing and tissue repair (52, 53). It is also of interest that dysfunctional intermediate filament turnover has been implicated in the pathogenesis of various internal conditions, including infectious hepatitis, pancreatitis, and inflammatory bowel disease (54).

Turnover of intermediate filaments, and soluble keratin pro-teins, in epithelial cells is a highly regulated and complex process that is known to involve the ubiquitin-proteasome pathway (UPP) (55, 56). It is not yet known whether the UPP contributes to the generation or degradation of KDAMPs. Our results (Figure 2A) showed a calcium-dependent effect on the generation of fragments from full-length K6A in human corneal epithelial cells, and UPP degradation of host proteins can be increased by elevated intracellular calcium (57). However, generation and degradation of KDAMPs could also reflect the activities of calcium-dependent proteases, e.g., calpains, which are known to target keratins (58, 59) and are present in corneal epithelial cells and other ocular tissues (60). Further studies are underway to investigate the mechanisms of KDAMP generation and regulation in epithelial cells.

In conclusion, this study reveals what we believe to be a novel function for human keratins as a source of endogenous antimicro-bial peptides contributing to antimicrobial activity of cell lysates and defense against bacterial adherence in vivo. Small synthetic analogs of these peptides exert broad-spectrum antimicrobial activity against diverse human pathogens, including P. aeruginosa, E. coli, S. pyogenes, and S. aureus. These peptides are active under physiological conditions and induce bacterial cell wall/membrane permeabilization after binding, which does not require cationic charge or α-helix formation but does require glycine residues. Fur-ther studies will be required to determine the mechanistic details of their bactericidal activity, whether other forms of keratin also give rise to KDAMPs, the mechanism(s) by which they are gener-ated in vivo, and their functional significance in the innate defense of mucosal epithelia and epithelial homeostasis. KDAMPs may also be of value as robust, biocompatible, and efficacious thera-peutic agents for combating infection.

**Methods**

**Bacterial strains.** *P. aeruginosa* corneal clinical isolates 6294 (invasive) and 6206 (cytotoxic), cystic fibrosis clinical isolate 2192, and laboratory strains PAO1 (invasive) and PA14 (cytotoxic) were used in this study. PAC1R and its series of isogenic LPS mutants (PAC1RΔLps, 557, 605, 611) were provided by Gerald B. Pier (Harvard Medical School, Boston, Massachusetts, USA). For antimicrobial spectrum analysis, *E. coli* (ATCC 25922), *S. marcescens* (ATCC 43862), *S. epidermidis* (ATCC 12228), *S. aureus* (ATCC 29213), and *S. pyogenes* (ATCC 19615) were used. For antimicrobial activity assays, lawn culture of *P. aeruginosa* was grown on tryptic soy agar plates at 37°C for 16 hours, while other bacteria were grown on nutrient agar plates for 16 hours at media and conditions specified by ATCC. Starting inoculum, at a concentration of approximately 10^6 CFU/ml, was prepared fresh by suspending bacteria in serum-free Keratinocyte Basal Medium (KBM) (Lonza Inc.) until OD_{600} reading was approximately 0.1. Specific bacterial concentration used in antimicrobial and cytotoxic assays was prepared by 100-fold dilution of this starting inoculum in the indicated test media (~10^6 CFU/ml). For experiments involving peptide effects on bacterial swimming motility or the time course of bacterial killing, broth cultures of *P. aeruginosa* were prepared by subculturing overnight culture in tryptic soy broth at 37°C until OD_{600} reading was approximately 0.3 (log phase). Bacteria were washed and resuspended in saline to an OD_{600} reading of approximately 0.1 (~10^6 CFU/ml).

**Cell culture and lysis fractionation.** hTCEpi cells (15) were maintained at 37°C/5% CO_2 in regular keratinocyte growth medium (KGM-2) containing 0.15 mM CaCl_2 (Lonza). hTCEpi cells in antibiotic-free KGM-2 media were grown to 90% confluence in 96-well tissue culture–treated plates and then switched high-calcium antibiotic-free KGM-2 media (containing 1.15 mM CaCl_2 for 32 hours. Cells were then lysed in 25 µl KBM (Lonza) per well (for antimicrobial and cytotoxicity assays) or 100 mM Tris-HCl (pH 8.5) (for mass spectrometry) by 3 cycles of freeze and thaw. Crude lysate was pooled and confirmed to have equal total protein concentration using a Bicinchoninic Acid Assay Kit (Pierce Biotechnology Inc., Thermo Scientific). Serial fractionation of crude lysate was performed at 4°C using sterile water–prerinised Microcon centrifugal filter devices, with membrane cutoffs at 100, 10, and 3 kDa (Millipore). Fresh vehicle was added back to all lysate fractions to maintain equal volume to that of their original crude lysates.

**Mass spectrometry.** Mass spectrometry was performed at the Proteomics/Mass Spectrometry Laboratory at University of California, Berkeley. A nano LC column was packed in a 100-µm inner diameter glass capillary with an emitter tip. The column consisted of 10 cm of Polaris c18 5-µm packing material (Varian). The column was loaded by use of a pressure bomb and washed extensively with buffer A (see below). The column was then directly...
coupded to an electrospray ionization source mounted on a Thermo-Fisher LTQ XL linear ion trap mass spectrometer. An Agilent 1200 HPLC equipped with a split line so as to deliver a flow rate of 30 nl/min was used for chromatography. Peptides were eluted with a 90-minute gradient from 100% buffer A to 60% buffer B. Buffer A was 5% acetonitrile/0.02% heptafluorobutyric acid (HFBA); buffer B was 80% acetonitrile/0.02% HFBA. Collision-induced dissociation and electron transfer dissociation spectra were collected for each m/z. The SEQUEST and DTASELECT programs were used to identify peptides and proteins from the human database (61, 62).

Bioinformatics. Peptide sequences were analyzed using web-based tools hosted on the Expert Protein Analysis System (ExPASY) proteome server of the Swiss Institute of Bioinformatics (http://ca.expasy.org/tools/#proteome). Specifically, ProtParam was used to compute theoretical PI, NetSurfP was used for secondary structure prediction, TMpred was used for transmembrane helices prediction, HeliQuest was used for amphipathicity and net charge computation as well as helical wheel presentations. Clustal Omega was used to align protein sequences.

Western blotting. hTCEpi cells were lysed on plates directly by tricine gel loading buffer containing SDS and β-mercaptoethanol (Bio-Rad). Detection of full-length and fragmented hK6A and GADPH proteins in the crude lysate (containing 10 μg of total protein) were performed by standard Western blotting using rabbit anti-19-mer antisera (prepared by New England Peptide) and rabbit polyclonal anti-GADPH antibody (sc-25778; Santa Cruz Biotechnology Inc.), respectively, followed by goat anti-rabbit IgG-HRP secondary antibody (Bio-Rad). The rabbit anti-19-mer antisera also recognized murine cytokeratin 6 (data not shown).

siRNA knockdown and real-time PCR. For in vitro knockdown, transfection (6 hours) in 96-well plate format was performed when hTCEpi cells were 80% confluent using Lipofectamine RNAi Max (Invitrogen) and siRNAs (10 pmol in 100 μl) against hK6A (5′-CGAAGGGCGUGUCAAGUC-3′, 5′-GAACAAGUUGUAGCUGCA-3′, 5′-GCAAGUUCACCAUCAAGUA-3′, and 5′-GAAUAUGACUCCUGAGA-3′; ON-TARGETplus SMARTpool L-012116-00-0005; Dharmacon) or scrambled control (ON-TARGETplus Non-targeting Pool D-001810-10-05; Dharmacon), followed by 16 hours of recovery in antibiotic-free KGM-2, and then 16 hours of differentiation in high-calcium antibiotic-free KGM-2 (containing 1.15 mM CaCl2). To collect lysate for antimicrobial activity assays, cells were lysed in 25 μl saline per well (0.9% NaCl) by 3 cycles of freeze and thaw, and crude lysate was pooled and confirmed to have equal total protein concentration prior to use. For in vivo knockdown, C57BL/6 wild-type mice (Charles River) were subcon junctively injected with siRNAs in PBS (5 μl of 8 μg per eye) against mouse K6A (5′-GCUCUGGCUUGCUGUA-3′, 5′-GCUCAGAAGUC- GGGCG-3′, 5′-GGUGAAUGGGAGGAGU-3′, 5′-GAUGUAGAG- UUCUGCUCA3′; ON-TARGETplus SMARTpool L-061465-01-0005; Dharmacon) or scrambled control (ON-TARGETplus Non-targeting Pool D-001810-10-05; Dharmacon). Injectons were performed once a day for 2 days before mice were sacrificed by lethal injection of anesthetic cocktail (21 mg/ml ketamine, 2.4 mg/ml xylazine, and 0.3 mg/ml acupepromazine). Whole eyes were enucleated and PBS-rinsed to exclude tear fluid, followed by incubation with 200 μl GFP-expressing P. aeruginosa strain PAO1 (i.e., 1010 CFU/ml of PA01-GFP) for 3 hours at 35°C, then imaged using a spectrophotometer (model 410, Biomedical Inc.) from 190 to 300 nm using a 0.1-cm path-length quartz cuvette. Peptides were dissolved in pure water or 50% water/50% TFE mixture or pure TFE to a final concentration of 0.1 mg/ml. Three scans were performed at 1-nm intervals and 1-s intervals. Spectra were averaged and substracted for blank control, and mean residue ellipticity values were presented. CD data were further analyzed using the web-based K2D2 program.

Phase-contrast and fluorescence microscopy. To assess bacterial motility, P. aeruginosa clinical isolate 6206 (700 μl; 106 CFU/ml of log-phase culture in saline) was incubated with or without peptide (100 μg/ml synthetic 19-mer peptide or its scrambled control peptide) at 37°C for 15 minutes in the presence of hTCEpi cells in 12-well tissue culture plates. Real-time bacterial motility was visualized with an Olympus IX70 epifluorescent microscope using a ×40 objective and phase contrast (total magnification, ×400). To assess peptide binding and membrane permeabilization, bacteria (200 μl; 106 CFU/ml of suspended lawn culture) were incubated at 37°C for 2.5 hours with or without indicated peptides, including TAMRA-tag peptides and TAMRA tag control (Invitrogen) at the equimolar concentration of 110 μM, followed by 30 minutes of incubation with the impermeant SYTOX blue nuclear acid stain (5 μM; Invitrogen). Bacteria were pelleted by centrifugation, and supernatant was removed before resuspension in 10 μl saline. Bacteria were visualized by phase-contrast microscopy, SYTOX blue stain, and TAMRA fluorescence under a ×100 objective (total magnification, ×1,000).

Statistics. Data are expressed as mean ± SD, calculated from triplicate samples per group in each experiment. Statistical significance of differences between 2 groups was determined by the Student’s t test (2 tailed). P < 0.05 was considered significant. All experiments were repeated at least 3 times, and representative experiments are shown in each figure.

Study approval. All animal procedures were performed in strict accordance with protocols approved by the Animal Care and Use Committee, University of California, Berkeley, which is an AAALAC-accredited institution.

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Address correspondence to: Suzanne M. Fleiszig, University of California, Berkeley, 688 Minor Hall, Berkeley, California 94720-2020, USA. Phone: 510.643.0990; Fax: 510.643.5109; E-mail: fleiszig@berkeley.edu.


