Unchecked thrombin is bad news for troubled arteries

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Thrombin is clearly a key trigger of thrombosis, the proximal cause of most morbidity and mortality in atherosclerotic cardiovascular disease. Might thrombin also contribute to longer-term, structural changes in the arterial wall that promote narrowing and cloting? A study in this issue of the JCI argues that it can. Aihara et al. report that haploinsufficiency of heparin cofactor II, a glycosaminoglycan-dependent thrombin inhibitor, exacerbates injury- or hyperlipidemia-induced arterial lesion formation in mice, possibly by excessive thrombin signaling through protease-activated receptors (see the related article beginning on page 1514).

Thrombin generation
As the main effector protease of the coagulation cascade (Figure 1), thrombin is required for hemostasis in response to injury. Thrombin is generated when tissue factor (TF), a transmembrane protein, binds plasma coagulation factors (1). Normally, TF is found mainly on cells in the extravascular compartment, and disruption of vascular integrity upon injury allows plasma coagulation factors to interact with TF. Factor VIIa complexes with TF to activate factor X, either directly or via factor IX activation, and the protease Xa converts prothrombin to thrombin. Active thrombin then cleaves fibrinogen to fibrin and triggers platelet activation and a variety of other cellular responses via G protein–coupled protease-activated receptors (PARs). Four main inhibitors keep thrombin generation and activity in check: TF pathway inhibitor (TFPI), antithrombin (AT, also known as ATIII), heparin cofactor II (HCII), and thrombomodulin (TM) through activated protease C (APC). Imbalances in this system can have important consequences, as evidenced by the multiple human genetic diseases associated with loss of coagulation factor or inhibitor function.

Pathological thrombin generation may have both acute and cumulative effects on arterial patency
In atherosclerotic plaques and injured arteries, TF expression is induced in smooth muscle cells and macrophages and is thus enriched and more proximal to the vessel lumen than in normal arteries (2). Thus even superficial injury, whether spontaneous or induced by percutaneous coronary intervention (PCI), exposes TF, lipid surfaces, and matrix proteins to circulating coagulation zymogens and platelets, triggering thrombin generation and platelet activation (Figure 2). In the short term, this leads to formation of thrombi that may or may not grow to a size sufficient to cause symptoms. In the long term, repeated or continuous exposure of endothelial cells, smooth muscle cells, and macrophages to active coagulation factors, fibrin, and platelet–released products might contribute to expansion of arterial lesions. Similar mechanisms might promote restenosis and/or thrombosis after stent placement, where compromised reendothelialization may result in prolonged exposure of TF to plasma.

How might exposure of mural cells to coagulation proteases promote atherosclerosis and neointima formation after arterial injury? Coagulation is one of the first responses to tissue injury, and coagulation proteases can regulate cellular behaviors through PARs (3). Thrombin activates PAR1 and PAR4; upstream proteases (VIIa and Xa) can activate PAR1 and PAR2. Responses to PAR signaling in cell culture hint that coagulation proteases and PARs together may help orchestrate not only hemostasis but also inflammation and repair after injury (4). Analogous to TF, both PAR1 and PAR2 are highly expressed in human atheroma relative to normal arteries and are induced in the arterial wall in response to injury in animal models (5). It is thus conceivable that hyperactivation of and hyperresponsiveness to coagulation proteases in the setting of arterial injury leads to excessive and therefore pathogenic repair responses (Figure 2).

A growing body of evidence from animal models supports a causal role for coagulation proteases and PAR signaling in arterial disease (Figure 1). Recombinant inhibitors (6–8) or PAR1 antagonists (9) ameliorate whereas anticoagulant deficiencies exacerbate (10–12) pathologic remodeling. Lack of effects of the anticoagulant heparin (13) and variable responses to another anticoagulant — hirudin (6) — in these models may point to roles for upstream proteases (6) or subtleties in anticoagulant function (discussed below). The study by Aihara et al. (14) in this issue of the JCI adds another piece to this puzzle with possible promise for a novel treatment strategy.

Anticoagulant activity of HCII
Until recently, HCII has been in the shadow of AT. Like AT, HCII is a serpin whose activity increases several orders of magnitude in the presence of heparin or heparan sulfate (HS) glycosaminoglycans (GAGs) (15). Yet HCII is limited to thrombin in specificity and is not only less abundant than AT but also both inferior in basal antithrombin activity and a secondary player in heparin–mediated anticoagulation. In mice, absence of AT, TFPI, TM, or APC results in coagulopathy and pre- or perinatal lethality (16), whereas HCII-knockout animals reach old age without signs of spontaneous thrombosis (17). Consistent with these findings, HCII deficiency does not appear to predict thrombophilia in humans (16).

What, then, is HCII’s raison d’être? Photochemically-induced arterial injury did uncover a hyperthrombotic phenotype in...
HCII-deficiency in adult mice (17), so HCII function does complement that of other anticoagulants. The key appears to be location; notable recent advances in HCII biology have come from studies of its tissue-specific activation (15). A unique feature of HCII is that, unlike AT, it can be activated by dermazon sulfate (DS) as well as HS GAGs. DS is enriched in areas low in anticoagulant HS such as the placenta and in the arterial wall; it may thus allow HCII to function where AT comes up short. HCII levels increase to 150% of normal during pregnancy but are 50% of normal in severe preeclampsia, suggesting a role for HCII in regulating placental hemostasis. DS appears to be the major anticoagulant GAG in the vessel wall (15), and unlike heparin, DS has been shown to inhibit smooth muscle hyperplasia in animal models (13, 15). Thus, a picture emerges where DS distribution may afford HCII a unique role in regulating extravascular thrombin activity after arterial injury (Figure 2). DS is modified and less able to support PAR1 signaling (Figure 2). DS is modified and less able to support PAR1 signaling (Figure 2).

**Perspectives**

A number of caveats regarding interpretation and extrapolation of the current study (14) should be mentioned. While decreased HCII levels may indeed yield increased thrombin activity, the report does not address the possibility that the effect of decreased HCII levels is due to dysinhibition of other proteases. While mutations that cause gain of function for thrombin might increase neointima formation, it does not necessarily follow that loss of function for thrombin would decrease formation. In this regard it is interesting to note that large doses of exogenous HCII did not reduce neointima formation in HCII-deficient mice beyond a mixed-strain background, so it is conceivable that they lost a vital modifier retained in the original knockout. They confirmed S' and 3' recombination, single integration, and unaltered expression of neighboring genes. The authors suggest that the original allele may have been hypomorphic, but evidence is inconclusive. Whether embryonic lethality supports a role for HCII in regulating placental hemostasis or suggests that HCII has other targets than thrombin was not assessed in this study.

Addressing the role of HCII in arterial homeostasis in adult mice, Aihara et al. (14) showed increased hyperplasia after cuff- or wire-induced arterial injury in their heterozygous survivors. Effects were striking as HCII activity was reduced by the mere 40%. Neointima formation increased with or without increased adventitial thickening, depending on the model; evidence of enhanced cell proliferation in these regions supported the outcome. Pheno type correction with purified human HCII provided elegant proof of specificity. Underscoring a critical role for HCII in regulating arterial homeostasis, the authors went on to show increased susceptibility to diet-induced atherosclerosis in an apoE-null background. HCII haploinsufficiency was associated with increased expression of PAR1 and induction of inflammatory and mitogenic gene products, some known to be regulated by PAR1, in remodeled lesions. Notably, the substantive findings in this study have been corroborated by Tollefsen and colleagues (15). Collectively, these data support a model where uninterrupted thrombin activity may promote inflammatory cell recruitment and smooth muscle expansion, possibly through PAR1 signaling (Figure 2).

**HCII deficiency**

In their study in this issue of the JCI, Aihara et al. report (14) on an independently generated HCII-knockout mouse. Contrasting the original report (17), homozygous deficiency was not compatible with life. The reason for the discrepancy is elusive; the original study established absence of circulating HCII antigen and activity, and both knockout alleles were inbred to the same strain background. Aihara et al. did not assess survival in a mixed-strain background, so it is conceivable that they lost a vital modifier retained in the original knockout. They confirmed S' and 3' recombination, single integration, and unaltered expression of neighboring genes. The authors suggest that the original allele may have been hypomorphic, but evidence is inconclusive. Whether embryonic lethality supports a role for HCII in regulating placental hemostasis or suggests that HCII has other targets than thrombin was not assessed in this study.

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Figure 2
How might HCII deficiency exacerbate responses to arterial injury? Arterial injury may trigger thrombin formation by exposing extravascular TF to circulating coagulation zymogens (see Figure 1). Thrombin activates platelets and converts fibrinogen to fibrin, thus triggering thrombosis and thus, potentially, acute vascular occlusion. Both thrombin and upstream proteases VIIa and Xa may also contribute long-term to arterial remodeling and narrowing (stenosis) by signaling to circulating and mural cells through PARs. At least in vitro, PAR activation triggers leukocyte chemotaxis, endothelial contraction/secretion/expression of adhesion molecules, and smooth muscle cell proliferation/migration. The thrombin inhibitor HCII may be uniquely suited to inhibiting thrombin in the vessel wall, since it is activated by DS, which is synthesized by smooth muscle cells and fibroblasts. AT, by contrast, is only activated by HS enriched in the subendothelial matrix. Tissue-specific thrombin inhibition may thus explain why even partial loss of HCII activity exacerbates arterial remodeling in response to injury or hyperlipidemia in mice, as shown by Aihara et al. in this issue of the JCI (14). m, mice; h, humans.
specific levels of the coagulation cascade on restenosis in a rabbit atherosclerotic femoral artery injury model. *Circulation* 92:3041–3050.