

Resistance to type 1 diabetes induction in 12-lipoxygenase knockout mice

David Bleich,¹ Songyuan Chen,¹ Brian Zipser,¹ Duxin Sun,² Colin D. Funk,² and Jerry L. Nadler¹

¹Division of Diabetes, Endocrinology, and Metabolism, City of Hope National Medical Center, Duarte, California 91010, USA

²Center for Experimental Therapeutics, University of Pennsylvania, Philadelphia, Pennsylvania 19104, USA

Address correspondence to: Jerry L. Nadler, Division of Diabetes, Endocrinology, and Metabolism, City of Hope National Medical Center, 1500 E. Duarte Road, Duarte, California 91010, USA. Phone: (626) 359-8111, ext. 2689; Fax: (626) 301-8212; E-mail: jnadler@smtplink.coh.org.

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Leukocyte 12-lipoxygenase (12-LO) gene expression in pancreatic β cells is upregulated by cytotoxic cytokines like IL-1 β . Recent studies have demonstrated that 12-LO inhibitors can prevent glutamate-induced neuronal cell death when intracellular glutathione stores are depleted. Therefore, 12-LO pathway inhibition may prevent β -cell cytotoxicity. To evaluate the role of 12-LO gene expression in immune-mediated islet destruction, we used 12-LO knockout (12-LO KO) mice. Male homozygous 12-LO KO mice and control C57BL/6 mice received 5 consecutive daily injections of low-dose streptozotocin to induce immune-mediated diabetes. Fasting serum glucose and insulin levels were measured at 7-day intervals, and the mice were followed up for 28 days. 12-LO KO mice were highly resistant to diabetes development compared with control mice and had higher serum insulin levels on day 28. Isolated pancreatic islets were treated with IL-1 β , TNF- α , and IFN- γ for 18 hours. Glucose-stimulated insulin secretion in cytokine-treated islets from C57/BL6 mice decreased 54% from that of untreated islets. In marked contrast, the same cytokine mix led to only a 26% decrease in islets from 12-LO KO mice. Furthermore, cytokine-induced 12-hydroxyeicosatetraenoic acid (12-HETE) production was absent in 12-LO KO islets but present in C57/BL6 islets. Isolated peritoneal macrophages were stimulated for 48 hours with IFN- γ + LPS and compared for nitrate/nitrite generation. 12-LO KO macrophages generated 50% less nitrate/nitrite when compared with C57BL/6 macrophages. In summary, elimination of leukocyte 12-LO in mice ameliorates low dose streptozotocin-induced diabetes by increasing islet resistance to cytokines and decreasing macrophage production of nitric oxide.

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Introduction

The leukocyte 12-lipoxygenase (12-LO) gene is expressed in pancreatic β cells and macrophages. Products of the 12-LO enzyme are proinflammatory and may be involved in immune-mediated conditions, but formal proof of this hypothesis is lacking. Leukocyte 12-LO is preferentially expressed over 5- and 15-lipoxygenase in pancreatic β cells and may be involved in cytokine-mediated β -cell damage through generation of the lipid hydroperoxide 12-hydroperoxyeicosatetraenoic acid (12-HPETE) (1–7). Reduction of 12-HPETE to 12-hydroxyeicosatetraenoic acid (12-HETE) is facilitated by the enzyme glutathione peroxidase, and recent work has demonstrated that 12-LO inhibitors can prevent glutamate-induced neuronal cell death when intracellular glutathione stores are depleted (8). Both neuronal cell death and peroxide generation required 12-LO activity in this model and were prevented by treatment with a lipoxygenase inhibitor, baicalein (10 μ M). Furthermore, the proinflammatory cytokine IL-1 β was shown to increase 12-LO product formation in pancreatic islets (9, 10), whereas nordihydroguaiaretic, a lipoxygenase inhibitor, protected rat islet cells from cytokine-induced destruction (11). Given that 12-HPETE and/or other 12-LO products may damage pancreatic β cells through

lipid peroxidation, it is possible that inhibition of the 12-LO pathway may protect pancreatic β cells from cytotoxic conditions.

To test this hypothesis, we administered low-dose streptozotocin (STZ) to 12-LO knockout (12-LO KO) mice and genetic controls to evaluate the development of diabetes. In addition, we isolated pancreatic islets and tested them for sensitivity to cytokine-induced dysfunction. We also evaluated the capacity of macrophages to generate nitric oxide (NO) and superoxide when stimulated. We chose the low dose STZ-induced diabetic model because previous studies have demonstrated that β -cell destruction is mediated in part by generation of free radicals and lipid hydroperoxides as well as by immune activation (12, 13). 12-LO KO mice were generated by homologous recombination in embryonic stem cells, as described previously (14).

In the present study, we found that 12-LO KO mice were significantly more resistant to STZ-induced diabetes than were C57BL/6 controls and that islets purified from 12-LO KO mice were protected from cytokine-induced dysfunction. Furthermore, macrophages from 12-LO KO mice generated ~50% of the nitrate/nitrite compared with macrophages from control C57BL/6 mice.

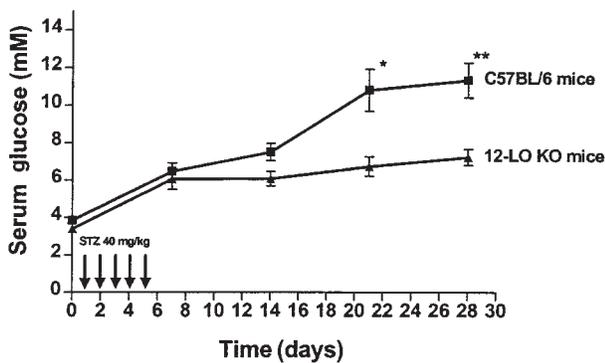


Figure 1
Serum glucose levels in low dose STZ-treated 12-LO KO and C57BL/6 mice. Eight 12-LO KO and 12 C57BL/6 mice received 40 mg/kg of STZ for 5 consecutive days as shown. The mean fasting serum glucose \pm SEM for each group of mice is shown at each time point. 12-LO KO mice were significantly more resistant to STZ-induced diabetes than C57BL/6 control mice. * $P = 0.01$ vs. 12-LO KO. ** $P = 0.003$ vs. 12-LO KO.

Therefore, disruption of the leukocyte 12-LO gene markedly reduces STZ-induced diabetes. The protective mechanisms appear to be due to resistance to cytokine-induced islet dysfunction and reduced macrophage release of NO.

Methods

Glucose determinations were made with a One Touch Profile (Lifescan Inc., Milpitas, California, USA) and validated with a glucose analyzer (Yellow Springs Instrument, Yellow Springs, Ohio, USA). Mouse insulin was measured with mouse-specific insulin RIA kits (Linco Research Inc., St. Charles, Missouri, USA). Anti-12-HETE antibody was purchased from PerSeptive Biosystems (Framingham, Massachusetts, USA). IL-1 β , TNF- α , and IFN- γ were purchased from R&D Systems Inc. (Minneapolis, Minnesota, USA). Cytochrome *c* was obtained from Sigma Chemical Co. (St. Louis, Missouri, USA). Protein determinations were performed according to the method of Bradford using Sigma Brilliant Blue G for detection and Sigma BSA Fraction V as a standard.

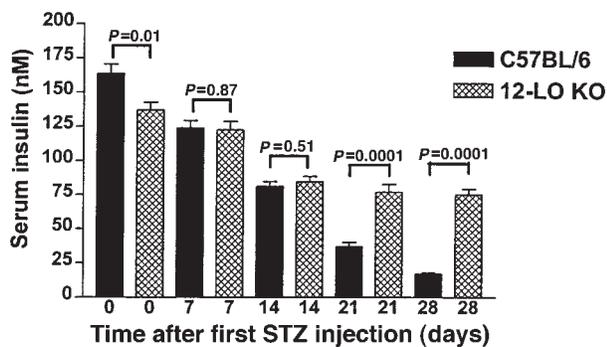


Figure 2
Serum insulin levels in low dose STZ-treated 12-LO KO and C57BL/6 mice. 12-LO KO mice demonstrated a 45% decrease in insulin levels from day 0 to day 28, whereas C57BL/6 mice had a 90% decrease in insulin levels after low-dose STZ treatment. A statistically significant difference in insulin levels between 12-LO KO and C57BL/6 mice is seen on days 21 and 28.

Generation of 12-LO KO mice. The generation of 12-LO KO mice has been described previously (14). The mice were backcrossed to the C57BL/6 genetic background 11 times (>99% C57BL/6). All animal studies were performed in accordance with guidelines set forth by the Research Animal Care Committees of City of Hope National Medical Center and University of Pennsylvania.

Low dose STZ-induced diabetes. Male C57BL/6 and 12-LO KO mice aged 12–16 weeks were used in this study. Mice received 40 mg/kg of STZ intraperitoneally on 5 consecutive days between 0900 and 1000 hours. The mice were housed in a temperature-controlled room with 12-hour light/12-hour dark cycles. Mice were given free access to water and standard laboratory chow during these studies. Blood was obtained weekly from the retro-orbital plexus via capillary tube and used for glucose and insulin determinations. All specimens were obtained after an overnight fast. Mice were followed up for a total of 28 days, at which time they were sacrificed. We defined diabetes as fasting glucose >7.8 mM (~140 mg/dL).

Purification of mouse islets and islet studies. Mice were sacrificed with CO₂ gas, and pancreata were surgically removed. The specimen was placed in a siliconized 25-mL Erlenmeyer flask with 5 mL of a 0.5% solution (wt/vol 15 mg/30 mL) of type V collagenase (Sigma Chemical Co.) diluted in cold HBSS. The pancreas was minced with scissors and incubated in a shaker-type water bath at 37°C for 8–10 minutes. The pancreatic digest was poured into a black-backed petri dish, and islets were picked by hand. Typically, we purified ~50 islets per mouse.

Twenty medium-sized mouse islets per experiment were transferred to 75-mm culture dishes in RPMI-1640 plus penicillin G (100 U/mL) and streptomycin (100 μ g/mL), plus 10% FCS, plus 10 mM HEPES, without glutamine, and cultured overnight.

Purification of mouse macrophages and macrophage studies. Mice were sacrificed with CO₂ gas, and immediately, 10 mL of cold RPMI-1640 medium was injected into the peritoneal cavity under sterile conditions. The medium was aspirated out and the peritoneal cells were counted. Typically, we obtained 10⁶ macrophages per mouse and used 10⁵ macrophages per well. Peritoneal cells were pipetted into 96-well plastic dishes, and macrophages were purified by differential adherence over 1 hour. The cells were gently washed 3 times with DMEM + 0.1% FCS without indicator for nitrate/nitrite determination or with HBSS + Mg²⁺ + Ca²⁺ medium for superoxide determination.

For nitrate/nitrite, macrophages were stimulated with 2 μ g/mL LPS + 100 U/mL IFN- γ , and the medium was sampled at 48 hours. We used Griess reagent to detect nitrate/nitrite formation (1 part 1% sulfanilamide in 5% concentrated H₃PO₄ plus 1 part 0.1% naphthylethylenediamine dihydrochloride plus 2 parts conditioned medium) (15). A standard curve was generated for each experiment using sodium nitrate (Sigma Chemical Co.) in serial dilutions.

For superoxide measurement, we used a cytochrome *c* assay that has been described previously (16, 17). Superoxide reduces ferricytochrome *c* (Fe³⁺) to ferrocyanochrome *c* (Fe²⁺) and can be monitored spectrophotometrically at 550 nm. Peritoneal cells, ~10⁵ per well, were adhered into 96-well plates and stimulated with 50 ng/mL PMA (Sigma Chemical Co.). Cytochrome *c* (160 μ M; Sigma Chemical Co.) was added to the macrophage preparation, and medium was sampled every 30 minutes. Medium containing cytochrome *c* alone was used as a control and subtracted out from the absorbance readings of PMA-stimulated macrophages. The rate of O₂⁻ production was estimated using a molar absorption coefficient of 21.1/(mM.cm).

Glucose-stimulated insulin secretion, nitrate/nitrite determination, and cytokine treatment. Twenty mouse islets per experiment were cultured overnight in 25-mm dishes with 1.5 mL of RPMI-1640 medium/5 mM glucose + 10% FCS + 10 mM HEPES + 100 U/mL penicillin G + 100 μ g/mL streptomycin. The islet cultures were maintained at 37°C in a (Forma Scientific Inc., Marjetta,

Ohio, USA) tissue culture incubator that provided an environment of 95% O₂/5% CO₂ gas. The next morning, islets were gently transferred to RPMI-1640 medium with 3 mM glucose and incubated for 1 hour, at which time the medium was sampled for insulin measurement. The medium glucose concentration was then increased to 17 mM, and the islets were incubated for an additional hour. Once again, the medium was sampled for later insulin measurement. In certain experiments, we pretreated mouse islets with IL-1 β (0.3 ng/mL) + TNF- α (15 ng/mL) + IFN- γ (15 ng/mL) for 18 hours overnight before measuring basal and glucose-stimulated insulin secretion.

For basal nitrate/nitrite determination, 40 medium-sized islets per well were incubated in a 96-well plate in 100 μ L DMEM (without indicator) + 5 mM glucose + 0.1% FCS + 10 mM HEPES + 100 U/mL penicillin G + 100 μ g/mL streptomycin. For cytokine-stimulated nitrate/nitrite determination, 40 medium-sized islets per well were treated as described above with the addition of IL-1 β (0.3 ng/mL) + TNF- α (15 ng/mL) + IFN- γ (15 ng/mL) at time 0. After 48 hours, 100 μ L of Greiss reagent was added to each well, and the absorbance was measured at 540 nm with a microtiter plate reader (Dynatech Laboratories, Chantilly, Virginia, USA).

12-HETE assay. One hundred mouse islets per experiment were incubated in 1.5 mL of RPMI-1640 medium + 0.2% fatty acid-free BSA in the presence or absence of IL-1 β (0.3 ng/mL) + TNF- α (15 ng/mL) + IFN- γ (15 ng/mL) for 1.5 hours at 37°C. The islets and medium were extracted with 225 μ L cold ethanol (15% ethanol final concentration) and stored at -70°C for later 12-HETE determination. HETEs were extracted from the supernatant using C18 Bond Elut Columns (Varian Sample Preparations, Harbor City, California, USA). The samples in 15% ethanol were acidified to pH 3.0–3.5 with 1 N HCl. The Bond Elut columns were prepared by washing with methanol and water, and the samples were placed onto the columns by syringe. The HETEs were eluted with ethyl acetate and then quantified by RIA. This method has been validated by HPLC as described previously (18–20).

Statistics. Statistical analysis was performed with the GraphPad Instat program (GraphPad Software for Science Inc., San Diego, California, USA) for the PC. For 2 comparisons of all data, we applied the Student's *t* test. All data are expressed as mean \pm SEM.

Results

12-LO KO mice are resistant to STZ-induced diabetes. Eight male 12-LO KO mice and 12 male C57BL/6 mice received intraperitoneal injections of STZ (40 mg/kg) for 5 consecutive days between 0900 and 1000 hours. Fasting serum glucose was measured before the first injection and every 7 days thereafter. As seen in Figure 1, a statistically significant difference in mean serum glucose in the 2 groups is evident at 21 days (12-LO KO: 6.74 \pm 0.52 mM vs. C57BL/6: 10.80 \pm 1.11 mM; *P* = 0.01). By day 28, the mean serum glucose for 12-LO KO mice vs. C57BL/6 mice was 7.24 \pm 0.43 mM and 11.33 \pm 0.93 mM, respectively (*P* = 0.003). Therefore, disruption of the 12-LO gene conferred resistance to STZ-induced diabetes. We also measured fasting serum insulin levels at each time point to assess *in vivo* residual β -cell function (Figure 2). C57BL/6 mice, despite similar glucose values before STZ injections, had statistically higher fasting insulin levels than did 12-LO KO mice (164 \pm 7 nM vs. 137 \pm 16 nM, respectively; *P* = 0.01). However, the serum insulin levels in C57BL/6 mice 28 days after STZ had decreased by 90% from baseline to 17 \pm 1 nM, whereas in the 12-LO KO mice, fasting insulin levels decreased only 45% to 75 \pm 4

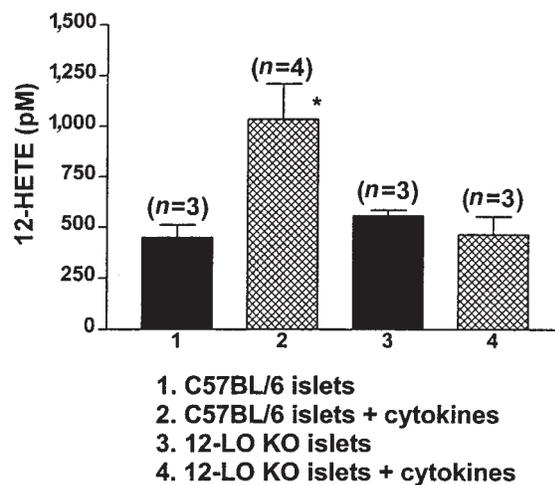


Figure 3

Differential effect of cytokines on 12-HETE production in C57BL/6 and 12-LO KO islets. C57BL/6 islets demonstrated a 2.5-fold increase in 12-HETE production in the presence of cytokines, whereas 12-HETE production in 12-LO KO islets was unchanged. The number in parenthesis represents the number of experiments performed for each condition. **P* < 0.05 vs. C57BL/6 islets.

nM. The insulin values in the C57BL/6 mice at 28 days were significantly lower than those in the 12-LO KO mice (*P* < 0.0001; Figure 2).

12-HETE generation in C57BL/6 and 12-LO KO islets. We measured cytokine-induced 12-HETE production in islets to determine whether there was any functional 12-LO protein in the 12-LO KO mice. One hundred islets per experiment were stimulated with the cytokines IL-1 β , IFN- γ , and TNF- α for 1.5 hours at 37°C. The islets were extracted into ethanol as described, and 12-HETE was measured. As seen in Figure 3, 12-LO KO islets did not generate 12-HETE in response to cytokine stimulation, whereas C57BL/6 islets produced a 2.5-fold increase, from 449 \pm 63 pM to 1,035 \pm 175 pM. This suggests that a functional cytokine-inducible 12-LO enzyme is absent in 12-LO KO islets.

Assessment of insulin secretion, cytokine-induced inhibition of insulin secretion, and NO generation in C57BL/6 and 12-LO KO islets. We purified islets from untreated C57BL/6 and 12-LO KO mice and measured basal (3 mM) and glucose-stimulated insulin secretion. Both mouse strains showed identical basal (nonstimulated) and 17 mM glucose-stimulated insulin secretion (Figure 4). 12-LO KO and C57BL/6 islets were subsequently exposed to cytokines, and glucose-stimulated insulin secretion was measured. Glucose-stimulated insulin secretion in cytokine-treated C57BL/6 islets decreased by 54% compared with untreated islets, whereas glucose-stimulated insulin secretion in 12-LO KO islets decreased only 26% compared with untreated islets (*P* < 0.03; Figure 5). Therefore, elimination of a functional 12-LO enzyme in islets leads to resistance to cytokine-induced islet dysfunction. Furthermore, the 12-LO KO islet cytokine resistance was not related to NO, as we found no difference in cytokine-stimulated nitrate/nitrite production (12-LO KO islets: 0.385 \pm

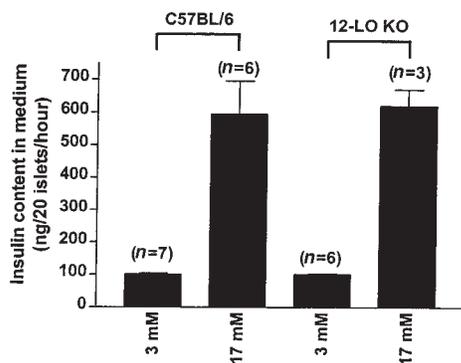


Figure 4 Basal and glucose-stimulated insulin secretion in 12-LO KO and C57BL/6 islets. Islet preparations were assessed for basal and glucose-stimulated insulin secretion as described in Methods. 12-LO KO islets secreted 102 ± 2 ng insulin/20 islets/h in the basal state and 620 ± 50 ng insulin/20 islets/h when stimulated with 17 mM glucose. In comparison, C57BL/6 islets secreted 102 ± 4 ng insulin/20 islets/h in the basal state and 595 ± 101 ng insulin/20 islets/h when stimulated with 17 mM glucose. The number in parenthesis represents the number of experiments performed for each condition.

0.125 nmol/40 islets/48 h, $n = 3$; C57BL/6 islets: 0.311 ± 0.04 nmol/40 islets/48 h, $n = 4$; $P = 0.55$).

Assessment of macrophage NO and superoxide production. Because 12-LO also is expressed in macrophages, we evaluated the possible alterations in macrophage function in 12-LO KO mice. Peritoneal macrophages were purified from C57BL/6 and 12-LO KO mice, and studies were performed to determine whether they differed in NO and superoxide production. For NO generation, we used the Griess reagent to measure nitrate/nitrite from stimulated macrophages. Figure 6 demonstrates that LPS + IFN- γ stimulation led to a 2-fold-greater increase in NO production in C57BL/6 macrophages than in 12-

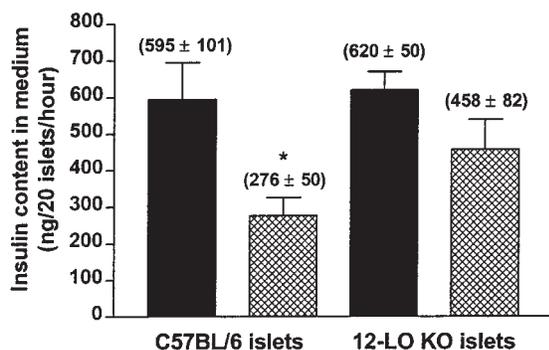


Figure 5 Effect of cytokine treatment on glucose-stimulated insulin secretion in 12-LO KO and C57BL/6 islets. Isolated islets were subjected to high-dose cytokine treatment as shown. Filled bars represent 17 mM glucose stimulation; hatched bars represent cytokine treatment (0.5 ng/mL IL-1 β + 30 ng/mL TNF- α + 30 ng/mL IFN- γ) plus 17 mM glucose stimulation. Cytokine treatment decreased glucose-stimulated insulin secretion by 26% and 54% in 12-LO KO and C57BL/6 islets, respectively. Shown is 1 representative experiment in triplicate out of 3 separate experiments. * $P < 0.03$ vs. C57BL/6 glucose-stimulated islets.

LO KO macrophages. In contrast, there was no difference in macrophage superoxide generation identified in the 2 mouse strains (Figure 7).

Discussion

It has been demonstrated previously that treatment of pancreatic islets with high-dose IL-1 β can increase 12-LO protein expression (9). Furthermore, IL-1 β has been shown to increase 12-HETE levels in rat islets by increasing the availability of free arachidonic acid (AA) (10). Given that 12-LO activation leads to hydroperoxide generation in the form of 12-HPETE, it is possible that 12-LO inhibition could decrease lipid hydroperoxide formation in islets, protect islets from cytokine-induced dysfunction, and prevent cytotoxic damage induced by STZ. We have not determined whether 12-LO gene disruption alters STZ metabolism and thereby ameliorates STZ-induced diabetes by decreasing the concentration of STZ delivered to pancreatic β cells. Nor have we formally identified whether the protective effect of 12-LO gene disruption on STZ-induced diabetes is through a pathway that prevents insulin secretory dysfunction rather than pancreatic β -cell apoptosis. However, when we performed immunohistochemistry on pancreatic sections taken from mice at 7-day intervals (data not shown), we observed preservation of islet size, number, and insulin content in 12-LO KO mice but significant decrease in these parameters in C57/BL6 mice. Therefore, the current data lend further support to the hypothesis that 12-LO inhibition may preserve islet β -cell function, and identify 12-LO as a potentially important enzyme that may be targeted for molecular inhibition.

Several studies also have addressed the question of whether lipoxygenase inhibitors can prevent pancreatic β -cell death. Zhou and colleagues recently demonstrated that thapsigargin-induced apoptosis in the MIN6 pancreatic β -cell line was significantly attenuated by lipoxygenase inhibitors. Furthermore, direct 12-HETE application in MIN6 cells reproduced the decrease in cell viability seen with thapsigargin (21). Earlier work by Rabinovitch and colleagues demonstrated that nordihydroguaiaretic acid, a lipoxygenase inhibitor, protected rat islets from cytokine-induced destruction (11). Others have demonstrated that lipoxygenase pathway products are activated in inflammatory disease states like inflammatory bowel disease (22), neuronal cells undergoing apoptosis (8), and human monocyte-mediated LDL oxidation (23). Therefore, lipoxygenase product generation may participate in a variety of inflammatory conditions.

Evidence also exists that lipoxygenase pathway products have a role in NO production. Kolb and colleagues demonstrated that nordihydroguaiaretic acid, a preferential lipoxygenase inhibitor, inhibited LPS-stimulated NO production in cultured macrophages in a dose-dependent fashion (24, 25). Our data are consistent with this report in that stimulated macrophages from 12-LO KO mice generated only 50% of the NO seen in similarly treated C57BL/6 macrophages. In contrast, AA metabolism does not seem to affect the production of superoxide by macrophages (15).

Conversely, it is tempting to speculate that high levels of NO may directly inhibit 12-LO enzyme activity through formation of iron-nitrosyl complexes similar to

mitochondrial aconitase (26, 27), as 12-LO is an iron-containing enzyme (28). However, Ma and colleagues demonstrated that low-dose application of IL-1 β (0.05 ng/mL or less) to pancreatic islets increased 12-HETE formation through an NO-dependent mechanism (10), which would imply that NO formation did not inhibit 12-LO activity in these studies. They demonstrated that NO generation increased the available pool of AA (for subsequent metabolism by 12-LO) by inhibiting enzymes that reacylate AA into membrane phospholipid. Therefore, changes in intracellular NO concentration may be one factor that regulates 12-LO product generation and, hence, modulates the redox state of the β cell through the formation of 12-HPETE.

The precise mechanism by which leukocyte 12-LO inhibition may prevent cytokine-induced pancreatic β -cell cytotoxicity is likely to be complex. 12-LO KO islets did have detectable levels of 12-HETE in the basal state; however, no appreciable increase in 12-HETE was evident after cytokine treatment. The presence of basal 12-HETE may be due to release of esterified 12-HETE from membrane phospholipid via phospholipase A₂ activation or through a yet uncharacterized isoform of 12-LO that is not induced by cytokines. Future studies will be necessary to clarify the mechanisms responsible for the basal 12-HETE in the 12-LO KO islets.

12-LO products also may act as intermediate signals in cellular inflammatory responses. 12-HETE can induce the stress-activated protein c-jun kinase (JNK) in islets and RIN m5F cells (29), and recent studies demonstrate that 12-HETE stimulates cAMP production in human fibroblasts (30). It is also possible that lipoxygenase products may activate certain forms of MAP kinases, including p38 MAP kinase, leading to downstream signaling effects.

In addition to generating hydroperoxides in pancreatic β cells and acting as intracellular signals in inflammatory responses, numerous studies have defined the role of AA and lipoxygenase metabolites in nutrient- and glucose-stimulated insulin secretion in islets. AA stimulates insulin release in islets when calcium is removed from the medium (because calcium chelates AA). Insulin secretion was independent of further AA metabolism by lipoxygenase enzymes, as it was not prevented by lipoxygenase inhibitors (31). Treatment of rat islets with *p*-hydromercuribenzoic acid, an inhibitor of arachidonyl-CoA synthase and/or lysophosphatidyl/acyl-CoA transferase (32), prevented reacylation of lysophospholipids with AA, resulting in insulin release at 1.7 mM glucose and potentiation of insulin release at 16.7 mM glucose (33). These data suggest that reacylation of fatty acids such as AA and/or 12-HETE into membrane phospholipid may play a critical role in insulin secretion. AA-induced insulin release from islets was blunted by pretreatment with phorbol ester, indicating that protein kinase C may be part of this metabolic pathway. In addition, indomethacin, a cyclooxygenase inhibitor, potentiated AA-induced insulin release (34).

Additional studies showed that 12-LO plays a role in glucose-stimulated insulin secretion (34, 35). Pretreatment of islets with LO inhibitors caused attenuation of glucose-stimulated insulin secretion, but the hydroperoxy product 12-HPETE was capable of stimulating

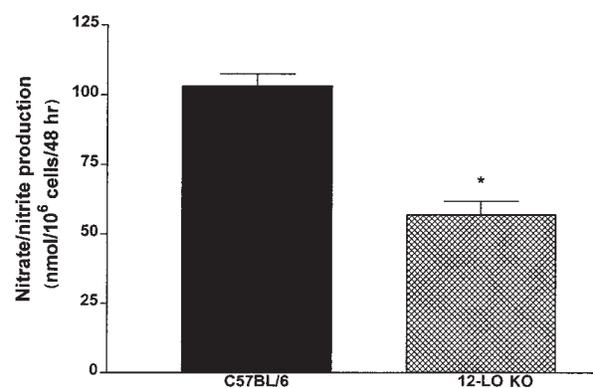


Figure 6

Nitrate/nitrite determination in 12-LO KO and C57BL/6 peritoneal macrophages. Peritoneal macrophages were purified as described in Methods and stimulated with LPS (2 μ g/mL) + IFN- γ (100 ng/mL) for 48 hours. We used Griess reagent to measure total medium nitrate/nitrite as an estimate of NO production. As shown, 12-LO KO macrophages generated ~50% of the NO of C57BL/6 macrophages under the conditions described. Six individual experiments were performed with macrophages isolated from both strains of mice. * $P < 0.0001$ vs. C57BL/6 macrophages.

insulin release. However, AA produced a greater increase in insulin release than 12-HPETE. 12-HPETE in doses ranging from 0.05 μ g/mL to 1 μ g/mL induced a 1.5-fold increase in glucose-stimulated (300 mg/dL) insulin release, whereas AA (5 μ g/mL) induced a 2-fold increase (36). 12-HETE did not alter glucose-stimulated insulin secretion in these experiments. Glucose- and AA-stimulated insulin release was further enhanced by diethylmalate, an agent that binds reduced glutathione and leads to accumulation of fatty acid hydroperoxides like 12-HPETE and epoxides.

The current results suggest that inhibition of 12-LO enzyme activity in islets does not appreciably alter basal

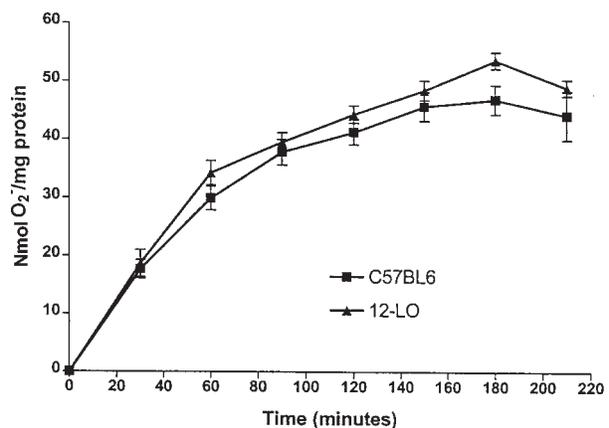


Figure 7

Generation of superoxide by 12-LO KO and C57BL/6 macrophages. Purified macrophages were treated with PMA (50 ng/mL), and superoxide generation was measured by monitoring the reduction of ferricytochrome c to ferrocyanochrome c at 550 λ over time. As shown, no difference was observed in superoxide generation. Eight individual experiments were performed with macrophages isolated from both strains of mice.

or glucose-stimulated insulin secretion. These data support previous studies showing that AA has the predominant effect on insulin secretion, whereas 12-LO products play a lesser role. Because inhibition of 12-LO does not appear to affect the overall stimulus-secretion coupling mechanism between glucose and insulin, it is possible that the 12-LO pathway provides fine tuning for AA-mediated insulin secretion.

In conclusion, 12-LO gene disruption significantly prevents STZ-induced diabetes through mechanisms that decrease pancreatic β -cell damage and reduce macrophage NO production. These results suggest that inhibition of leukocyte 12-LO in pancreatic β cells may be an effective approach to prevent cytokine-induced cytotoxicity in models of type 1 diabetes and islet transplantation rejection.

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