Mechanisms of thyroid hormone action

Gregory A. Brent

Department of Medicine, VA Greater Los Angeles Healthcare System, and Departments of Medicine and Physiology, David Geffen School of Medicine at UCLA, Los Angeles, California, USA.

Our understanding of thyroid hormone action has been substantially altered by recent clinical observations of thyroid signaling defects in syndromes of hormone resistance and in a broad range of conditions, including profound mental retardation, obesity, metabolic disorders, and a number of cancers. The mechanism of thyroid hormone action has been informed by these clinical observations as well as by animal models and has influenced the way we view the role of local ligand availability; tissue and cell-specific thyroid hormone transporters, corepressors, and coactivators; thyroid hormone receptor (TR) isoform–specific action; and cross-talk in metabolic regulation and neural development. In some cases, our new understanding has already been translated into therapeutic strategies, especially for treating hyperlipidemia and obesity, and other drugs are in development to treat cardiac disease and cancer and to improve cognitive function.

Introduction
Thyroid hormone regulates a wide range of genes after its activation from the prohormone, thyroxine (T4), to the active form, triiodothyronine (T3) (1). The signaling pathway is complex and highly regulated due to the expression of cell and tissue-specific thyroid hormone transporters, multiple thyroid hormone receptor (TR) isoforms, and interactions with corepressors and coactivators (2, 3). Furthermore, in many cases, thyroid signals are involved in cross-talk with a range of other signaling pathways (4, 5). Here, we review how clinical observations and animal models have shaped our understanding of this pathway, and how this insight might be translated to therapeutic approaches for a range of conditions (Table 1).

Overview of thyroid hormone action
Thyroid hormone is produced by the thyroid gland, which consists of follicles in which thyroid hormone is synthesized through iodination of tyrosine residues in the glycoprotein thyroglobulin (6, 7). Thyroid stimulating hormone (TSH), secreted by the anterior pituitary in response to feedback from circulating thyroid hormone, acts directly on the TSH receptor (TSH-R) expressed on the thyroid follicular cell basolateral membrane (8). TSH regulates iodide uptake mediated by the sodium/iodide symporter, followed by a series of steps necessary for normal thyroid hormone synthesis and secretion (9). Thyroid hormone is essential for normal development, growth, neural differentiation, and metabolic regulation in mammals (2, 3, 10) and is required for amphibian metamorphosis (11). These actions are most apparent in conditions of thyroid hormone deficiency during development, such as maternal iodine deficiency or untreated congenital hypothyroidism, manifesting as profound neurologic deficits and growth retardation (6). More profound thyroid hormone deficiency or untreated congenital hypothyroidism, manifesting as profound neurologic deficits and growth retardation (6). More profound thyroid hormone deficiency or untreated congenital hypothyroidism, manifesting as profound neurologic deficits and growth retardation (6).

There are two TR genes, TRα and TRβ, with different patterns of expression in development and in adult tissues (2, 13). TRα has one T3-binding splice product, TRα1, predominantly expressed in brain, heart, and skeletal muscle, and two non–T3-binding splice products, TRα2 and TRα3, with several additional truncated forms. TRβ has three major T3-binding splice products: TRβ1 is expressed widely; TRβ2 is expressed primarily in the brain, retina, and inner ear; and TRβ3 is expressed in kidney, liver, and lung (2). Human genetics, animal models, and the use of selective pharmacologic agonists have been informative about the role and specificity of the two major isoforms (2, 14, 15). The selective actions of thyroid hormone receptors are influenced by local ligand availability (1, 16); by transport of thyroid hormone into the cell by monocarboxylate transporter 8 (MCT8) or other related transporters (17); by the relative expression and distribution of the TR isoforms (13) and nuclear receptor corepressors and coactivators (18); and, finally, by the sequence and location of the thyroid hormone response element (TRE) (refs. 19, 20) (Figure 1). In addition, nongenomic actions of thyroid hormone, those actions not involving direct regulation of transcription by TR, have been increasingly recognized (21). Membrane receptors, consisting of specific integrin αv/β3 receptors, have been identified (22) and found to mediate actions at multiple sites, including blood vessels and the heart (23). Several studies have identified direct actions of TR on signal transduction systems (2, 24), which may be especially significant in relation to actions in cell proliferation and cancer.

The broad range of genes whose expression is modified by thyroid hormone status makes studying the effect of thyroid hormone action a daunting challenge (25). Many of the actions of thyroid hormone are the result of potentiation or augmentation of other signal transduction pathways (Table 2 and ref. 5). In metabolic regulation, this includes potentiation of adrenergic signaling (26–29) as well as direct interaction with metabolic-sensing nuclear receptors (30–32). Similar direct receptor-to-receptor interactions and competition for overlapping DNA response elements are seen in neural differentiation, as TR interacts with chicken ovalbumin upstream transcription factor 1 (COUP-TFI) and retinoic acid receptor (RAR) (3, 33).

TR isoforms differ in length at both amino and carboxy termini and are differentially expressed developmentally and spatially (Figure 1). The structure of TRα and TRβ are similar in the DNA and ligand domains and differ most in the amino terminus, and it is thought that the increased potency of TRα is related to its amino terminus (34). Fundamental differences in the ligand-binding pocket have permitted the design of ligands that specifically interact with TRα or TRβ (35), and these have been important tools in the dissection of isoform-specific actions.
<table>
<thead>
<tr>
<th>Clinical condition</th>
<th>Mechanism</th>
<th>Key manifestations</th>
<th>Thyroid function studies</th>
<th>Treatment/reversibility</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>RTH</td>
<td>TRβ heterozygous dominant-negative A</td>
<td>Variable, can include goiter; hearing deficit; hyperactive behavior; learning disability; developmental delay; tachycardia</td>
<td>Elevated serum T4 and T3; nonsuppressed TSH</td>
<td>No single treatment and should be individualized; high-dose T3 every other day can suppress TSH and reduce goiter; treatment to reduce T4 may be indicated during pregnancy to reduce miscarriage rate; other targeted treatment based on symptoms</td>
<td>63, 64, 85</td>
</tr>
<tr>
<td>Severe-phenotype RTH</td>
<td>TRβ homozygous dominant-negative mutation</td>
<td>Large goiter; dysmorphic features; severe tachycardia; developmental and growth delay; mental retardation; hearing deficit</td>
<td>Elevated serum T4; elevated serum T3; TSH, 35–490 mU/l</td>
<td>Developmental defects not likely to be reversible; cardiac protection with β-adrenergic blockers; optimal thyroid hormone levels not known</td>
<td>70</td>
</tr>
<tr>
<td>RTH variant from TRα mutations</td>
<td>TRα heterozygous dominant-negative mutation</td>
<td>Short stature; delayed bone development; transient delay in motor development; mild impairment of cognitive development; chronic constipation</td>
<td>Serum free T4 and rT3 in low-normal range; T3 in high-normal range; normal TSH</td>
<td>T4 treatment to normalize TSH improved growth and metabolic rate in a few patients studied</td>
<td>67, 68</td>
</tr>
<tr>
<td>Short stature, delayed bone age</td>
<td>SECISBP2 mutation</td>
<td>Short stature and delayed bone age; selenoprotein deficiency partial; only children studied, so phenotype into adulthood not known</td>
<td>Elevated serum T4; low serum T3 relative to elevated T4; elevated rT3; slightly elevated TSH</td>
<td>TSH suppressed with T4 or T3 treatment; benefit of selenium treatment not known</td>
<td>52, 64</td>
</tr>
<tr>
<td>Profound mental retardation, Allan-Herndon-Dudley syndrome</td>
<td>MCT8 mutation</td>
<td>Profound developmental delay; truncal hypotonia and poor head control; mental retardation; spasticity</td>
<td>Elevated serum T3; low serum T4 and rT3; TSH normal or slightly elevated</td>
<td>DITPA does not require MCT8 to reach brain, but likely needs to be given at an early developmental stage for benefit</td>
<td>54, 55</td>
</tr>
<tr>
<td>Linkage of D2 polymorphisms to obesity and type 2 diabetes</td>
<td>Reduced D2 activity; less local production or activation of T3 from T4</td>
<td>Subtle reduction in local T3; perhaps some reduction in resting energy expenditure</td>
<td>Normal serum levels of T4, T3, and TSH</td>
<td>May respond to treatment with T3; challenge is how to monitor and follow when trying to normalize local T3 levels</td>
<td>45, 46</td>
</tr>
</tbody>
</table>

A TRβ gene homozygous deletion in initial family (63), 15% of which displayed no TR mutation. BFor example, β-blocker for tachycardia. C3- to 6-fold upper limit normal. D7- to 10-fold upper limit normal. EReference range, 0.3–4.5, mU/l. FMutation mediates insertion of selenocysteine into selenoproteins.
TR isoform selectivity for TRE sequences in genes that mediate thyroid hormone response have been seen in some studies, but not all. TRE sequences influence TR isoform interaction with ligand (36) and may influence coactivator recruitment (37). TR interaction with TREs is not static; as has been reported with other nuclear receptors, there is variation in the pattern of binding that may be influenced by the TRE (37). In vitro studies have shown some TR isoform preferences for specific TREs (38), although the ability to translate these findings to in vivo observations are likely limited. Liver gene profiling in TRα and TRβ gene knockouts demonstrates little in the way of specific genes linked to a TR isoform (25). A recent study, however, suggests that the relative potency of activation may be controlled more by the relative expression of TRα or TRβ in a tissue, rather than by TR isoform specificity for a specific TRE (39).

Cell membrane thyroid hormone transport and local ligand availability

Local activation of T3 from the prohormone T4 at the tissue level is increasingly recognized as an important mechanism of regulation of thyroid hormone action (40). The activity of type 2 5′-deiodinase (D2) is regulated by a ubiquitinase/deubiquitinase mechanism. T4 deiodination by D2 results in exposed lysine residues in D2: ubiquitination of these residues reduces D2 activity, and deubiquitination increases D2 activity (41, 42). Rodents derive circulating T3 primarily by the action of type 1 5′-deiodinase (D1), but humans rely primarily on D2 (1). The inactivation of T4 to form reverse T3 (rT3), mediated by type 3 5-deiodinase (D3), is also important in regulating tissues levels of T3, especially in thyroid axis regulation and sensory development (43, 44). Some — but not all — human genetic linkage studies of polymorphisms in D2 have shown an association with obesity and diabetes (45, 46).

The relationship between the level of serum T4 and serum TSH, termed the set point, is stable for an individual when repeatedly measured prospectively, but varies significantly between individuals (47). This variability in set point in the population suggests that there is a genetic influence involving one or more genes in the thyroid hormone pathway (48). D2 polymorphisms have been associated with an altered pituitary set point of TSH (49) and with a blunted increase in serum T4 after thyrotropin-releasing hormone–stimulated (TRH-stimulated) acute increase in serum TSH (50). Specific D2 polymorphisms were linked to an improved response in hypothyroid patients to replacement with combined therapy of T4 and T3, rather than T4 alone (51).
patients may have reduced conversion of T4 to T3 at the tissue level and benefit from replacement with T3. Selenium is required for the enzyme function of all three deiodinases. Individuals with abnormal thyroid hormone metabolism have been described with defects in the SECISBP2 gene, which is required for the synthesis of selenoproteins (52), thus confirming the essential role of this mineral in thyroid metabolism (Table 1).

Thyroid hormone is hydrophobic and was long thought to enter the cytoplasm by passive diffusion. Thyroid transporters, such as the monocarboxylate (MCT) family and organic anion transporters (OATPs), were identified based on measurable in vitro activity, but the physiologic significance of these transporters was not established early on (17). MCT8 was identified as a specific transporter of thyroid hormone and was reported to be located on the X chromosome (53). Individuals with a severe mutation have myelination delays, which are thought to be caused by impaired thyroid hormone action on oligoden-drocytes (59). MCT8 is expressed in the hypothalamus, a major site of integration of thyroid hormone feedback and gene regulation (60). Exogenous T3, even in the presence of functional MCT8 transporters, does not act on fetal rat brain, due to the requirement for local production of T3 from T4 (61). Studies of MCT8 have shown that the thyroid hormone metabolite diiodothyropropionic acid (DITPA) does not require MCT8 to enter into cells and is a potential therapy for those affected by MCT8 mutations (62). It is likely, however, that DITPA therapy will require treatment at an early stage of brain development to be effective. Thus, thyroid hormone action in the brain is modulated by both regional activation and selective uptake into cells, identifying multiple selective targets for therapeutic interventions.

Expanded spectrum of resistance to thyroid hormone: TRα and TRβ gene mutations

The major clinical condition associated with impaired nuclear action of thyroid hormone, resistance to thyroid hormone (RTH), was first described in 1967 (63). Clinical features include goiter, elevated circulating thyroid hormone levels, nonsuppressed serum TSH level, clinical euthyroidism, and tachycar-dia; some individuals also demonstrate attention deficit disorder and deficits of linear growth, hearing, and bone formation (64). The RTH genetic defect was firmly established by a report published more than twenty years ago of a TRβ mutation in an RTH kindred (65).

The potential phenotype of a TRα mutant RTH syndrome was considered based on the phenotype in animal models with TRα deletion or mutation (66). Recently, two families with different inactivating point mutations in TRα that resulted in receptors with dominant-negative properties have been reported (67, 68). The individual with an E403X TRα mutation had chronic constipation, developmental delay, and short stature, with some improvement after levothyroxine therapy (67). The index patient and her father—who was found to have an insertion of thymine at codon 397 of TRα, resulting in a frameshift and stop codon at 406—had short stature, delayed bone development, transient delay in motor development, and mildly impaired cognitive development; they also had some improvement with T4 treatment (68). Levels of free T4 and rT3 in these patients were in the low-normal range, and T3 in the high-normal range, with normal TSH. These reports are a long-awaited complement to the well-characterized TRβ mutations and provide very strong support to the results of genetic and pharmacologic studies indicating that TR isoforms have distinct roles.

### Table 2

<table>
<thead>
<tr>
<th>Pathway/nuclear factor or target</th>
<th>Process or tissue</th>
<th>TR isoform</th>
<th>Nature of interaction</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Retinoic acid</td>
<td>Neural development</td>
<td>TRα1</td>
<td>Inhibits T3 action generally by direct inhibition of TR</td>
<td>96–98</td>
</tr>
<tr>
<td>COUP-TF1</td>
<td>Brain development</td>
<td>TRα1</td>
<td>Stimulates MCT8 expression and thyroid transport</td>
<td>103</td>
</tr>
<tr>
<td>PPARα</td>
<td>Liver</td>
<td>TRα1</td>
<td>Blocks TR binding to a TRE and inhibits T3 induction of gene expression</td>
<td>99, 100</td>
</tr>
<tr>
<td>PPARγ</td>
<td>Liver</td>
<td>TRβ</td>
<td>Fatty acid oxidation</td>
<td>26</td>
</tr>
<tr>
<td>LXRα</td>
<td>Liver</td>
<td>TRβ</td>
<td>Lipid homeostasis</td>
<td>110</td>
</tr>
<tr>
<td>LXR</td>
<td>Brain</td>
<td>TRα1</td>
<td>Cholesterol metabolism</td>
<td>111</td>
</tr>
<tr>
<td>p85α subunit of PI3K</td>
<td>Thyroid and liver</td>
<td>TRβ4</td>
<td>Cortical layering</td>
<td>113</td>
</tr>
<tr>
<td>β-Catenin</td>
<td>T3 stimulates expression in intestinal epithelium</td>
<td>TRα1</td>
<td>Cell proliferation; tumorigenesis</td>
<td>24, 130</td>
</tr>
<tr>
<td>Adrenergic signaling</td>
<td>White fat</td>
<td>TRα1</td>
<td>Proliferation; tumorigenesis</td>
<td>133</td>
</tr>
<tr>
<td>Adrenergic signaling</td>
<td>Brown fat</td>
<td>TRα1β</td>
<td>Promotes lipolysis</td>
<td>30</td>
</tr>
<tr>
<td>Adrenergic signaling</td>
<td>Heart</td>
<td>TRα1</td>
<td>Adaptive thermogenesis</td>
<td>27–29</td>
</tr>
<tr>
<td>Adrenergic signaling</td>
<td>Bone</td>
<td>TRα1c</td>
<td>Tachycardia</td>
<td>30</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>Increased bone turnover and bone loss</td>
<td>89, 135</td>
</tr>
</tbody>
</table>

- Wild type and mutant.
- TRβ regulates UCP1.
- Role also for TRβ.

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It is of particular interest to compare the phenotype of TRα and TRβ mutations in humans and determine the in vivo role of TR isoform specificity. An important difference is that TRβ mediates thyroid hormone feedback to TRH/TSH, and a mutation blunts this feedback, such that more thyroid hormone is produced (8). In a limited study of RTH patients, the reduction in T3 affinity of the TRβ mutant correlated with the slope of the serum TSH to serum T4 (69). The higher concentration of T4 and T3 in individuals with a TRβ mutation may compensate for the impaired receptor signaling. In patients with TRα mutations, thyroid hormone feedback is not impaired to the same extent, so thyroid hormone levels are not elevated. This may result in peripheral hypothyroidism, and also points to a potential benefit of levothyroxine therapy in these individuals.

Recently, a report of several patients that are homozygous for TRβ mutations demonstrated phenotypic features that represented a combination of those found in individuals heterozygous for a mutation only in TRα or in TRβ (70). Patients homozygous for TRβ mutations have a more severe phenotype of RTH — goiter, hearing loss, and much greater elevations of serum T3, T4, and TSH — than heterozygous individuals. Those homozygous for a TRβ mutation also have intellectual deficits and growth retardation, more characteristic of deficient action of TRα (67, 68). This shows that the mutant TRβ, expressed at sufficiently high levels, antagonizes the actions of TRα.

Role of TR interaction with cofactors

The essential function of gene repression by transcription factor corepressors in development and homeostasis is being increasingly recognized (71, 72). Initial in vitro transfection studies with TR expression vectors showed that the unliganded receptor had a repressive effect on genes positively regulated by TRα and an activating effect on genes normally repressed by TRβ (73). The significance of this property has subsequently been demonstrated by several in vivo models. The mouse model with complete absence of TRα and TRβ has a milder phenotype than a hypothyroid mouse (74). In the setting of TRα gene deletion, the structural effects of induced neonatal hypothyroidism on the mouse brain were not seen (75). The repressive actions of the unliganded receptor, therefore, have a greater physiologic effect than having no receptor at all (18). The interaction of TR with corepressors has been carefully mapped and tested (76, 77). Astapova et al. recently described a mouse model that expressed a version of nuclear receptor corepressor (NCoR) with a mutation in the region that binds TR (78). The disruption of this interaction resulted in a blunted TSH response to thyroid hormone, but enhanced peripheral tissue sensitivity, as the animals were euthyroid despite lower circulating thyroid hormone levels. Interestingly, a mutant NCoR ubiquitously expressed in the background of a TRβ RTH mutant reversed much of the resistance phenotype seen in that model (79). This indicates that constitutive TRβ interaction with a corepressor is an important mechanism for RTH.

In a similar approach, mutation of the coactivator interacting domain in TRβ resulted in resistance to the action of thyroid hormone (80). The interaction of NCoR with histone deacetylase 3 seems to be important for both T3-induced gene activation and repression (81). Another approach to determine the importance of TR coactivator interactions is to determine the impact of coactivator knockouts on thyroid hormone action (82, 83). Mice deficient in the coactivator SRC1 showed increased resistance to the action of thyroid hormone (84). These models may provide a mechanistic basis for the approximately 15% of individuals with an RTH phenotype who lack mutations in TRβ, although no cofactor gene mutations have yet been identified in these patients (85).

TR isoforms and neural development

Highly selective TR isoform requirements have been shown most clearly in models of sensory development, with marked and selective defects of structure and function in the setting of TR isoform inactivation (86). These include development of the inner ear and the cone photoreceptors in the retina (87, 88). Another site with specific TR isoform function is bone, both developmentally and in the adult (89). The developmental importance of TR isoforms is coupled with a requirement for specific transporter expression, such as MCT8 expression in the mouse cochlea (58), as well as a requirement for D2 expression to provide local T3, and for D3 to inactivate thyroid hormone and protect from excessive T3 action during sensitive periods (86, 90).

Thyroid hormone interfaces with other signaling pathways in neural development (Table 2). There is a close developmental link between retinoic acid action in early neurologic development and thyroid hormone action (3). In most model systems studied, retinoic acid acts first, followed by thyroid hormone action. Several studies have shown thyroid hormone targets in early neurologic development and a requirement for TRα expression (91, 92). There are multiple genes whose expression is known to be regulated by both TR and RAR at the TRE (93–95). TR and RAR interact in promoting neural differentiation (96, 97), including a repressive action of the unliganded RAR, as has been shown for unliganded TR (73, 98).

The orphan nuclear receptor COUP-TF1 is expressed early in neurodevelopmental, when thyroid hormone is present, but before the brain is responsive to it (33). Thyroid hormone responsiveness of the brain is associated with reduced expression of COUP-TF1. Numerous thyroid hormone gene targets have been identified with overlapping TR and COUP-TF1 response elements (99, 100). The expression of COUP-TF1 blocks TR from binding the TRE, consistent with protection from early T3 stimulation. Calcium calmodulin–dependent kinase IV (CamKIV), a major thyroid hormone target gene in the developing brain, contains a TRE and COUP-TF1 binding site (101). CamKIV is regulated directly by T3 in primary cultured neurons from fetal cortex and promotes the maturation and proliferation of GABAergic interneurons from their precursor cells (102). The timing of the transport of thyroid hormone is tied to RA based on the stimulation of MCT8 gene expression. Using a neuronal cell model, RA was shown to stimulate MCT8 mRNA expression and to confer thyroid hormone transport (103).

TRα protein is expressed in embryonic postmitotic neurons and most adult neurons in the mouse brain, which suggests that thyroid hormone may also have a significant role in the adult brain (104). Thyroid hormone acting through TRα regulates adult hippocampal neurogenesis, which is important in learning, memory, and mood (105, 106). Expression of a mutant TRα is associated with more depressive behavior traits in mice (107).

TR isoforms and metabolic regulation

Specific actions of TR isoforms have been demonstrated for metabolic regulation, including in white fat and brown adipose tissue (BAT). TRα potentiates adrenergic action in white fat, and when TRα is mutated, visceral fat accumulates (30). BAT expresses both TRα and TRβ, which have selective roles in adaptive thermogenesis (28, 29). Adaptive thermogenesis requires adrenergic stimula-
tion, T3, D2, uncoupling protein 1 (UCP1), and both TRα and TRβ (27). The TRα isoform is required for adrenergic signaling (30), and the TRβ isoform is required for stimulation of UCP1 (29). In addition to these examples of thyroid hormone potentiation of peripheral adrenergic signaling, thyroid also influences adrenergic signaling centrally (108).

RTH patients with dominant-negative TRβ mutations have some growth retardation and skeletal defects, but do not consistently present with metabolic abnormalities. However, in a recent study of RTH patients, increased resting energy expenditure was reported (109). These findings suggest that TRα actions mediating adrenergic sensitivity and fatty acid oxidation may be activated by the higher thyroid hormone levels in RTH patients, as is seen in the heart (5).

TRs engage in cross-talk with a range of nuclear metabolic receptors, including PPARα (26), PPARγ (110), and liver X receptor (LXR), in metabolic regulation (111, 112) and in brain cortical layering (Table 2 and ref. 113). The role of thyroid hormone receptor as an endocrine modulator of metabolic interaction, interacting with other nuclear receptors, PPARγ coactivator 1 (PGC-1), and p160 coactivators and corepressors, has been well described (114).

Our understanding of metabolic cross-talk has been applied directly to therapy with the use of TRβ agonists for lipid lowering and weight loss (15, 115). TRβ agonists have approximately 10-fold greater affinity for TRβ than TRα. Initial studies with TRβ agonists showed a marked preference for action in the liver, efficacy in lowering of cholesterol, and, for some compounds, weight loss, all with little effect on heart or bone. A phase 2 trial of the TRβ agonist eprotirome in patients who had not reached LDL target level with a statin demonstrated that the addition of eprotirome produced a dramatic improvement in LDL (116), although cartilage damage in longer-term dog models has led to the withdrawal of these compounds from clinical trials. Although these actions speak strongly for TR isoform specificity, a significant part of the specificity of action of these agents is much greater concentration of the selective agonist compound in the liver compared with the heart (117). A recent study found that the changes in gene expression in the liver were the same after T3 stimulation or exposure to the selective TRβ agonist GC1 (118). MB07811 achieves liver specificity by being activated after entering hepatocytes by the action of cytochrome P450 to generate the TRβ agonist MB07344 (119). Interestingly, providing hypothyroid human subjects with only T3 rather than T4, but keeping their TSH in the normal reference range, also results in reduced LDL cholesterol and slight weight loss (120). This modest local hepatic excess of T3 may be sufficient to lower cholesterol and produce weight loss, even when systemic levels are in the normal range.

The thyroid hormone analog DIPTA was found to have some specificity for action on the heart and was studied in a prospective randomized control study in patients with severe heart failure (121). Although improvement in some cardiac parameters was seen, the metabolic effects of weight loss were profound, and the study stopped. The metabolic effects of DIPTA — reduction in body weight and LDL cholesterol — provide encouragement for beneficial effects of this class of compound, although stimulation of bone turnover and bone loss by DIPTA will limit its therapeutic use (122).

The clinical utility of a TR antagonist has been considered primarily to antagonize the cardiac effects of thyroid hormone, such as ischemia and arrhythmias (123). The structure of the apo TR, without ligand, has not been solved, but important features have been identified from studies of the liganded receptor with agonists and antagonists (35, 123). Helix 12 is the carboxyterminal helix of TR, which folds in response to ligand and is essential for TR interaction with coactivators and corepressors (124). TR antagonists have been designed by adding extension groups on TR agonists that interfere with Helix 12 folding (123), although this approach is not specific for TRα or TRβ.

**Association of thyroid hormone receptor mutations with cancer**

The viral oncogenes v-erbA and v-erbB are the mediators of avian erythroblastosis retrovirus (AEV) induction of erythroleukemias and fibrosarcomas in chickens, first recognized in 1935 (125–127). v-erbA was later recognized as a mutant version of TRα, with features that favor oncogenic activity, including deletion of the Helix 12 TR domain, which prevents T3 binding.

The link between the origins of TR and oncogenes is consistent with the role of thyroid hormone signaling and mutant TRs in several forms of cancer. The PV model, in which animals harbor a specific truncation of TRβ, is associated with the development of thyroid cancer (128). In related studies, TRβ mutations have been identified in a range of cancers, including hepatocellular carcinoma, renal cell carcinoma, erythroleukemias, and thyroid cancer (127, 129). TSH-secreting pituitary tumors have also been linked to TRβ mutations. TRβ mutants are associated with direct interaction with the regulatory p85α subunit of PI3K, which leads to activation of PI3K and increased phosphorylation of Akt and mTOR and results in cellular proliferation and migration (24, 130). Mutations in TRβ promote metastatic spread of thyroid cancer (131). TRβ mutants have also been linked to pituitary tumors by activation of the cyclin D1/cyclin-dependent kinase/retnoblastoma/E2F pathway (132). TRβ directly stimulates transcription of the β-catenn gene in intestinal epithelial cells and may play a role in tumorigenesis in that tissue (133). Expression of D3, which inactivates thyroid hormone, has been associated with proliferation of malignant keratinocytes in basal cell skin carcinomas (134).

**Summary**

The elements required for thyroid hormone action are well recognized, but the interaction among the various pathways has been challenging to understand. Thyroid hormone interacts with a wide variety of signaling pathways, and its action is modulated based on nutritional and iodine status. A range of conditions with disordered thyroid signaling has allowed us to identify key regulatory pathways that are potential therapeutic targets. The availability of TR isoform-selective agonists and the recent reports of patients with RTH due to TRα mutations, as well as those homozygous for TRβ mutations, are strong evidence for TR isoform specificity. The role of the pituitary in responding to a defect in a thyroid hormone action pathway is central to the resulting phenotype. These pathways, as well as the role of thyroid hormone in metabolism, cardiac function, and oncogenesis, are likely to be the focus in applying these findings.

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Address correspondence to: Gregory A. Brent, Department of Medicine, VA Greater Los Angeles Healthcare System, 11301 Wilshire Blvd., Los Angeles, California 90073, USA. Phone: 310.268.3125; Fax: 310.268.4818; E-mail: gbrent@ucla.edu.


5. Liu YY, Bergh JJ. Thyroid hormone crosstalk with nuclear receptor signaling in metabolic regulation. Trends Endocrinol Metab. 2010;21(3):166–173.


10. Andersen S, Pedersen KM, Bruun NH, Laurberg P. Narrow individual variations in serum T(4) and T(3) in normal subjects: a clue to the understanding of subclinical thyroid disease. J Clin Endocrinol Metab. 2011;96(9):2529–2534.


science in medicine


67. Weiss RE. “They have ears but do not hear” (Psalms 135:17)–non-thyroid hormone receptor beta (non-TRbeta) resistance to thyroid hormone. Thyroid. 2008;18(1):5–13.


84. Tan XJ, et al. Thyroid hormone receptor alpha and beta and thyroid hormone receptor alpha in brain cortical layering. Proc