The role of B lymphocyte stimulator (BLyS) in systemic lupus erythematosus

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SLE, a chronic, multisystem autoimmune disorder with a broad range of symptoms, involves defective B cell selection and elimination of self-reactive B cells. B lymphocyte stimulator (BLyS), a soluble ligand of the TNF cytokine family, is a prominent factor in B cell differentiation, homeostasis, and selection. BLyS levels affect survival signals and selective apoptosis of autoantibody-producing B cells. High levels of BLyS may relax B cell selection and contribute to autoantibody production, exacerbating the SLE disease state. This review discusses the mechanism of BLyS action on B cells, its role in SLE, and specific targeting of BLyS in the treatment of SLE.

The B lymphocyte stimulator family of cytokines and receptors

B lymphocyte stimulator (BLyS, also known as the B cell–activating factor belonging to the TNF family, or BAFF; ref. 1) was identified as a novel TNF family ligand nearly 10 years ago (1–4) and has proven to be a key factor in the selection and survival of B cells. The BLyS protein is expressed by a wide variety of cell types, including monocytes, activated neutrophils, T cells, and DCs (5–7). BLyS is expressed as a cell surface protein that is then organized into a trimer, furin cleaved, and released into the circulation. Although standing levels of BLyS are constitutively generated, its expression and secretion can be potentiated by inflammatory cytokines, such as IL-2, TNF-α, and IFN-γ (6, 8, 9). A splice variant of BLyS that is inefficiently cleaved from the cell surface also exists (10, 11), and a 60-mer form of soluble BLyS (BLyS-60) has been observed in mice (12, 13).

BLyS can bind to 3 receptors: BLyS receptor 3 (BR3; also known as BAFF-R), transmembrane activator–1 and calcium modulator and cyclophilin ligand–interactor (TACI), and B cell maturation antigen (BCMA). BLyS is the sole ligand for BR3, whereas TACI and BCMA each can bind either BLyS or another TNF family ligand known as a proliferation-inducing ligand (APRIL; Figure 1 and ref. 14). These ligand-receptor interactions vary in affinity: BLyS binds more strongly to BR3 than to TACI or BCMA, whereas APRIL displays the reverse affinity hierarchy. Recent in vitro evidence suggests that BLyS-60 binds to TACI with 100-fold higher affinity than the canonical, trimeric form (14). However, the existence of a soluble BLyS-60 remains to be determined in humans.

The 3 BLyS family receptors vary in their expression patterns and levels across different B cell subsets (Figure 2). During B cell development, the capacity to bind BLyS emerges concomitant with B cell receptor (BCR) expression (15), with higher binding found on CD23+ immature B cells emerging from the bone marrow (16). BLyS binding capacity increases through the transitional (TR) stages (17–19), and the highest expression of BR3 is found in the follicular (FO) and marginal zone (MZ) subsets of B cells (16). The activation of mature B cells leads to the up- or downregulation of TACI and BR3, with the exact effects dependent on the nature of the activating stimuli. Finally, long-lived plasma cells primarily express BCMA. The various receptor expression combinations and patterns foreshadow the biological actions of the different receptor-ligand pairs.

Among newly formed and mature primary B cells, BLyS-BR3 interactions are absolutely necessary for viability, because they provide essential and nonredundant survival signals (14, 20). The requisite for BLyS-mediated BR3 survival signals is first imposed during the TR stages of B cell differentiation, concomitant with the initiation of BR3 expression. This differentiative stage is the point at which newly formed B cells leave the bone marrow and enter the circulation and spleen. As detailed below, this stage is the last major checkpoint for elimination of potentially autoreactive primary B cells prior to their maturation (21, 22). BLyS signaling via BR3 antagonizes apoptosis at these stages, thus allowing further differentiation and entrance of cells into the mature, preimmune B cell populations—the FO and MZ compartments (20, 23). Moreover, BLyS signaling via BR3 continues to be required for B cells to survive in these mature preimmune subsets.

In contrast to the actions of BR3, neither the development nor the maintenance of primary B cells requires TACI or BCMA (24, 25), as mice deficient in either of these BLyS family receptors generate normal preimmune B cell subsets (26). Instead, the data suggest that APRIL-TACI and APRIL-BCMA interactions influence antigen-experienced B cell subsets, such as activated B cells, memory B cells, and long-lived plasma cells. For example, APRIL-TACI interactions can affect both survival and Ig class-switching characteristics of B cells during T cell–dependent immune responses, and TACI-knockout mice have subtle defects in T cell–independent responses and MZ B cell numbers (27). Roles for BCMA in antigen-experienced subsets are indicated by BCMA expression on long-lived plasma cells, as well as by evidence that the numbers of such cells are reduced in BCMA-knockout mice. In addition,
APRIL promotes plasma cell survival and is produced by some resident bone marrow cell types (28). These interactions may prove to be important considerations from the perspective of therapeutic intervention, because B cell subsets that rely on TACI and BCMA and their interaction with APRIL are likely to be relatively independent of BlyS and BR3.

**BlyS, B cell survival, and self tolerance**

The importance of BlyS-BR3 interactions in primary B cell maturation and survival was first noticed in BR3-deficient and BR3-mutant mice (29–31), which display profoundly reduced peripheral B cell numbers and drastically shortened life span of cells in the TR and mature B cell subsets (16, 32, 33). Moreover, these defects could be rescued by overexpression of the antiapoptotic Bcl-2 family member Bcl-XL, suggesting that BlyS-BR3 interactions promote survival by preventing or overcoming apoptosis (34). Conversely, either exogenous administration or overexpression of BlyS rapidly led to B cell hyperplasia (2, 35) and autoimmune symptoms reminiscent of SLE and Sjögren syndrome (36). This relationship with autoimmunity has fostered intense investigation aimed at discovering how BlyS-mediated survival influences B cell tolerance.

Developing B cells that express autoreactive Ig receptors are purged at 2 checkpoints (37, 38). The first occurs in the bone marrow, where autoreactive cells either are eliminated through deletion or lose self-reactivity through a process called receptor editing (38–42). The second checkpoint is in the spleen and circulation, where TR B cells differentiate en route to becoming mature FO or MZ B cells (21, 22). In general, very strong BCR signals drive death or editing in the bone marrow, whereas less avid self-reactive clones become anergic and are then lost at the TR checkpoint (15, 43). Recent studies have shown that B cells expressing antibodies characteristic of humoral autoimmune syndromes, including anti-DNA and polyreactive specificities, are lost at the TR checkpoint (38).

The requirement for BlyS-mediated BR3 signals at the TR stages suggested that this might be the point at which BlyS affects tolerogenic processes. This has proven to be the case, because excess BlyS can cause autoreactive clones that normally die at this checkpoint to be rescued and allowed to mature (44–47). These findings indicate that anergic cells compete poorly for available BlyS and thus can survive only when BlyS levels are elevated. This ability to adjust tolerogenic stringency reflects cross-regulatory relationships between the intracellular signaling pathways used by the BCR and BR3 (48–51).

Together, these findings imply that BlyS serves 2 roles in the establishment and maintenance of primary B cells. First, it is the key regulator of primary B cell homeostasis, governing the overall numbers of mature, preimmune B cells by controlling their final step of differentiation and their life span. Second, it plays a central role in maintaining B cell tolerance by balancing the stringency of anergic cell elimination at the TR checkpoint against the need for additional preimmune B cells. Under normal conditions, this balance is achieved without substantial risk of autoreactive B cell maturation. However, in circumstances where B cell production from the bone marrow or the number of mature B cells falls outside these norms, or where BlyS levels are unusually high for extended periods, the likelihood that autoreactive cells will be allowed to mature is increased (43). Accordingly, the molecular mechanisms whereby BlyS-BR3 interactions mediate primary B cell homeostasis and the way these processes intertwine with mechanisms governing autoreactivity have been the subject of intense investigation over the last several years (43, 52, 53).

The binding of BlyS to BR3 triggers intracellular signaling pathways that antagonize apoptosis in B cells, thus promoting survival. Mounting evidence indicates that members of the NF-κB transcription factor family play central roles in these processes (54–56). Thus, BlyS signals via BR3 act primarily through activation of the so-called nonclassical NF-κB2 pathway, which facilitates upregulation of several antiapoptotic Bcl-2 family members.

The roles that BlyS family receptors and ligands play in activated and antigen-experienced subsets remain less understood. Based on the patterns of receptor expression among these cells, it is likely that TACI and BCMA will be the most prominent mediators of BlyS and APRIL activity in such cells. For example, TACI becomes elevated in B cells that have been activated through BCR or TLR stimulation, while BCMA is the prevalent receptor on long-lived plasma cells. Because APRIL binds these receptors with greater avidity than does BlyS, it is likely to exert greater influence than BlyS in these subsets. Most antigen-experienced B cell subsets appear to be independent of BlyS for survival (28, 57). Whether this indicates reliance on APRIL or instead reflects complete independence for BlyS family members remains unclear. In contrast to BR3, both TACI and BCMA signal through the classical NF-κB1 pathway, which has been generally associated with promotion of inflammatory responses and innate immunity (43, 55, 58).

**Implications for B cells and BlyS in SLE**

SLE is a chronic multisystem autoimmune disease with a broad range of clinical manifestations (59, 60). SLE can present with many different symptoms, which include photosensitive skin...
SLE patients often present with nonspecific symptoms such as fatigue, headache, Raynaud phenomenon, oral ulcers, and mild hair loss (61–63), and diagnosis may be delayed. SLE occurs primarily in women, with African and Asian ancestry groups having the highest risk (59, 64–66). Although SLE is a chronic illness that most often results in debilitated health, it can be life threatening if major organs are affected (67). It is becoming increasingly clear that accelerated atherosclerosis associated with SLE may contribute to premature mortality (68, 69).

Serologically, SLE is characterized by the presence of high titers of autoantibodies and abnormal B cell activation and differentiation (70). Autoantibodies observed in SLE include those against a variety of nuclear antigens, including dsDNA, the ribonucleoprotein (RNP) complex Ro, the RNA-binding protein La, RNPs, the RNA molecule/protein complex Sm, the C1 complement component subunit C1q, and phospholipids. Complement levels are often reduced as a consequence of active disease, but in rare cases, they may be caused by a congenital complement deficiency (71). Autoantibodies have been detected 2–9 years before the onset of clinical features of SLE (72). However, the appearance of anti-dsDNA, anti-Sm, and anti-RNP antibodies suggest a peak in the intensity of the autoimmune response prior to the onset of clinical illness (73).
### Table 1
B cell–targeted therapies for SLE

<table>
<thead>
<tr>
<th>Therapy</th>
<th>Example</th>
<th>Class of therapy</th>
<th>Mechanism of action</th>
<th>B cell lineages affected</th>
<th>Clinical trials</th>
<th>Clinical outcomes</th>
<th>Refs.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Anti-BR3</td>
<td>NA</td>
<td>Mouse mAb</td>
<td>Binds BR3 receptor to block BLyS</td>
<td>NA</td>
<td>None</td>
<td>Preclinical data show reduction of B cell counts</td>
<td>(102)</td>
</tr>
<tr>
<td>BLyS targeted</td>
<td>BR3-Fc</td>
<td>Recombinant FP</td>
<td>Binds BLyS to prevent interaction with receptor on B cells</td>
<td>Immature B cells to short-lived plasma cells</td>
<td>Phase I (N = 56) in RA</td>
<td>Well tolerated; efficacy NA</td>
<td>(97)</td>
</tr>
<tr>
<td>BLyS targeted</td>
<td>AMG-623</td>
<td>Peptide-Fc FP</td>
<td>Binds BLyS to prevent interaction with receptor on B cells</td>
<td>Immature B cells to short-lived plasma cells</td>
<td>Phase 1a (N = 54), phase 1b (N = 63) in SLE</td>
<td>Well tolerated; efficacy NA</td>
<td>(98)</td>
</tr>
<tr>
<td>BLyS targeted</td>
<td>Belimumab</td>
<td>Fully human mAb</td>
<td>Binds BLyS to prevent interaction with receptor on B cells</td>
<td>Immature B cells to short-lived plasma cells</td>
<td>Phase II RCT plus open label extended (N = 449) in SLE</td>
<td>Serologically active patients show significant clinical improvement; overall endpoint not met</td>
<td>(91)</td>
</tr>
<tr>
<td>BLyS/APRIL targeted</td>
<td>Atacicept</td>
<td>Recombinant FP</td>
<td>Binds both BLyS and APRIL</td>
<td>Immature B cells to long-lived plasma cells</td>
<td>Phase IIb RCT (N = 47), phase II/III RCT (N = 200)</td>
<td>Patients with moderate SLE show clinical improvement compared with placebo; atacicept/MMF treatment causes significant infection risk</td>
<td>(95, 96)</td>
</tr>
<tr>
<td>Anti-CD20</td>
<td>Rituximab</td>
<td>Chimeric mAb</td>
<td>Binds to CD20, leading to B cell apoptosis, ADCC, C-mediated lysis, and enhanced CD20+ lymphocyte clearance</td>
<td>Pre-B cells through memory cells</td>
<td>Open label (N = 24), prospective (N = 11), phase II/III (N = 18), phase II/III RCT (N = 257) in SLE</td>
<td>Patients show significant clinical improvement; overall endpoint not met</td>
<td>(103–108)</td>
</tr>
<tr>
<td>Anti-CD20</td>
<td>Ocrelizumab</td>
<td>Humanized mAb</td>
<td>Binds to CD20, leading to B cell apoptosis, ADCC, C-mediated lysis, and enhanced CD20+ lymphocyte clearance</td>
<td>Pre-B cells through memory cells</td>
<td>Phase VII (N = 237) in RA</td>
<td>Well tolerated, B cell depletion comparable to rituximab</td>
<td>(109)</td>
</tr>
<tr>
<td>Anti-CD20</td>
<td>Ofatumumab</td>
<td>Fully human mAb</td>
<td>Binds to CD20, leading to B cell apoptosis, ADCC, C-mediated lysis, and enhanced CD20+ lymphocyte clearance</td>
<td>Pre-B cells through memory cells</td>
<td>Phase II RCT (N = 226) in SLE</td>
<td>Early results show significant clinical improvement</td>
<td>(110)</td>
</tr>
<tr>
<td>Anti-CD22</td>
<td>Epratuzumab</td>
<td>Fully human mAb</td>
<td>Binds to CD22, leading to B cell apoptosis</td>
<td>Pro-B cells through memory cells</td>
<td>Open label (N = 14)</td>
<td>Clinical improvement in all patients tested</td>
<td>(111, 112)</td>
</tr>
</tbody>
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*A* Phase II trial for lupus nephritis that included combination treatment with atacicept and mycophenolate mofetil (MMF) was suspended because of a high risk of severe infections. 

*B* The FDA issued a safety alert regarding treatment with rituximab when 2 incidences of progressive multifocal leukoencephalopathy were seen in lupus patients. 

*C* Two clinical trials for epratuzumab in SLE have been prematurely terminated because of manufacturing quality assurance failures. ADCC, Ab-dependent cellular cytotoxicity; FP, fusion protein; RCT, randomized controlled trial.
Numerous factors, both environmental and genetic, contribute to and trigger SLE development and autoantibody production. B cells and BLyS have recently emerged as important factors in the pathology of SLE, regardless of triggering factors. As noted above, BLyS-BR3 binding and signaling is critical for B cell survival or selection at the TR checkpoint. If this checkpoint is impaired or compromised in any way, it could allow moderately autoreactive or polyreactive B cells to survive selection instead of undergoing elimination by apoptosis. A primary source of BLyS secretion is DCs, which are often located in mucosal, glandular, and cutaneous regions (74). Because much of the pathology of SLE is dermal and oral, it is reasonable to speculate that overactivity of the DCs may contribute to elevated serum levels of BLyS, and thus to SLE disease flares.

Elevated serum levels of soluble BLyS protein have been observed in patients with autoimmune disease, including those with SLE; evidence has shown that moderately autoreactive B cells in the spleen require higher concentrations of BLyS to survive the TR checkpoint than do B cells that produce non-self-reactive antibodies (36, 75). Additionally, mice treated with exogenous BLyS have shown increased numbers of antichromatin B cells, which suggests that high levels of BLyS contribute to elevated production of antDNA antibodies, which are common in SLE patients (76). Transgenic mice that overexpress the BLyS protein develop autoimmune disorders similar to SLE and Sjogren syndrome (36). On the other hand, mouse models have confirmed that deletion of either BLyS or BR3 severely impairs B cell development beyond the TR developmental stages (30, 31, 36, 77). These data indicate that elevated BLyS levels may correlate with SLE disease activity. In addition, the development of class-switched autoantibodies in BLyS-transgenic mice appears to be T cell independent (78). This finding is interesting in light of the production of autoreactive B cells in human autoimmune disease; it suggests that there may be subsets of SLE or RA patients who develop autoantibodies in a T cell–independent manner and may benefit from BLyS-targeted therapy.

SLE has also been associated with defective clearance of apoptotic bodies. Apoptosis is a continual process that results in the release of subcellular debris and intracellular antigens, such as DNA, into the immune system. The clearing of apoptotic bodies is critical to prevent 2 events: first, uptake of the bodies by macrophages, which would present antigens to T and B cells, thus leading to an immune reaction against intracellular components (79); and second, presentation of intracellular TLR ligands to innate immune cells. The most common intracellular self-antigens are ssDNA, dsDNA, dsRNA, and RNP, among others, and each of these activates TLRs, resulting in elevated expression of BLyS (80). It is reasonable to speculate that increased levels of BLyS resulting from the presentation of self-antigens to innate immune cells—which express BLyS upon antigenic stimulation—may further amplify the autoantibody production by B cells and exacerbate the SLE disease state. Moreover, IFNs are often profoundly dysregulated in SLE, and both IFN-α and IFN-γ have been shown to induce BLyS expression (5, 81, 82). A number of gene expression profile analyses have identified an IFN-related signature in SLE patients; more recently, SLE disease activity, as measured by Systemic Lupus Erythematosus Disease Activity Index–2K (SLEDAI-2K), was found to correlate with a strong IFN signature in a group of SLE patients (83, 84). Based on these data, it is tempting to speculate a potential cooperative action of BLyS and IFNs in the etiology of SLE. These observations suggest that BLyS may figure prominently in the development of autoimmune disease, particularly SLE, and may be a valid target for SLE therapy.

### BLyS-targeted therapy for SLE

Elevated BLyS levels have been implicated in abnormal B cell development, including (but not limited to) autoantibody production, lymphadenopathy development (75), and lymphomas (85). By specifically antagonizing the modulatory protein, BLyS-targeted agents provide an effective way to control B cell activity (Table 1). BLyS-specific targeted therapy specifically affects early-stage B cells in the periphery without affecting late-stage compartments, such as memory or bone marrow plasma cells, and without compromising the immune system (86).

**Belimumab.** Belimumab is a fully human monoclonal Ab (IgG1) that binds soluble BLyS (86) and inhibits its binding to TACI, BCMA, and BR3 (Table 1). The specificity and affinity of belimumab for BLyS suggests that it may decrease B cell survival that results from an abundance of BLyS. Preclinical data demonstrate a role for belimumab in inhibiting BLyS; the administration of belimumab was found to inhibit the effects of BLyS in mice expressing the exogenous human protein, including decreases in spleen weight and serum IgA levels (86). This effect may stem, at least in part, from an attenuation of class switching in autoreactive B cells. Recent nonclinical data have shown that anti-BLyS treatment appears to leave secondary immunity intact. A study of anti-BLyS treatment in mice determined that memory B cells, long-lived plasma cells, and secondary immune responses transiently rose, but remained whole, during and after exposure (87, 88). IgG-bearing memory B cells and natural Ab–secreting B cells were insensitive to anti-BLyS treatment, which suggests that they are BLyS independent and may be manipulated separately (57). A common phenomenon associated with BLyS-targeted therapies is the early transient rise in memory B cell numbers, which return to normal levels within 2 weeks after treatment (89).

The safety and efficacy of belimumab in SLE patients has been studied in a phase I randomized controlled clinical trial (90). In this dose-ranging study, belimumab treatment resulted in significantly greater percent reductions in CD20+ B cells than did placebo, consistent with the mechanism of action of this Ab. This reduction was observed in patients treated with 1 or 2 doses of belimumab versus placebo-treated patients. A similar percent decrease in serum IgS was observed in belimumab-treated patients compared with placebo-treated patients; however, this decrease did not reach statistical significance. Overall, there was no difference in disease activity—as assessed by Safety of Estrogens in Lupus Erythematosus: National Assessment–SLEDAI (SELENA-SLEDAI) score—between the 2 treatment groups, but this may be because of the short term of the trial. Belimumab was well tolerated in the phase I study, with the majority of adverse events (AEs) being mild to moderate in severity, and the incidence of AEs similar in the placebo and belimumab treatment groups. In addition, there was no increase observed in the number of infections in belimumab-treated patients versus patients given placebo (37% and 62% of the study population, respectively).

Belimumab appears to have a latency period before SLE disease improvement is seen, and clinical response was most evident in a subgroup of serologically active patients (91); each of these observations may account for the phase II trial not meeting its primary endpoint (Table 1). The long onset of efficacy for belimumab may be due to a slow off-time for BLyS at BR3, coupled with highly occupied receptors in SLE patients. A study of SLE patients showed reduced levels of available free BR3 on B cells in comparison with healthy controls (92). The reduced amount of
available BR3 resulted in continuous receptor-ligand engagement. The slow off-time for BlyS at BR3, coupled with the abundance of BlyS, correlated with an increase in disease activity (92). BlyS also has a slow turnover rate, which may explain why there is a lag time or slower rate of disease response with belimumab treatment. The safety and efficacy of belimumab in SLE is currently being further validated in 2 large, global, phase III randomized controlled trials (BLISS S2 and BLISS 76; http://www.clinicaltrials.gov; trial nos. NCT00424476 and NCT00410384, respectively).

Atacicept. Atacicept (TACI-Ig) is a soluble, recombinant fusion protein of the human IgG1 Fc and the extracellular domain of the TACI receptor that binds BlyS and APRIL and inhibits their action on B cells (Table 1). Similar to BlyS blockade, atacicept administration in a murine model of SLE led to delayed onset in SLE disease progression and reductions in B cell populations, spleen size, and DC activation (93, 94). However, in parallel to the anti-BlyS data above, it appears that TACI blockade reduces IgM-producing, but not IgG-producing, plasma cells. Whether this same effect occurs in humans remains to be seen.

In a dose-escalating phase I clinical trial (95), patients with moderate SLE showed a trend toward clinical improvement, as assessed by SELENA-SLEDAI scores, after treatment with atacicept. The small patient number (n = 12) precludes a definitive conclusion regarding the effect of atacicept on disease activity, although dose-dependent reductions in mature and total B cell levels and IgG were observed. The reductions in B cells were sustained for up to 43 days in the single-dose group and up to 64 days in the repeated-dose group. Atacicept was well tolerated in this trial, with no significant differences in AE frequency or type observed between placebo and treatment groups. Although the number of infection-related events was similar in placebo and atacicept groups in the phase I trial, a recent phase II trial of atacicept in lupus nephritis was stopped because of a high risk of severe infections (96). A new phase II/III trial of atacicept in generalized SLE is currently open and recruiting patients (http://www.clinicaltrials.gov; trial no. NCT00624338).

Additional BlyS-targeted therapies. New biologic therapies targeted against BlyS are presently in development. These include BR3-Fc, AMG-623, and anti-BR3 Ab (Table 1). Of these, only BR3-Fc and AMG-623 have begun clinical trials, and while both appear to be well tolerated by patients, their efficacy in treating SLE and improving disease activity remains unknown (97, 98).

Other B cell–targeted therapies in SLE. Several therapies with a mechanism of action that targets B cells directly, in contrast to targeting BlyS, have also been tested in SLE patients and other autoimmune patients. Rituximab, ofatumumab, and epratuzumab have all demonstrated biological effect in SLE, while ocrelizumab has shown efficacy in RA (Table 1). Each of these has been shown to deplete the total population of peripheral B cells with less specificity than BlyS-targeted drugs. Furthermore, the depleted state of B cells may persist for up to 10 months after treatment is terminated (99).

It is reasonable to speculate that reducing the number of B cells with anti-CD20 Ab therapy may effectively elevate the concentration of free BlyS, leading to increased incidence of autoreactive B cells surviving selection (43). A recent study of non-SLE B cell disorders showed that treatment with rituximab resulted in a 5-fold increase in serum BlyS levels (P < 0.001; ref. 100). In addition, recent clinical data reflecting the measurement of BlyS levels and autoreactive Abs in SLE during rituximab therapy (n = 25) showed that patients with higher BlyS levels at baseline were more likely to harbor anti-Ro and anti-RNP/Sm Abs after treatment; patients in this subgroup were significantly more likely to experience an SLE disease flare within 1 year of cessation of therapy (101). In this study, patients with lower BlyS levels prior to anti-CD20 therapy fared better in the long term than did their higher-BlyS counterparts (101). These data suggest that certain patients, especially those with high baseline BlyS serum levels, may not be the ideal candidates for rituximab therapy.

Conclusion
The definitive role of BlyS in SLE continues to be assessed. However, the elevated serum BlyS levels frequently observed in SLE patients, coupled with murine models of BlyS overexpression, suggest a significant role in the autoimmune process. The use of a fully human monoclonal Ab that binds soluble BlyS (i.e., belimumab) in serologically active SLE patients has resulted in reductions in disease activity and B cell populations without an observed increased risk of infection or major AEs. The continued clinical investigations into BlyS-targeted SLE therapy, as well as B cell deletion therapies, will aid in defining the role of BlyS and B cells in the development and perpetuation of SLE, as well as their place in future therapies for this heterogeneous patient population.

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