



The MEF2D transcription factor mediates stress-dependent cardiac remodeling in mice

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The adult heart responds to excessive neurohumoral signaling and workload by a pathological growth response characterized by hypertrophy of cardiomyocytes and activation of a fetal program of cardiac gene expression. These responses culminate in diminished pump function, ventricular dilatation, wall thinning, and fibrosis, and can result in sudden death. Myocyte enhancer factor-2 (MEF2) transcription factors serve as targets of the signaling pathways that drive pathological cardiac remodeling, but the requirement for MEF2 factors in the progression of heart disease in vivo has not been determined. MEF2A and MEF2D are the primary MEF2 factors expressed in the adult heart. To specifically determine the role of MEF2D in pathological cardiac remodeling, we generated mice with a conditional MEF2D allele. MEF2D-null mice were viable, but were resistant to cardiac hypertrophy, fetal gene activation, and fibrosis in response to pressure overload and β -chronic adrenergic stimulation. Furthermore, we show in a transgenic mouse model that forced overexpression of MEF2D was sufficient to drive the fetal gene program and pathological remodeling of the heart. These results reveal a unique and important function for MEF2D in stress-dependent cardiac growth and reprogramming of gene expression in the adult heart.

Introduction

Acute or chronic injury to the adult heart activates a pathological response characterized by hypertrophic growth of cardiomyocytes, assembly of additional sarcomeres to enhance contractility, and activation of a fetal cardiac gene program, ultimately causing ventricular wall thinning, chamber dilation, heart failure, and sudden death (1–3). A variety of abnormalities cause pathological hypertrophy and heart failure, including hypertension, myocardial infarction, endocrine disorders, and inherited mutations in structural and contractile proteins of cardiomyocytes. These insults have been shown to activate a complex set of interwoven signaling pathways that culminate in the nucleus to activate various transcription factors, including nuclear factor of activated T cells (NFAT), GATA, and myocyte enhancer factor-2 (MEF2) factors, that drive the expression of fetal cardiac and stress response genes (3, 4). There has been great interest in deciphering the molecular mechanisms responsible for transcriptional reprogramming of the failing heart and in developing “transcriptional” therapies for enhancing cardiac function (4, 5).

Members of the MEF2 family of MADS (MCM1, agamous, deficiens, serum response factor) box transcription factors function as stress-dependent regulators of gene expression and have been implicated in multiple aspects of striated muscle development and disease (6). The 4 vertebrate MEF2 factors, MEF2A, -B, -C, and -D, display overlapping expression patterns in embryonic and adult tissues (7–9), and loss-of-function studies in mice have revealed distinct functions for the *Mef2a* and *Mef2c* genes in the develop-

ing heart. Mice lacking *Mef2a* survive until the perinatal period when they succumb to lethal cardiac arrhythmias accompanied by abnormalities in expression of contractile protein and metabolic genes (10). Mice lacking *Mef2c* die during early embryogenesis from a spectrum of cardiovascular defects (11). However, despite the key roles of MEF2 factors as regulators of cardiac gene expression during development, relatively little is known of their functions in the adult heart.

Using transgenic mice harboring a MEF2-dependent reporter gene, we showed previously that hypertrophic stimuli augment MEF2 activity in the adult heart (12). Stress-activated signaling pathways that drive pathological cardiac remodeling stimulate the transcriptional activity of MEF2 through multiple mechanisms. Activation of calcium/calmodulin-dependent protein kinase and protein kinase D stimulates MEF2 activity by promoting the phosphorylation and nuclear export of class II histone deacetylases, which function as MEF2 corepressors (13, 14). MAPK signaling leads to the phosphorylation of MEF2 and stimulation of transcriptional activity, and calcineurin promotes MEF2 activity through multiple mechanisms (15–17).

The major MEF2 isoforms in the adult heart are MEF2A and -D, which form heterodimers (10, 18). The lethal phenotype of *Mef2a*-null mice indicates that MEF2D alone cannot support normal postnatal cardiac function (10). Whether MEF2A can support cardiac function in the absence of MEF2D and whether MEF2D may also be essential in the adult heart has not been determined.

To define the potential functions of MEF2D in the heart, we generated mice with a *Mef2d* loss-of-function mutation. In contrast to the lethal cardiac phenotypes resulting from genetic deletion of *Mef2a* or *Mef2c*, we show that mice lacking *Mef2d* are viable, but display an impaired response to stress signals that normally lead to cardiac hypertrophy, fibrosis, and fetal gene activation. Conversely, forced overexpression of MEF2D in the heart is sufficient to evoke severe cardiomyopathy. These findings demonstrate that MEF2D

Nonstandard abbreviations used: FS, fractional shortening; HW/TL, heart weight/tibia length; ISO, isoproterenol; LVlDd, LV end-diastolic diameter; LVlDs, LV end-systolic diameter; MEF2, myocyte enhancer factor-2; MHC, myosin heavy chain; NFAT, nuclear factor of activated T cells; TAC, thoracic aortic constriction.

Conflict of interest: The authors have declared that no conflict of interest exists.

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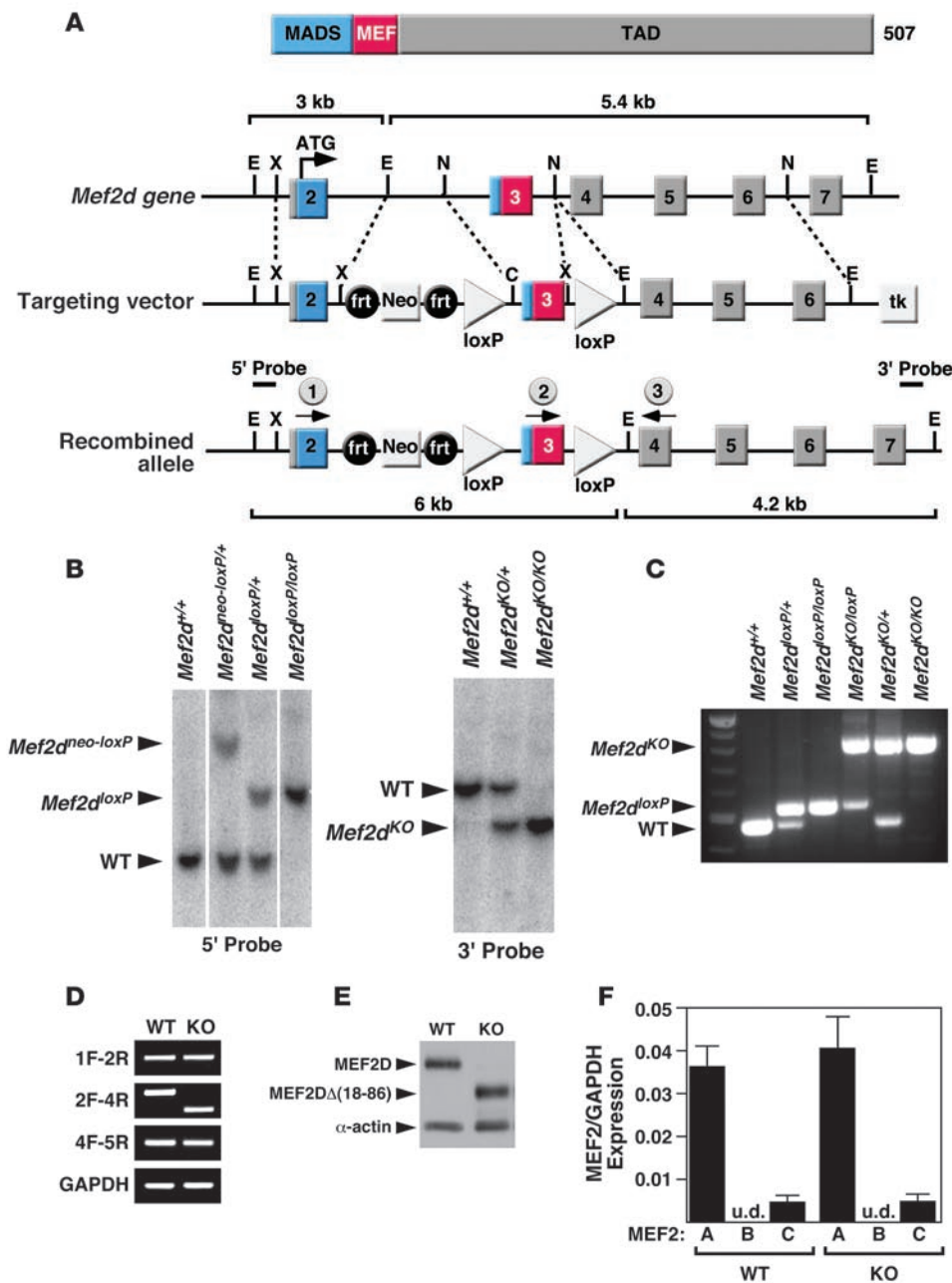


Figure 1

Generation of mice with a conditional *Mef2d* mutation. (A) Schematic representation of the mouse *Mef2d* locus and targeting strategy. Positions of 3' and 5' probes used for Southern blots are shown. The positions of the PCR primers used for genotyping mutant alleles are marked with arrows (circles labeled 1, 2, and 3). C, ClaI; frt, FLP recombinase target; E, EcoRI; TAD, transactivation domain; MADS, MCM1, agamous, deficiens, serum response factor; N, NcoI; X, XhoI. (B) Southern blot analysis of *Mef2d* mutant alleles. Genomic DNA was digested with EcoRI and hybridized to a 5' probe in the left panel and to a 3' probe in the right panel. WT, wild-type allele; *Mef2d*^{neo-loxP}, conditional allele; *Mef2d*^{loxP}, conditional allele with the Neo cassette removed; *Mef2d*^{KO}, null allele. (C) PCR genotyping to distinguish different *Mef2d* alleles. The positions of the primers that produce these PCR products are labeled (1, 2, and 3) and circled in A. All 3 primers were added to the PCR reactions and the PCR products were loaded into lanes 2–7. (D) Expression of wild-type and mutant *Mef2d* detected by RT-PCR. *Mef2d* mutant allele lacks exon 3, which encodes the MADS- and MEF2-specific domains. GAPDH was used as a loading control. Labels on the left side of the panel indicate exon location and direction of primers used for RT-PCR. (E) Western blot analysis to detect WT and mutant MEF2D (KO) proteins. The mutant MEF2D protein is truncated due to deletion of exon 3. α -Actin protein was used as a loading control. (F) Expression level of *Mef2d* detected by quantitative PCR. Error bars indicate \pm SEM. u.d., undetectable.

plays a key role in mediating stress-dependent gene expression in the adult heart.

Results

Generation of *Mef2d*-null mice. Because *Mef2a*- and *Mef2c*-null mice are not viable (10, 11), we generated a conditional *Mef2d*-null allele, called *Mef2d*^{loxP} in order to determine the functions of *Mef2d* in the adult heart. The conditional allele contained loxP sites flanking the third exon, which encodes the MADS- and MEF2-specific domains that mediate DNA binding, dimerization, and cofactor interactions (Figure 1A). Deletion of the corresponding region between the loxP sites of *Mef2a* or *Mef2c* results in a null allele (10, 11).

To determine the complete loss-of-function phenotype of *Mef2d*, we initially deleted the gene using a *Meox2*-Cre transgene, which

expresses Cre recombinase in epiblast-derived tissues as early as embryonic day 5 (19). Unexpectedly, mice homozygous for this deletion were viable and showed no obvious phenotypic abnormalities. Complete deletion of the genomic region between the 2 loxP sites was confirmed by Southern blot and PCR of genomic DNA (Figure 1, B and C). By RT-PCR of RNA from adult heart, we found that exon 2 was spliced to exon 4 in the mutant allele, resulting in an in-frame deletion within the coding region of the *Mef2d* mRNA (Figure 1D). Western blot analysis of cardiac extracts confirmed that the mutant MEF2D protein lacking the MADS and MEF2 domains was expressed at a level comparable to the wild-type protein (Figure 1E).

To confirm that the residual MEF2 protein did not function as a dominant-negative mutant, we coexpressed it with the wild-type

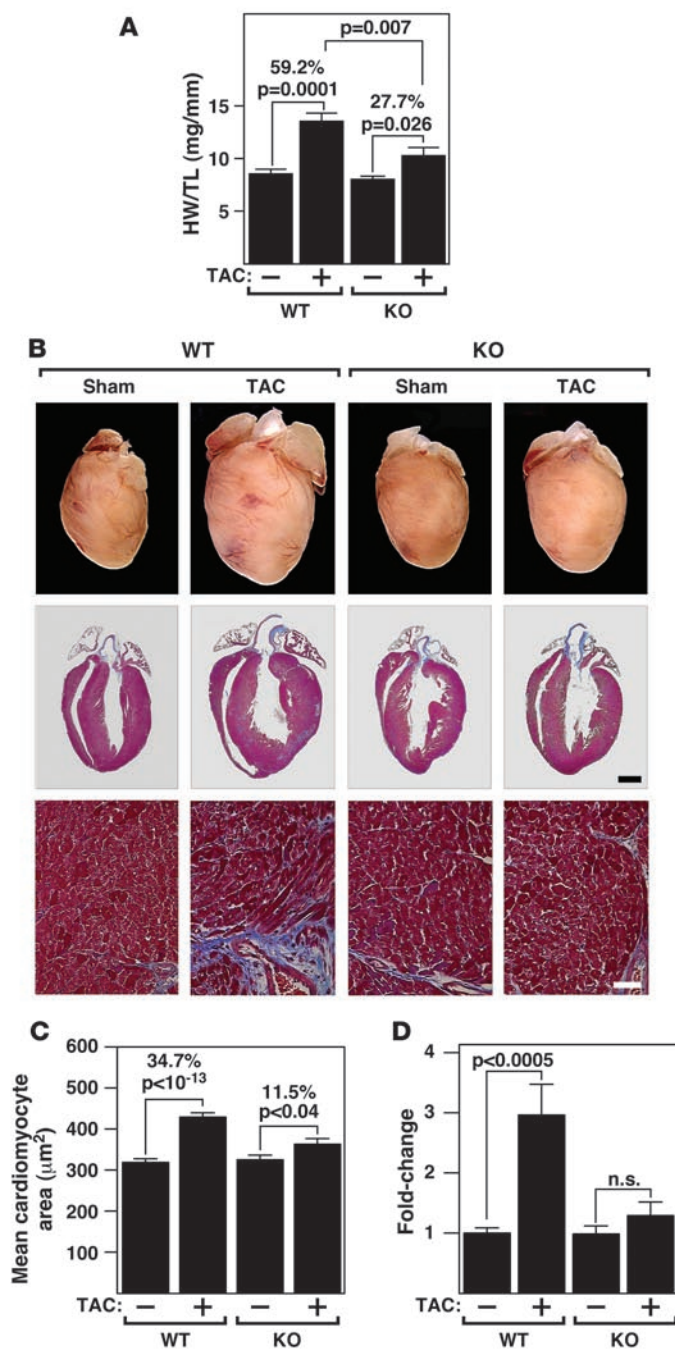


Figure 2

Blunted hypertrophy of *Mef2d* mutant mice following TAC. (A) HW/TL ratios (\pm SEM) of WT and *Mef2d* mutant mice were determined 21 days after TAC. (B) Hearts from WT and *Mef2d* mutant mice subjected to either a sham operation or pressure overload (TAC) are shown at the top. Histological sections stained with Masson's trichrome are shown on the bottom. Masson's trichrome staining of WT and *Mef2d* mutant hearts indicates lack of fibrosis in *Mef2d*^{-/-} hearts in response to pressure overload by TAC. Scale bars: 1 mm (middle panel); 40 μ m (bottom panel). (C) Mean cross-sectional area of cardiomyocytes (\pm SEM) in WT and *Mef2d* mutant mice was measured 21 days after TAC. (D) Using morphometric analysis of picrosirius red stained heart sections, the amount of myocardial fibrosis was assessed. Fibrosis was apparent in the hearts of WT mice following TAC. In contrast, virtually no fibrosis was detected in the hearts of *Mef2d* mutant mice. Values indicate fold changes of fibrosis in each group compared with a group of sham-operated WT mice (\pm SEM).

expression in *Mef2d* mutant hearts showed no compensatory upregulation of these genes (Figure 1F).

Mef2d-null mice are resistant to cardiac remodeling and fibrosis in response to pressure overload. To investigate whether MEF2D was required for pathological remodeling of the heart in response to pressure overload, we performed thoracic aortic constriction (TAC) on adult mice. As shown in Figure 2A, the hearts of wild-type and *Mef2d*^{-/-} mice were comparable in size at baseline. Twenty-one days following TAC, wild-type mice showed a 59.2% increase in heart weight/tibia length (HW/TL). In contrast, *Mef2d* mutant mice showed only a 27.7% increase in HW/TL (Figure 2A). Based on histological sections, less cardiac remodeling was observed in *Mef2d* mutant mice compared with wild-type mice (Figure 2B). Consistent with these findings, following pressure overload, we observed a 34.7% \pm 0.25% increase in cross-sectional area of cardiomyocytes in wild-type mice, in comparison to only a 11.5% \pm 0.40% increase in mutant mice (Figure 2C).

Pressure overload hypertrophy in wild-type mice was accompanied by extensive fibrosis of the ventricular wall as detected by Masson's trichrome staining (Figure 2B). Remarkably, there was virtually no ventricular fibrosis in *Mef2d* mutant mice. These findings were corroborated by morphometric analysis of picrosirius red stained sections of wild-type and mutant hearts showing that *Mef2d* mutant mice had much less fibrosis after pressure overload compared with wild-type mice (Figure 2D).

Functional analysis of hearts of Mef2d-null mice. The response to cardiac stress of *Mef2d* mice was assessed by echocardiography. At baseline, there were no substantial differences seen in the LV end-diastolic diameter (LVIDd) or heart rate (HR), and only minimal differences seen in the LV end-systolic diameter (LVIDs) and fractional shortening (FS) in wild-type and mutant *Mef2d* mice (Table 1 and Supplemental Table 1). Three weeks after TAC surgery, wild-type mice showed a dramatic increase in LVIDd accompanied by a pronounced reduction in cardiac contractility, as indicated by a decreased FS. In contrast, *Mef2d* mutant animals were largely resistant to LV dilation and its concomitant decrease in contractility (Figure 3). Additionally, compared with *Mef2d* mutant animals, wild-type animals experienced a significant reduction in HR following TAC, which is indicative of the pathological state of the heart. These data demonstrate that in response to pressure

protein in transfected COS-7 cells and monitored expression of a MEF2-dependent luciferase reporter. As shown in Supplemental Figure 1 (supplemental material available online with this article; doi:10.1172/JCI3255DS1), the MEF2 Δ (18-86) protein was devoid of transcriptional activity and showed no dominant-negative effect on wild-type MEF2A protein.

We analyzed numerous adult tissues, including skeletal muscle, heart, liver, thymus, spleen, and brain of *Mef2d* mutant mice and observed no histological abnormalities. *Mef2d* mutant mice also appeared normal by other criteria, including behavior, weight, fertility, and lifespan. We conclude that MEF2D is not required for viability in mice. Quantification of *Mef2a* and *Mef2c* mRNA

**Table 1**

Echocardiographic data (\pm SEM) indicate less LV remodeling in the *Mef2d* mutant (KO) mice in response to TAC compared with WT littermates

	LVIDd (mm)	LVIDs (mm)	FS (%)	HR (min^{-1})	AWd (mm)	PWd (mm)
WT sham	2.10 \pm 0.06	0.28 \pm 0.03	83.98 \pm 0.81	681.70 \pm 10.50	0.83 \pm 0.02	0.60 \pm 0.03
WT TAC	3.41 \pm 0.36	2.09 \pm 0.17	41.63 \pm 4.40	587.56 \pm 15.95	0.69 \pm 0.08	0.77 \pm 0.03
KO sham	2.14 \pm 0.05	0.50 \pm 0.04	75.62 \pm 1.68	668.49 \pm 5.60	0.80 \pm 0.05	0.70 \pm 0.05
KO TAC	2.87 \pm 0.17	0.98 \pm 0.21	65.03 \pm 4.46	685.14 \pm 26.71	0.95 \pm 0.04	0.88 \pm 0.02

$n = 5-7$ per group. All values are shown as mean \pm SEM. AWd, anterior wall thickness in diastole; HR, heart rate; PWd, posterior wall thickness in diastole.

overload *Mef2d* mutant hearts are resistant to cardiac remodeling and functional deterioration.

Fetal gene activation is blunted in Mef2d-null mice in response to pressure overload. MEF2D was required not only for hypertrophic growth and fibrosis in response to pressure overload, but was also essential for fetal gene activation, a prototypical consequence of cardiac stress. As shown in Figure 4, upregulation of the hypertrophic gene markers atrial natriuretic peptide (*ANP*), brain natriuretic peptide (*BNP*), myosin, heavy polypeptide 7, cardiac muscle, β (*Myh7*), and actin, $\alpha 1$, skeletal muscle (*Acta1*) was severely blunted in mutant mice. Also, induction of connective tissue growth factor (*Ctgf*), procollagen, type I, $\alpha 2$ (*Col1a2*), and procollagen, type III, $\alpha 1$ (*Col3a1*), which are upregulated during cardiac fibrosis, was compromised in *Mef2d*^{-/-} mice. Baseline expression of several fetal cardiac genes was also diminished in *Mef2d*^{-/-} mice, suggesting that MEF2D is required for their normal expression in the absence of stress (Figure 4 and Supplemental Table 1). These data are consistent with Masson's trichrome and picosirius red staining, and support the conclusion that cardiac hypertrophy and fibrosis induced by pressure overload are suppressed in *Mef2d*^{-/-} mice.

We observed a decrease in *Mef2a* and *Mef2d* expression level following TAC; however, previous reports showed that MEF2 activity is increased following cardiac remodeling (12, 20). We believe that the decrease in MEF2 expression may represent a compensatory mechanism to diminish pathological signaling via MEF2 in the heart.

Mef2d-null mice are resistant to cardiac remodeling in response to β -adrenergic stimulation. We also investigated whether MEF2D was necessary for cardiac remodeling in response to chronic β -adrenergic stimulation. Isoproterenol (ISO), a β -adrenergic receptor agonist, was administered over 7 days to mice using osmotic minipumps (8.8 mg/kg/day). Compared with wild-type mice, which showed a 25.9% increase in HW/TL ($P = 0.0008$), mutant mice

showed an insignificant increase in HW/TL (Figure 5, A and B). This finding suggests that MEF2D acts within signaling pathways activated not only by pressure overload, but also β -adrenergic stimulation.

Echocardiography confirmed that ISO administration resulted in a more pronounced increase in LV wall thickness in wild-type mice compared with *Mef2d* mutant mice, indicative of cardiac hypertrophy (Table 2). In addition, the wild-type animals experienced enhanced LV remodeling in

response to ISO, as shown by more LV dilation and a decrease in contractility (Table 2 and Figure 5C).

Forced expression of MEF2D causes severe cardiomyopathy. To determine whether MEF2D is sufficient to induce pathological cardiac remodeling, we generated transgenic mice in which the α -myosin heavy chain (α -MHC) transgene directed cardiac-specific expression of MEF2D. Three independent stable transgenic mouse lines (lines 1, 2, and 3) were established that, based on Western blot analysis, expressed MEF2D in the heart approximately 18-, 15-, and 2.5-fold higher, respectively, than the wild-type animals. We focused on transgenic line 2, which showed clear signs of cardiomyopathy – extensive fibrosis, atrial enlargement, and thrombi as well as severely congested lungs and liver – at 8 weeks of age.

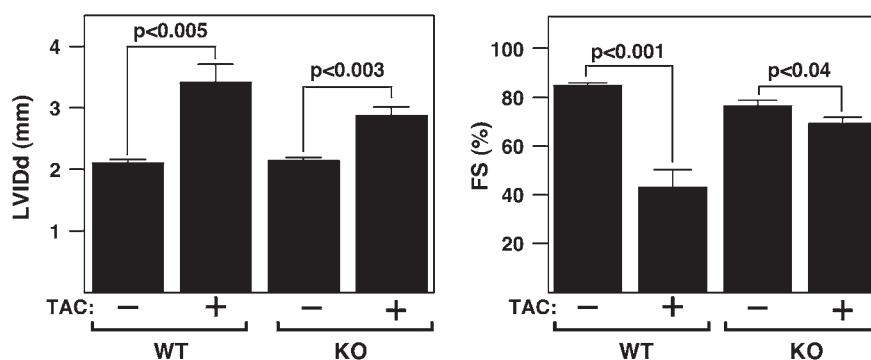
Fetal cardiac genes as well as connective tissue growth factor (*Ctgf*), a marker of fibrosis, were dramatically upregulated in transgenic lines 1 and 2 and were upregulated to a lesser extent in line 3, consistent with the lower level of MEF2D expression in this line (Figure 6C). MEF2D overexpression in transgenic lines 1 and 2 also caused a diminution in expression of myosin, heavy polypeptide 6, cardiac muscle, α (*Myh6*) and upregulation of *Myh7*, a hallmark of pathological cardiac remodeling and reduced cardiac contractility. We conclude that MEF2D is sufficient to induce cardiac remodeling and cardiomyopathy.

Discussion

The results of this study demonstrate that MEF2D plays a unique and important role as a mediator of stress-dependent pathological remodeling of the adult heart. In the absence of MEF2D hypertrophy, chamber dilatation, fibrosis, and fetal gene activation were profoundly blunted following pressure overload or chronic β -adrenergic stimulation. Conversely, elevating MEF2D activity by transgenic overexpression was sufficient to substitute for stress

Figure 3

Echocardiographic analysis of *Mef2d* mutant mice following TAC. *Mef2d* mutant mice display less LV dilation during diastole and a less pronounced decrease in FS in response to TAC than WT mice.



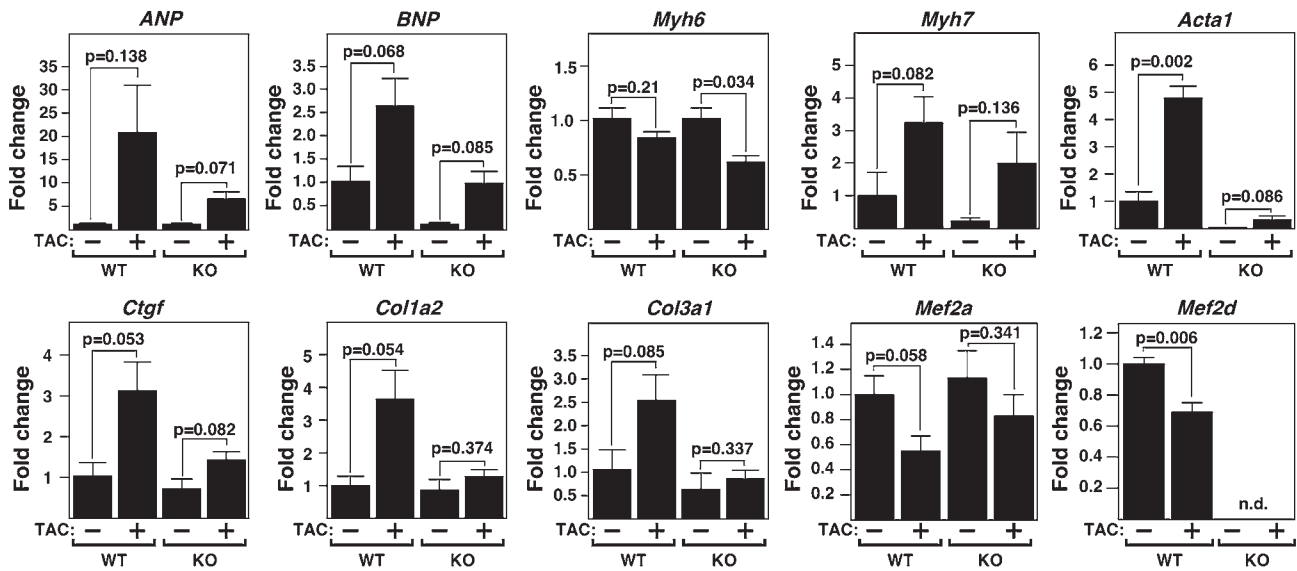


Figure 4

Resistance of *Mef2d* mutant mice to fetal gene activation following TAC. Expression of cardiac hypertrophic markers, fibrosis markers, *Mef2a*, and *Mef2d* in WT and *Mef2d* mutant hearts was detected by quantitative PCR 21 days after TAC ($n = 3-5$ per group). Values indicate expression level relative to a WT sham-operated group (\pm SEM). ANP, atrial natriuretic peptide; BNP, brain natriuretic peptide; *Myh6*, myosin, heavy polypeptide 6, cardiac muscle, alpha; *Myh7*, myosin, heavy polypeptide 7, cardiac muscle, beta; *Acta1*, actin, alpha 1, skeletal muscle; *Ctgf*, connective tissue growth factor; *Col1a2*, procollagen, type I, alpha 2; *Col3a1*, procollagen, type III, alpha 1; n.d., non-determined.

signals and drive pathological cardiac remodeling. The viability of *Mef2d*-null mice contrasts with the early embryonic lethality of *Mef2c*-null mice (11) and perinatal lethality of *Mef2a*-null mice (10), pointing to distinct functions for each MEF2 isoform at different stages of heart development.

Signaling pathways leading to cardiac remodeling. Our results indicate that MEF2D is a critical component of the signaling pathways through which pressure overload and adrenergic signaling drive cardiac remodeling and fetal gene activation. In this regard, prior studies showed that MEF2 regulates an array of genes involved in ion handling, extracellular matrix remodeling, and metabolism during pathological cardiac remodeling (21, 22). Some of these genes are direct targets of MEF2, whereas others are regulated indirectly by MEF2 through protein-protein interactions with other transcription factors and through subordinate MEF2-regulated transcription factors. Recent studies pointing to the importance of stress-inducible microRNAs in the cardiac remodeling response reveal an additional layer of potential regulation by MEF2 (23, 24).

Forced expression of MEF2A or -C in transgenic mice has been shown previously to cause dilated cardiomyopathy and fetal gene activation with little or no hypertrophy (21, 22). Overexpression of MEF2 or expression of a superactive MEF2-VP16 chimeric protein in primary cardiomyocytes also evokes a phenotype of myocyte elongation and sarcomere disorganization (21, 22). Similarly, our results show that overexpression of MEF2D causes ventricular dilation and fibrosis in transgenic mice.

Expression of a dominant-negative MEF2 mutant protein in the adult heart also reduces chamber dilation and improves contractility in the presence of constitutive calcineurin activation (22). Notably, dominant-negative MEF2 results in a minimal block to hypertrophy, suggesting that MEF2 function is required primarily for late stages of pathological cardiac remodeling. Since MEF2

associates with NFAT and GATA factors, both of which have been shown to mediate pathological cardiac signaling (17, 25-27), the blockade of cardiac remodeling by dominant-negative MEF2 likely involves the inhibition of multiple downstream transcriptional effectors in addition to endogenous MEF2. The diminution of cardiac remodeling in *Mef2d* mutant mice therefore provides a more direct illustration of the specific contribution of this MEF2 isoform to the cardiac stress response. It will be of interest to examine mice with a cardiomyocyte-specific deletion of *Mef2d* to determine whether the reduction of cardiac remodeling in *Mef2d* mutant mice results exclusively from the lack of MEF2D in cardiomyocytes.

While hypertrophy, fetal gene activation, and fibrosis were substantially blunted in *Mef2d* mutant mice, we observed residual hypertrophy and fetal gene activation in these animals. This likely reflects redundant functions of MEF2A, the other major MEF2 factor expressed in the adult heart, as well as other stress-responsive transcription factors such as NFAT, GATA, and CAMTA (25-27).

Myriad functions of Mef2 genes in vivo. The phenotypes of *Mef2a*, -c, and -d mutant mice reveal unique roles for each of these MEF2 factors at different steps in cardiac development: MEF2C regulates early cardiac morphogenesis (11), MEF2A regulates genes involved in contractility and energy metabolism after birth (10), and MEF2D serves as a mediator of stress-dependent gene expression in the adult heart. These unique functions are likely to reflect the distinct expression patterns of the different *Mef2* genes and, possibly, intrinsic differences in the transcriptional activities and protein-protein interactions of the MEF2 factors due to their amino acid sequence differences.

In addition to their functions in the heart, MEF2 factors are expressed in most or all tissues, and are likely to play key roles in diverse gene programs for cell differentiation and stress responsiveness. Conditional gene deletion of *Mef2c*, for example, has revealed

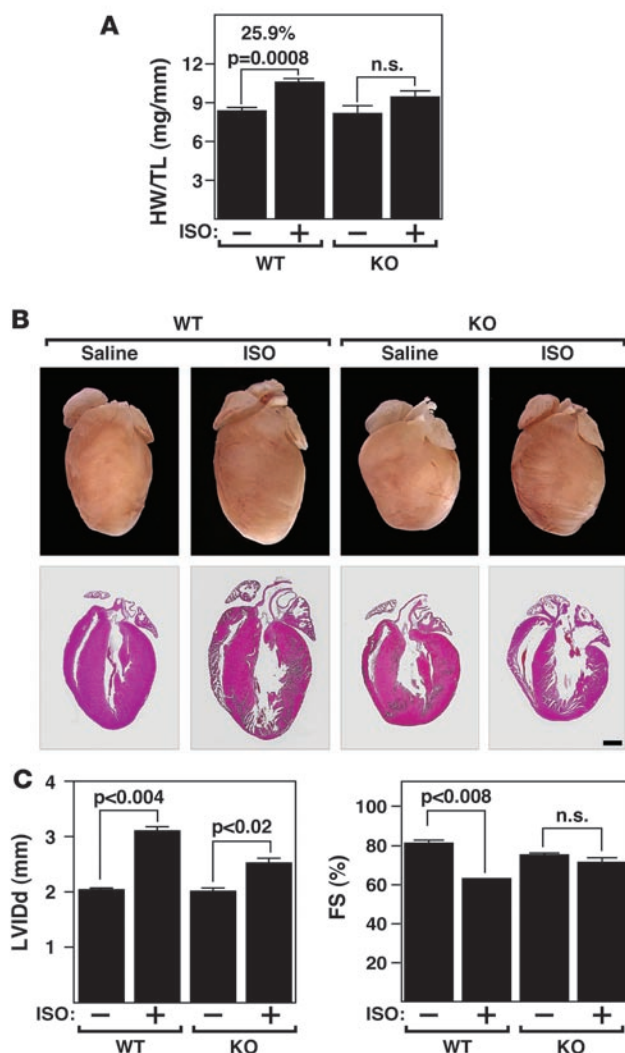


Figure 5

Decreased hypertrophic response of *Mef2d* mutant mice following β -adrenergic stimulation. **(A)** WT and *Mef2d* mutant mice were chronically infused with either saline vehicle or ISO (8.8 mg/kg/day) for 7 days and sacrificed for assessment of cardiac remodeling. HW/TL ratios (\pm SEM) are shown as bar graphs ($n = 7-9$ per group). n.s., non-significant. **(B)** Whole hearts of WT and *Mef2d* mutant mice treated with either saline vehicle or ISO (8.8 mg/kg/day) are shown at the top. Hearts were sectioned and stained with H&E as seen on bottom. Scale bar: 1 mm. **(C)** Representative bar graphs indicating less LV dilation and a decrease in FS in *Mef2d* mutant mice compared with WT mice in response to ISO treatment.

region) that encoded exon 3 was digested by *NcoI* restriction enzyme, blunted and inserted in between loxP sites of the targeting vector via *ClaI* and *XhoI* restriction sites. A 3.3-kb genomic fragment (long arm) that encodes exons 4-6 was digested and blunt-end cloned into the *EcoRI* restriction site downstream of the loxP sites in the targeting vector. This *EcoRI* site was not destroyed by the blunt cloning and was used for screening for positive recombination by Southern blot analysis (Figure 1A).

The *Mef2d* targeting vector was linearized by digestion with *NotI* and electroporated into 129SvEv-derived embryonic stem cells. Following positive-negative selection with G418 and FIAU, resistant colonies were screened by Southern blot analysis of *EcoRI* digested DNA using 5' and 3' probes (Figure 1, A and B). Three correctly targeted clones were expanded and injected into C57BL/6 blastocysts, which were transferred to the uterus of pseudopregnant females. High-percentage chimeric male mice (*Mef2d*^{neo-loxP/+}) were bred into a C57BL/6 background to obtain germline transmission. Heterozygous *Mef2d*^{neo-loxP} mice were intercrossed with hACTB:FLPe transgenic mice (37) to remove the neomycin resistance cassette. Southern blot analysis and PCR were performed to confirm the removal of the neomycin resistance cassette in the genome of *Mef2d*^{loxP/+} mice. To obtain mice with a *Mef2d*-null allele, the intercross of *Mef2d*^{loxP/+} mice to *Meox2-Cre* transgenic mice (19) was performed. Southern blot analysis and PCR were performed to confirm mice with *Mef2d* heterozygous- or homozygous-null (*Mef2d*^{KO}) alleles (Figure 1, B and C). All experiments on mice were conducted in 129SvEv/C57BL/6 mixed background. All animal studies were approved by the IACUC of the University of Texas Southwestern Medical Center.

Southern blot analysis and PCR genotyping. DNA sequences for PCR genotyping primers are as follows: 5'-TTCTGCAGACCATGAAGTACTAGT-3'; 5'-TAGAGCAAGTACCAGAATAGCCAGG-3'; and 5'-ACAGGAAAGGA-CAATACAAGAAAGCAC-3'. Southern blot probes were generated by PCR using the following primer sets: 5' probe forward, 5'-AAGCTTAGAATTTA-CAGCCTGCAGCGCTGA-3', reverse, 5'-GGATCCGAGCTAAGGTTTTGT-TAAGTTTCC-3'; and 3' probe forward, 5'-CTCGAGTCATGCATCCTGT-GTGAA-3'; reverse, 5'-GGATGACTTTGTTTAGGCTGTTGC-3'. In brief, tail genomic DNA was digested with *EcoRI* and analyzed using a standard Southern blot protocol. The 25 μ l PCR genotyping reaction contained 1 μ l

important roles for this gene in regulation of skeletal muscle fiber type and resistance to fatigue, as well as in endochondral bone development (28, 29). MEF2 factors also participate in neural and vascular development (30-32) and, recently, have been implicated in coronary artery disease, cancer and hepatic fibrosis (33-36). Thus, strategies to modulate the MEF2 activity in adult tissues may be therapeutically beneficial in the settings of a variety of diseases.

Methods

Generation of *Mef2d* mutant mice. The mouse *Mef2d* genomic locus was isolated from a 129SvEv mouse BAC library (BACPAC Resource Center at CHORI, Oakland, California, USA). A 1.3-kb genomic DNA fragment (short arm) that includes exon 2 and flanking region was digested with *XhoI* and *EcoRI* restriction enzymes. The *EcoRI* restriction site of this genomic DNA fragment was blunted and subcloned into the *XhoI* site located upstream of the FLP recombinase target-flanked (FRT-flanked) neomycin resistance cassette of the pDelboy targeting vector. A 1.2-kb genomic DNA fragment (the deleted

Table 2

Mef2d mutant animals show less LV remodeling compared with WT in response to ISO treatment

	LVIDd (mm)	LVIDs (mm)	FS (%)	HR (min ⁻¹)	AWd (mm)	PWd (mm)
WT saline	2.04 \pm 0.03	0.27 \pm 0.01	80.75 \pm 1.73	643.45 \pm 18.10	0.79 \pm 0.05	0.62 \pm 0.03
WT ISO	3.10 \pm 0.08	1.18 \pm 0.03	62.86 \pm 0.29	571.45 \pm 49.84	1.03 \pm 0.08	0.89 \pm 0.04
KO saline	2.01 \pm 0.06	0.40 \pm 0.02	75.03 \pm 0.53	679.27 \pm 12.63	0.78 \pm 0.04	0.70 \pm 0.01
KO ISO	2.52 \pm 0.09	0.81 \pm 0.06	71.12 \pm 3.15	684.10 \pm 8.50	0.86 \pm 0.08	0.77 \pm 0.02

$n = 3-4$ per group. All values are shown as mean \pm SEM.

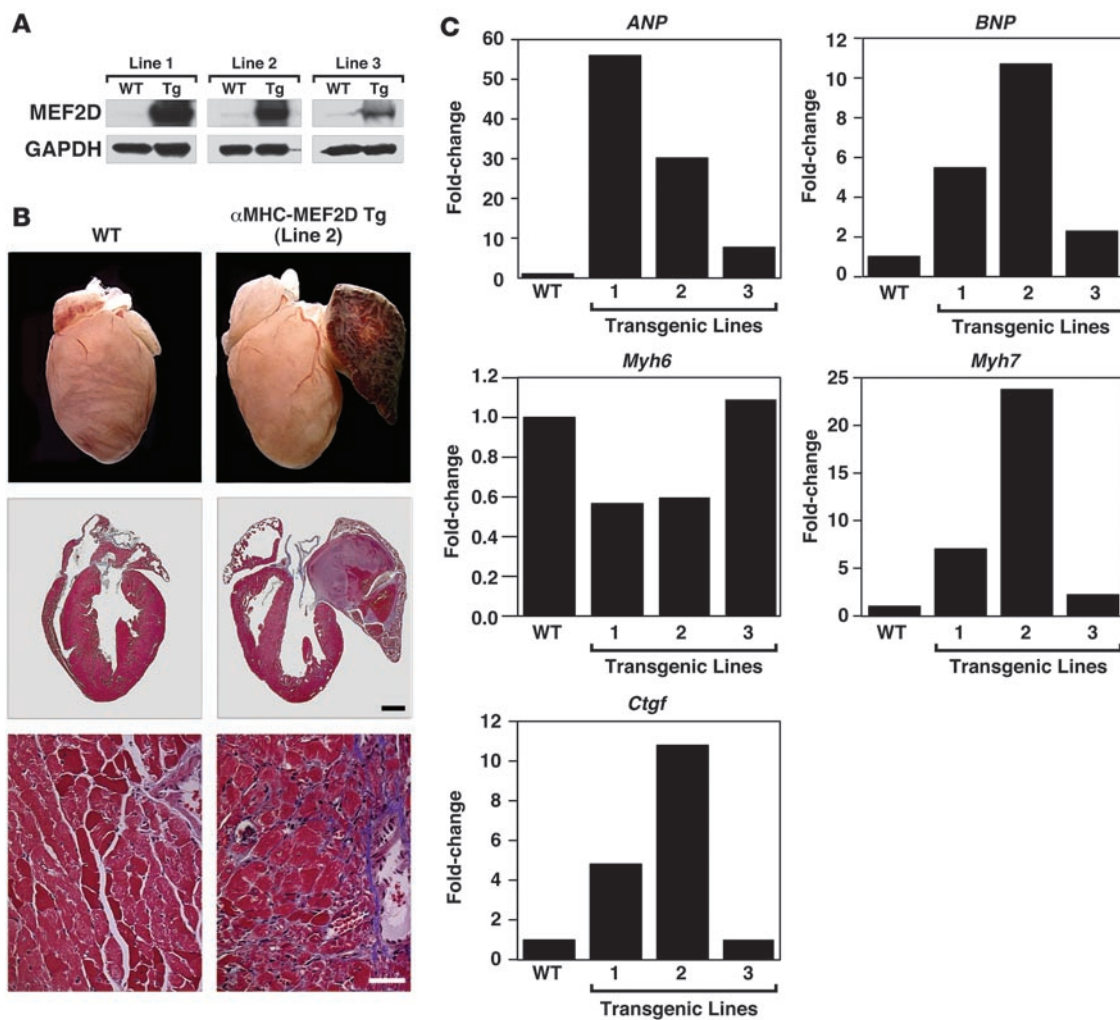


Figure 6 Pathological cardiac remodeling induced by overexpression of MEF2D. (A) Proteins were extracted from ventricles of α -MHC-MEF2D transgenic mouse lines at 6 weeks to 8 weeks of age for Western blot analysis using an antibody against MEF2D. GAPDH was used as a loading control. (B) Hearts were dissected from α -MHC-MEF2D male mice (line 2) and WT littermates at 8 weeks of age. Note the enlarged left atrium in the α -MHC-MEF2D heart. The lower panels are histological sections stained with Masson’s trichrome to detect fibrosis. Scale bar: 1 mm (middle panel); 40 μ m (bottom panel). (C) Expression level of various hypertrophy and fibrosis markers was evaluated by quantitative PCR in each transgenic line. Note the correlation between expression level of MEF2D (A) and that of hypertrophy and fibrosis markers.

of tail DNA as template, 2.5 U of Taq polymerase (Promega) in 0.5 μ l, and a final concentration of 2 mM MgCl₂. An annealing temperature of 70°C was used and the PCR products were electrophoresed on a 2% agarose gel.

RNA analysis. Total RNA was purified from isolated heart ventricles with TRIzol reagent (Invitrogen) according to the manufacturer’s instructions. Two micrograms of RNA was used as template to synthesize cDNA using random hexamers. Primer sequences used for RT-PCR are as follows: 1 forward, 5’-TTCGCTGTTTCCCGTCGGAG-3’; 2 reverse, 5’-TGGC-GGTTCCGTTTCATCAGTG-3’, forward, 5’-GTCCCCAGACATTCAT-GAGC-3’; 4 reverse, 5’-GCTCTGCTCCAGTGAGTCCT-3’, forward, 5’-CCCTGAGGAAGAAGGGTTTC-3’; 5 reverse, 5’-CAGGGAAGGAGT-GACCAGAG-3’; GAPDH forward, 5’-AGACAGCCGCATCTTCTTGT-3’, reverse, 5’-TGATGGCAACAATGTCCACT-3’. Quantitative PCR was performed using the following Taqman probes purchased from ABI: *Acta1*, Mm00808218_g1; *ANP*, Mm01255748_g1; *BNP*, Mm00435304_g1; *Col1a2*, Mm00483888_m1; *Col3a1*, Mm00802331_m1; *Ctgf*,

Mm00515790_g1; *Mef2a*, Mm01318991_m1; *Mef2d*, Mm00504931_m1; *Myh6*, Mm00440354_m1; *Myh7*, Mm01319006_g1.

Histology. Hearts were fixed in 4% paraformaldehyde in phosphate-buffered saline, embedded in paraffin, and sectioned at 5- μ m intervals. H&E staining, Masson’s trichrome staining, and picrosirius red staining were performed using standard procedures.

Measurements of mean cardiomyocyte area and myocardial fibrosis. Cardiomyocyte size was assessed on H&E-stained sections using ImageJ software (<http://rsb.info.nih.gov/ij/>) (NIH). About 100–150 randomly chosen cardiomyocytes from each group ($n = 2-3$) were analyzed to measure cross-sectional cardiomyocyte area. ImageJ software (NIH) was used to measure the amount of myocardial fibrosis on picrosirius red stained sections. About 30–50 randomly chosen frames from each group ($n = 2-3$) were analyzed.

Transthoracic echocardiography. Cardiac function and heart dimensions were evaluated by 2-dimensional echocardiography on conscious mice using a Vingmed System (GE Vingmed Ultrasound) and a 11.5-MHz linear



array transducer. M-mode tracings were used to measure anterior and posterior wall thicknesses at end diastole and end systole. LV internal diameter (LVID) was measured as the largest anteroposterior diameter in either diastole (LVIDd) or systole (LVIDs). The data were analyzed by a single observer blinded to mouse genotypes. LV FS was calculated according to the following formula: FS (%) = [(LVIDd - LVIDs)/LVIDd] × 100.

Statistics. Gene expression was normalized to GAPDH expression level and calculated as relative change. Differences in morphological, physiological, and biochemical parameters between groups were analyzed by 2-sided Student's *t* test using Microsoft Excel 2002. Non-significant (n.s.) difference indicates a *P* value greater than 0.05.

TAC and ISO administration. Male mice (8- to 12-week-old) either underwent a sham operation or were subjected to TAC as described (38). Perioperative mortalities (within 24 hours of surgery) were similar (about 10%) in wild-type and mutant mice subjected to TAC. There was no excess mortality during follow-up in any of the 4 groups (Sham or TAC, wild-type or mutant mice). Hearts were isolated 21 days following TAC for assessment of cardiac hypertrophy. ISO (Sigma-Aldrich) (8.8 mg/kg/day) or saline vehicle was administered using miniosmotic pumps (Model 2001; Alzet) subcutaneously implanted on the back of 8- to 10-week-old male mice. Cardiac hypertrophy was evaluated by measuring HW/TL of sacrificed mice 7 days after pump implantation.

Generation of transgenic mice. A cDNA encoding mouse FLAG-tagged MEF2D was cloned into a plasmid containing the α -myosin heavy chain transgene and human growth hormone (hGH) poly(A)⁺ signal (39), and transgenic mice were generated by standard techniques. Genotyping was performed by PCR analysis using genomic DNA, and transgenic expression of MEF2D in the heart was confirmed by Western blot analysis using mouse anti-MEF2D antibody (1:1,000; BD Transduction Laboratories) and mouse anti-GAPDH antibody (1:5,000; Chemicon).

Tissue culture and transfection. Standard procedures were used and described in detail in Supplemental Methods.

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