While immune responses in neurodegeneration were regarded as little more than a curiosity a decade ago, they are now increasingly moving toward center stage. Factors driving this movement include the recognition that most of the relevant immune molecules are produced within the brain, that microglia are proficient immune cells shaping neuronal circuitry and fate, and that systemic immune responses affect brain function. We will review this complex field from the perspective of neurons, extra-neuronal brain cells, and the systemic environment and highlight the possibility that cell intrinsic innate immune molecules in neurons may function in neurodegenerative processes.

Introduction

The immune system serves to protect the organism from external and internal danger by using an elaborate network of immune sensors. In the past, immunologists believed that most of these sensors served to detect pathogens, but there is a growing recognition, elegantly formulated by P. Matzinger (1), that the ability of immune sensors to recognize pathogens may have an evolutionary origin in the detection of molecules on distressed cells from within the organism itself. This point of view is of particular interest for our understanding of immune responses in neurodegeneration in the absence of pathogenic triggers. Importantly, immune sensors are not restricted to dedicated immune cells but specialized sensors are expressed by most cell types including neurons; what’s more, immune sensors are also present inside cells, allowing them to detect internal dangers. Once triggered, the immune system can employ cellular or soluble arbiters, which attempt to remove the cause of injury and initiate repair processes, thereby employing all or part of what is called an inflammatory response. However, in the event that the injurious trigger cannot be removed, the immune response may turn into chronic inflammation or cell death.

It is now clear that the immune response forms an integral part of neurodegeneration, and we are gaining a better understanding of how this system may promote or inhibit neurodegenerative processes. The complement system and microglial cells, for example, are invariably activated in brain regions with neurodegeneration, and genetic studies in animal models demonstrate that these immune effectors can be either protective or harmful (Table 1). A growing number of studies also show associations between neurodegeneration in the brain or spinal cord and immune changes in the periphery. These findings have been discussed extensively (2–4) and will not be reiterated here. Instead, we explore immune responses in neurodegeneration at three levels: (a) cell-autonomous immune mechanisms in neurons, (b) immune responses in the extra-neuronal brain tissue, and (c) immune responses associated with neurodegeneration at a systemic level (Figure 1). Theoretically, neurodegeneration may start at any of these levels, and a failure of the immune responses at each of them may thus contribute to or result in neurodegeneration. While the basic understanding of immune responses in neurodegeneration is growing, the clinical implications for humans are still largely unclear.

Immune responses by neurons

Neurodegeneration ultimately targets neurons and can range from damage to synapses or neurites to cell death. It seems therefore obvious to ask whether cell autonomous immune responses in neurons may have a role in neurodegeneration. However, only in the past few years have scientists started to explore how neurons sense danger using intracellular or surface immune sensors (Figure 2). Once a danger signal is detected, neurons can activate intracellular defense mechanisms and can alert neighboring cells via cell-cell interactions or the release of signaling proteins, neurotransmitters, and other messengers.

Innate immune sensors of cell-autonomous distress. The best studied innate immune sensors are TLRs, which signal through various adaptor molecules to activate NF-κB and/or Stat signaling pathways to induce the production of mostly pro-inflammatory cytokines and chemokines (reviewed in ref. 5). Mammalian TLRs not only recognize pathogen-derived molecules such as lipoproteins, peptidoglycans, single-stranded RNA (ssRNA), double-stranded RNA (dsRNA), and unmethylated CpG motifs, but also endogenous molecules produced by stressed and injured cells, including hsp, mRNA, and fibrinogen. Neurons and neural progenitor cells express several TLRs, including TLR2, -3, -4, -7, -8, and -9, and their functions range from regulating neurogenesis to triggering neurite retraction or cell death (reviewed in ref. 6). Because TLRs can be functionally expressed in the ER and in endolysosomes (7, 8), molecules associated with injury or aging could in principle trigger a pathogenic cascade from within neurons. In support of this idea, mitochondrial DNA (mtDNA), which is released from damaged mitochondria, is recognized by TLR9 in the endosome and can lead to an inflammatory response (9). While very little is known at this point about the role of innate immune sensors in neurons, studies in macrophages show that stimulation of TLR3, -4, and -7 can be beneficial by inducing autophagy (10, 11). Autophagy, which is a degradation pathway for long-lived proteins and organelles (12), has largely beneficial functions in postmitotic neurons, and it is possible that damaged mitochondria or intracellular protein aggregates in aged neurons may trigger TLR-dependent autophagy for clean up. Defects in autophagy, as they occur in neurodegenerative diseases (13–16), might thus interfere with such an innate immune process.

Another group of innate immune sensors, nucleotide binding oligomerization domain–like (NOD-like) receptors, form cytosolic protein complexes called inflammasomes, which can activate caspase-1, IL-1, and NF-κB–dependent inflammatory responses (17, 18). Recent studies show that NOD-like receptors (NLK) can also trigger autophagy (19–21), which supports an intriguing link among

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recognition of abnormal molecular patterns, initiation of inflammatory pathways, and activation of autophagy. This mechanism may well be evolutionarily conserved and could allow cells to purge themselves of abnormal protein aggregates, aged mitochondria, or other cellular junk apart from intracellular pathogens. That such a scenario may, at least in part, take place in neurons in a cellular model of Parkinson disease (PD) using the neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), HMGB1 binds to CD11b/Mac1 on microglia to induce a neurotoxic response (28). Additionally, studies in medicinal leech, which is able to regenerate the structure and function of its CNS after mechanical lesions (23), show that both intracellular leech NLR and TLR1 are temporally and spatially associated with regeneration of neurons (24). It will therefore be most interesting to test whether intracellular innate immune sensors are capable of recognizing abnormal patterns or assemblies of the disease-related proteins β-amyloid (Aβ), α-synuclein, huntingtin, or TAR DNA-binding protein 43 (TDP43). It will also be important to determine in which cellular compartment such protein aggregates may be accessible to TLRs, NLRs, and related receptors.

**Alerting the neighbors.** Once neurons have detected cell intrinsic abnormalities, they will alert their neighbors, which include microglia, astrocytes, and possibly other, less abundant cells in the CNS. Injured neurons can release ATP, ions, neurotransmitters, growth factors, cytokines, and other injury response factors, or they can retract inhibitory molecules that keep microglia in a quiet, surveillant state (25). Neurons may release nucleic acids or hsp, which are ligands for TLRs and other receptors, into the extracellular space. For example, hsp60 released from injured neurons can activate microglia via TLR4, causing enhanced nitric oxide production and in turn, neuronal cell death (26). An important mediator alerting other cells to neuronal injury is the DNA-binding protein high mobility group box 1 (HMGB1), which can be released actively from injured cells or passively from dying cells. HMGB1 promoted neurodegeneration in a mouse model of ischemic stroke by activating microglia through receptor for advanced glycation end products (RAGE) (27). Likewise, in a cellular model of Parkinson disease (PD) using the neurotoxin 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), HMGB1 binds to CD11b/Mac1 on microglia to induce a neurotoxic response (28).

**Neuronal sensing of tissue injury through immune receptors.** Neurons are remarkably well equipped with immune receptors and sensor molecules to detect injury in their local environment. In addition to TLRs, they express cytokine and growth factor receptors, major histocompatibility class I, and complement receptors. While dozens of studies have manipulated immune pathways in mouse models of neurodegeneration (Tables 1 and 2), most used global or

### Table 1

<table>
<thead>
<tr>
<th>Molecule</th>
<th>Disease model for:</th>
<th>Genetic manipulation(^a)</th>
<th>Outcome</th>
<th>Reference</th>
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<tbody>
<tr>
<td>C1q</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased neurodegeneration</td>
<td>98</td>
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<tr>
<td>C1q</td>
<td>Prion disease</td>
<td>Deletion</td>
<td>Prevention of prion disease</td>
<td>99</td>
</tr>
<tr>
<td>C3</td>
<td>Prion disease</td>
<td>Deletion</td>
<td>Delayed neuroinvasion</td>
<td>99</td>
</tr>
<tr>
<td>Caspase-1</td>
<td>ALS</td>
<td>Deletion</td>
<td>Extension of life span, decreased gliosis and neurodegeneration</td>
<td>50</td>
</tr>
<tr>
<td>CCL2</td>
<td>AD</td>
<td>Overexpression in astrocytes</td>
<td>Increased plaque load</td>
<td>100</td>
</tr>
<tr>
<td>CD14</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased plaque load</td>
<td>101</td>
</tr>
<tr>
<td>CD21/CD35</td>
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<td>Deletion</td>
<td>Delayed neuroinvasion</td>
<td>99</td>
</tr>
<tr>
<td>CD36</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased neurovascular dysfunction, reduced vascular oxidative stress; no changes in plaque pathology</td>
<td>102</td>
</tr>
<tr>
<td>CD40L</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased plaque load and gliosis</td>
<td>103</td>
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<td>Cry</td>
<td>AD</td>
<td>Overexpression under metallothionein promotorm</td>
<td>Increased plaque load and neurodegeneration</td>
<td>104</td>
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<tr>
<td>CX3CR1</td>
<td>AD</td>
<td>Deletion</td>
<td>Prevention of neuron loss</td>
<td>45</td>
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<td>CX3CR1</td>
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<td>Deletion</td>
<td>Reduced plaque load</td>
<td>47</td>
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<td>AD</td>
<td>Deletion</td>
<td>Decreased Aβ production</td>
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<td>H2-BF/C2</td>
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<td>Deletion</td>
<td>Prevention of prion disease</td>
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<td>IFNγR1</td>
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<td>Deletion</td>
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<td>106</td>
</tr>
<tr>
<td>IL-1β</td>
<td>ALS</td>
<td>Deletion</td>
<td>Extended life span, decreased gliosis and neurodegeneration</td>
<td>50</td>
</tr>
<tr>
<td>PTGER2</td>
<td>AD</td>
<td>Deletion</td>
<td>Decreased plaque load and oxidative damage</td>
<td>107</td>
</tr>
<tr>
<td>RAG1</td>
<td>PD</td>
<td>Deletion</td>
<td>Attenuation of neuronal cell death</td>
<td>76</td>
</tr>
<tr>
<td>RAGE</td>
<td>AD</td>
<td>Overexpression in neurons</td>
<td>Accelerated abnormalities in learning and memory</td>
<td>29</td>
</tr>
<tr>
<td>RAGE</td>
<td>AD</td>
<td>Overexpression of dominant-negative form in neurons</td>
<td>Preservation of learning and memory, decreased neurodegeneration</td>
<td>29</td>
</tr>
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<td>S100B</td>
<td>AD</td>
<td>Overexpression under human S100B promotorm</td>
<td>Increased plaque load and gliosis</td>
<td>108</td>
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<td>TCRβ</td>
<td>PD</td>
<td>Deletion</td>
<td>Attenuation of neuronal cell death</td>
<td>76</td>
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<tr>
<td>TGF-β</td>
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<td>Overexpression in astrocytes</td>
<td>Increased amyloidosis in the cerebral vasculature</td>
<td>109</td>
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<td>Overexpression of dominant-negative form in cells of the myeloid lineage</td>
<td>Decreased plaque load and vascular Aβ deposition</td>
<td>110</td>
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<td>Deletion</td>
<td>Decreased Aβ generation and plaque load</td>
<td>36</td>
</tr>
<tr>
<td>TNFR1/2</td>
<td>PD</td>
<td>Deletion</td>
<td>Protection against dopaminergic cell death</td>
<td>35</td>
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</tbody>
</table>

\(^a\)Genetic modification is present in all cells. C3, complement component 3; Crry, complement-receptor related protein; H2-BF, complement factor B; IFNγR1, IFN-γ receptor 1; PTGER2, prostaglandin E2 receptor; RAG1, recombination activating gene 1; S100B, S100 calcium binding protein B.
Notable exception was the directed expression of RAGE, one of cytokines, chemokines, complement proteins, and nitric oxide, many of which have neurotoxic effects (25). For example, blocking TNF-α attenuates loss of dopaminergic neurons in mouse models of PD (34, 35) and reduces pre-plaque pathology and learning and memory deficits in mouse models of AD (36, 37). Attempts to divide microglia into functional subsets have been hampered by the lack of reliable cellular markers, which may be due in part to the plasticity of these cells and the possibility that they can assume a continuum of different activation states (38). In a surveillant state (previously erroneously referred to as “resting”), microglia sample their environment for danger signals (39) and they seem to maintain this state through a number of close interactions with neurons. If the neuronal ties are severed, microglia become activated and, depending on the context, may support nearby neurons or kill them. Neuronal-microglial ties include CD200 and CD200R or CD47 and CD172a, (each expressed on neurons and microglia, respectively) (40, 41). The best-studied of these cellular interactions is between neuronal fractalkine (CX3CL1) and the microglia-restricted fractalkine receptor (CX3CR1) (42). Mice that are genetically deficient in CX3CR1 display increased neurotoxicity in response to MPTP, overexpressed mutant SOD, and systemic LPS injections (43). Importantly, LPS toxicity was transferred by transplantation of CX3CR1-deficient microglia from LPS-treated mice into brains of wild-type mice, indicating that microglial toxicity is cell autonomous (43). In contrast, lack of CX3CR1 in a model of stroke reduced neuronal cell loss (44) and produced distinct effects in four independent mouse models for AD pathology (Tables 1 and 2). While CX3CR1 deficiency also reduced neuronal loss in mice that express a combination of familial AD mutations in APP, presenilin 1, and t (45), it promoted neuronal t pathology and behavioral deficits in mice that express only mutant t (46). Furthermore, CX3CR1 deficiency reduced amyloid deposition, without reported effects on neurons, in mice expressing human mutant amyloid precursor protein (APP) alone or in combination with presenilin 1 (47).

How the same genetic defect can be both good and bad for neurons is still unclear, but it is likely that microglial function in vivo is shaped by signals from a multitude of receptor pathways. For example, the presence of other triggers of innate inflammatory pathways (e.g., TLR ligands) can induce caspase signaling in microglia, turning them into neurotoxic cells in vivo (48). Interestingly, SOD1 is required for activation of caspase-1, and thus production of mature IL-1, in response to LPS, suggesting a link between innate immune sensors and neurodegeneration. (49). Moreover, expression of an intracellular mutant SOD1 that is linked to genetic forms of amyotrophic lateral sclerosis (ALS) triggered an inflammasome response in microglia, and genetic deletion of caspase-1 and IL-1β extended life span and reduced pathology in mutant SOD1 transgenic mice (Tables 1 and 2; ref. 50).
A key functional state that microglia can assume and that has received much attention recently is the phagocytic state. The clearance of cellular debris or protein aggregates is evidently a positive function of microglia, but phagocytosis of synapses—so-called synaptic pruning—or removal of entire neurites could be a double-edged sword. For example, CX3CR1 deficiency results in increased numbers of dendritic spines and more immature synapses during postnatal brain development, suggesting a role for microglia in the removal of synapses and the maintenance of neuronal plasticity (51). After entorhinal cortex lesion, microglia are involved in the removal of denervated neural processes and reorganization of the brain, and again, CX3CR1 deficiency impairs this process (52). Although these studies suggest that CX3CR1 activation promotes phagocytosis, an independent study found that treatment with CX3CL1 reduces phagocytic engulfment of latex beads in cultured microglia and that mice lacking CX3CR1 show increased uptake of stereotactically injected Aβ fibrils into microglia (47). Clearly, more work is necessary to sort out these differences.

Microglial phagocytosis is also regulated by the purinergic receptor P2Y6, possibly upon binding of UDP released from injured neurons (53) or by complement proteins C1q and C3, which have been linked to synaptic pruning during development (54). However, the role of microglia in (abnormal) synaptic pruning and removal of other neuronal structures in neurodegeneration is still unclear. Likewise, it is debated how much microglia contribute to the clearance of protein aggregates such as those that are characteristic of AD. While several studies show that activation of microglia can lead to reduced Aβ accumulation and plaque formation in APP transgenic mice (55–58), surprisingly, conditional deletion of microglia in two transgenic mouse models did not change Aβ load or development of neuritic dystrophy (58). Since microglia are clearly capable of phagocytosing Aβ assemblies in vitro or on brain slices ex vivo (59, 60), it is possible that the engulfed peptide impairs intracellular degradation (61, 62) or that Aβ or other age-related factors render microglia ineffective.

In addition to microglia, astrocytes become activated in response to neuronal injury, and they too can produce many classical immune factors with important functions in neuronal survival and death (4). Examples include the chemokine CCL2, TGF-β1, IL-1, and IL-6, and some of their effects on amyloid deposition and neurodegeneration are listed in Tables 1 and 2.

**Immune responses in the systemic environment**

Immune activation in neurons or the CNS can propagate to the organism as a whole and induce alterations in immune status. In turn, immune activation in the periphery can affect CNS cells and neurons (Figure 1). The blood-brain barrier (BBB), which is formed by endothelial cells, astrocytic end feet, and basement membrane, restricts access of large molecules and cells from the
blood to the CNS (63). In general, systemic immune molecules present in blood or tissues may signal to the brain through at least four different routes: active transport across the BBB, signaling at the luminal side of the BBB and propagation of secondary factors into the CNS, diffusion via circumventricular organs, and signaling via peripheral nerves such as the vagus (64–66). Blood-based immune cells may gain access to the brain through transendothelial migration across the BBB into the perivascular space, via the choroid plexus and the cerebrospinal fluid, or through post-capillary venules at the pial surface (reviewed by ref. 67).

Immune cells infiltrating the brain. While infiltrating immune cells are critical in CNS diseases such as multiple sclerosis, the extent to which blood-borne cells have a role in classical neurodegeneration is less well understood. In the APP transgenic mouse model, recruitment of peripheral macrophages to plaques or perivascular spaces appears to have beneficial effects, and several reviews have discussed the clinical relevance and mechanistic underpinnings of these findings (68, 69). The chemokine CCL2 and its receptor CCR2 are strong candidates as mediators of macrophage infiltration into the CNS and have captured the attention of researchers (refs. 70, 71; Tables 1 and 2); a recently described model expressing red fluorescent protein from the CCR2 locus and GFP from the CX3CR1 locus may provide new opportunities to study migration of these cells (72).

Besides myeloid cells, T lymphocytes have been detected in post-mortem tissue of AD, PD, and ALS patients (73–75), and at least in corresponding mouse models, T cells may indeed have a functional role in the pathogenesis of these diseases. Accordingly, the MPTP

| Table 2 |

<table>
<thead>
<tr>
<th>Column 1</th>
<th>Column 2</th>
<th>Column 3</th>
<th>Column 4</th>
<th>Column 5</th>
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<td>Beclin 1</td>
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<td>Deletion (haplo-insufficiency)</td>
<td>Increased plaque load and neurodegeneration</td>
<td>16</td>
</tr>
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<td>C3</td>
<td>AD</td>
<td>Deletion</td>
<td>Increased plaque load and neurodegeneration</td>
<td>111</td>
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<td>CCR1</td>
<td>Prion disease</td>
<td>Deletion</td>
<td>Increased Aβ levels, decreased survival</td>
<td>71</td>
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<tr>
<td>CCR2</td>
<td>AD</td>
<td>Deletion</td>
<td>Increased plaque load and memory impairment</td>
<td>113</td>
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<td>PD</td>
<td>Deletion</td>
<td>No effect</td>
<td>114</td>
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<td>Increased levels of endogenous mouse Aβ42</td>
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<td>ALS</td>
<td>Deletion</td>
<td>Increased neuron loss and microglial activation</td>
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</tr>
<tr>
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<td>PD</td>
<td>Deletion</td>
<td>Increased neuron loss and microglial activation</td>
<td>43</td>
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<td>IFN-γ</td>
<td>AD</td>
<td>AAV-mediated overexpression in brain</td>
<td>Increased plaque load, increased microglial activation</td>
<td>117</td>
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<td>IL-1R1</td>
<td>AD</td>
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<td>No changes in plaque load</td>
<td>118</td>
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<td>IL-18</td>
<td>ALS</td>
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<td>No changes in life span</td>
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<td>IL-1β</td>
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<td>Deletion</td>
<td>No effect on life span or motor axon degeneration</td>
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<td>120</td>
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<tr>
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<td>Decreased plaque load, increased gliosis</td>
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<tr>
<td>MyD88</td>
<td>Prion disease</td>
<td>Deletion</td>
<td>No changes in infection or histopathology</td>
<td>122</td>
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<td>AD</td>
<td>Overexpression in astrocytes</td>
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<td>123</td>
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<td>Increased plaque load and dendritic loss</td>
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<td>124</td>
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<td>TLR4</td>
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<td>Deletion</td>
<td>Increased plaque load</td>
<td>125</td>
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</table>

*Genetic modification is present in all cells. C3, complement component 3; IL-1R1, IL-1 receptor 1; MyD88, myeloid differentiation primary response gene 88; TGFBR2, TGF-β receptor 2.*
A growing number of studies recognize TNF-α as a main perpetuator in cognitive dysfunction associated with systemic inflammation or injury. Thus, peripheral injection of LPS into wild-type mice can lead to prolonged increases in cerebral TNF-α levels and loss of dopaminergic neurons in the substantia nigra, effects which are ameliorated in mice lacking both TNF-α receptors (87). TNF-α was also shown to be a main mediator of post-operative cognitive dysfunction following orthopedic surgery in mice (88). Interestingly, TNF-α may also have a role in familial forms of frontotemporal dementia (FTD) with loss-of-function mutations in progranulin. This anti-inflammatory protein was shown recently to bind to TNF-α receptors and inhibit TNF-α-mediated inflammation in a mouse model of arthritis (89), and it is conceivable that it has similar functions in FTD.

Systemic immunity can be coordinated by neural circuits that operate reflexively. The central components of this immunological reflex are the afferent and the efferent arc of the vagus nerve, which sense and inhibit the production of pro-inflammatory stimulation seems to reduce or slow cognitive decline in some of noradrenaline and subsequently acetylcholine in the spleen, patients with AD (90).

Additional evidence linking systemic inflammation with neurodegeneration comes from epidemiological studies, which show that prolonged use of nonsteroidal anti-inflammatory drugs (NSAIDs) reduces the risk of developing AD (32) or PD (33) by as much as 50%. These beneficial effects do not extend into the clinical phase of disease, and clinical trials using several different NSAIDs have not resulted in cognitive improvement (91). While it may simply be too late to target inflammation with these drugs once clinical symptoms are apparent, it is equally possible that more specific modulators of inflammatory processes targeting, for instance, prostanoid receptors, would show benefits.

Additional evidence that neurodegeneration is associated with systemic changes in the immune response comes also from screens of peripheral blood cell transcriptomes (92) and of cellular signaling proteins (93) in blood from patients with AD and controls. While such studies are still underpowered and proteomic studies suffer from technical challenges, our group was recently successful in identifying several overlapping proteins in two studies, which used different detection methods and samples (94). Among these proteins, G-CSF and macrophage-CSF were reduced in the plasma of AD patients in studies conducted by our group and led to reduced amyloidosis and neuroinflammation in APP transgenic mice following systemic administration by two independent groups (95, 96). These results suggest that changes in peripheral immune factors can influence neurodegenerative processes. Interestingly, cognitive decline with aging may be linked, in part, to increased levels of inflammatory protein in the systemic environment. These factors in the blood of old mice were capable of inhibiting hippocampus-dependent spatial learning and memory, long-term potentiation, and adult hippocampal neurogenesis in young mice, and systemic injection of one of these factors, CCL11/eotaxin, was sufficient to elicit some of these effects (97). Together, these observations provide the intriguing possibility that the peripheral immune environment in neurodegenerative diseases could be targeted toward the development of novel therapies that regulate CNS function to slow cognitive decline and neurodegeneration.

Conclusion
While it took 36 years to generate the first 1,200 publications containing the key words “immunology” and “neurodegeneration,” it took only 9 more years to double that number. The small fraction of these studies discussed here make it clear that immune responses are integral to neurodegeneration and that we are just at the beginning of understanding this complex relationship. Exciting new studies linking innate immune receptors to cellular protein homeostasis through phagocytosis and autophagy may provide new insight into immune dysfunction and neurodegeneration. Rapid progress is also being made in our understanding of microglia, both as protectors of neurons and as their possibly worst enemy. Fortunately, we can make active progress toward the development of useful therapies even as research into the basic mechanisms continues.

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Address correspondence to: Tony Wyss-Coray, Stanford University School of Medicine, 1201 Welch Road, Rm. P208, MSLS Bldg., Stanford, California 94305-5489, USA. Phone: 650.852.3220; Fax: 650.849.1983; E-mail: twc@stanford.edu.


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