hypertension rat model, elastase activity precedes MMP activity and subsequently induces MMP activity. The matrix degradation by MMPs and elastases further interacts with growth factors such as EGF, resulting in further induction of VSMC proliferation. These data raise the question of whether targeting this pathway using elastase inhibitors will be another effective therapeutic approach. Additional investigation is needed to determine which, if any, of these abnormalities initiate PAH and which ones are the best to target for therapeutic gain. It is clear that PAH has a multifactorial pathobiology. As we increase our understanding of the pathways involved and their interactions, we will hopefully increase the efficacy of novel therapeutic strategies for this severe disease and further improve the palliative therapy for PAH and ultimately cure PAH.

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A new cause of hypertriglyceridermia

The function of lipoproteins in vascular wall biology continues to unfold in an increasingly complex story. It was more than 20 years ago that the majority of apolipoproteins known to play a major role in lipid metabolism were identified, and 9 of them (APOA1, APOA2, APOA4, APOB48, APOB100, APOC1, APOC2, APOC3, and APOE) have long been known to be most relevant to the regulation of lipoproteins. In contrast, additional apolipoproteins have only recently been identified, and their roles in lipoprotein metabolism are still being uncovered. For instance, APOM and its important function in HDL formation were just described this year (1). An important discovery in triglyceride biology was made in 2001 when a new member of the apolipoprotein family, human apolipoprotein A-V (APOA5), was identified by postgenomic techniques. It was described independently by 2 groups; the first group used comparative sequencing (2), and the second group identified rat Apoa5 after observing the marked upregulation of the Apoa5 gene during liver regeneration (3). Since its plasma concentration is only about one-thousandth that of some other apolipoproteins (e.g., APOA1 and APOA2) (4), the relevance and function of APOA5 was not immediately fully recognized.

In this issue of the JCI, Marçais et al. describe a truncation mutation of APOA5 associated with severe familial hypertriglyc-

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**Give me A5 for lipoprotein hydrolysis!**

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Two theories of APOA5 function

Since APOA5 was discovered, 2 mechanisms of APOA5 action have been proposed. One is that APOA5 may inhibit hepatic VLDL production (15–17), the other that APOA5 is an activator of intravascular triglyceride hydrolysis by LPL (18–20). Arguments can be made to support both theories; based on its structural properties (it is strongly hydrophobic), some homology with microsomal triglyceride transfer protein, and its abundant subcellular accumulation in the endoplasmic reticulum as well as the results of secretion studies performed in transfected cells, it was proposed that APOA5 may interfere with VLDL assembly (15–17). In mice, adenoviral overexpression of murine apolipoprotein A-V (Apoa5) resulted in a decreased production rate of VLDL triglycerides (20). However, these data were not confirmed by studies in Apoa5 transgenic mice (19). Most importantly, as determined by APOB kinetic studies using stable isotopes and reported in this issue of the JCI, Marçais et al. show that the VLDL production rate was normal but the fractional catabolic rate of VLDL-APOB was decreased more than 20-fold in patients lacking normal APOA5 (5). This finding is in accordance with mouse Apoa5 overexpression data; both transgenic and adenoviral overexpression of Apoa5 resulted in a faster turnover of apoB and triglycerides from VLDL and chylomicrons (18–20).

APOA5 augments plasma triglyceride hydrolysis . . .

Taking all the data together, there seems to be a strong argument for APOA5 being an activator of plasma triglyceride hydrolysis. Here is where it gets difficult; LPL is the key enzyme in intravascular triglyceride hydrolysis. Bound to endothelial proteoglycans, it acts predominantly in skeletal muscle and adipose tissue, supplying fatty acids as an energy source for these tissues. How can APOA5 at such low concentrations influence plasma hydrolysis? Usually, LPL is measured by its hydrolytic activity in the plasma after heparin injection (postheparin plasma). In the current study, Marçais et al. found lower LPL activities and mass in their APOA5-deficient patients (5). However, careful examination of mouse lines with Apoa5 deficiency or overexpression did not reveal any differences in hepatic lipase and LPL activities (19). Frucht-Najib et al. have reported an increased hydrolysis of VLDL in vitro by free LPL in the presence of very high amounts of recombinant APOA5 (600 µg/ml; about 1,000-fold of plasma concentration) (18). Similarly, Schaap et al. showed an influence of APOA5 on LPL-mediated lipolysis of triglyceride-rich, apolipoprotein-free emulsions (20). On the other hand, using 3 different substrates, Lookene and coworkers did not detect any APOA5-mediated changes in triglyceride hydrolysis by non-proteoglycan-bound LPL (21).

. . . only if LPL is bound to proteoglycans

Several investigators (22–24) have postulated that plasma lipolysis is limited by factors other than the amount of postheparin plasma LPL activity. Instead, binding of triglyceride-rich lipoproteins to endothelial proteoglycans could be rate limiting. In consideration of the physiological condition in which LPL is only bound to endothelial proteoglycans, the influence of APOA5 on LPL hydrolysis was measured in the presence and absence of proteoglycans. Without proteoglycans, APOA5 derived either from triglyceride-rich lipoproteins,

Figure 1

Triglyceride-lowering effect of APOA5. (A) Triglyceride-rich lipoproteins such as VLDL are hydrolyzed by the lipolytic action of dimeric LPL, which is bound to heparan sulfate proteoglycans on the vascular endothelium. We propose that APOA5 targets VLDL to proteoglycans, placing VLDL in close proximity to LPL. At the same time, APOA5 may activate proteoglycan-bound LPL by stabilizing the dimerized conformation or by binding to an LPL allosteric site. After hydrolysis, remnant particles (Rem) are released into the circulation, and APOA5 can be transferred and reused by other VLDL particles. In the postprandial situation, HDL can act as an APOA5 donor for intestinally derived chylomicrons. Thus, APOA5 is a potent activator of the natural lipolytic system. (B) Proposed situation in patients with the Q139X mutation in APOA5 (Q139X-APOA5). Truncated APOA5 is not associated with lipoproteins and cannot sufficiently target triglyceride-rich lipoproteins to proteoglycans. Instead, binding of the Q139X mutation in APOA5 to proteoglycan-bound LPL results in detachment and degradation of LPL.
from APOA5 transgenic mouse HDL, or from a recombinant source did not alter the LPL hydrolysis rate (19, 21). In the presence of proteoglycans, however, we have shown that APOA5 led to a significant and dose-dependent increase of LPL-mediated hydrolysis of VLDL and chylomicron triglycerides (19). Lookene et al. identified a proteoglycan-binding site on APOA5 (residues 186–227). Using surface plasmon resonance binding studies, they found that APOA5 could specifically increase the binding of chylomicrons and VLDL to proteoglycans. This effect was even increased by the presence of LPL and was abolished by heparin (21).

How APOA5 lowers plasma triglyceride levels

Based on these data, we propose the following model for the function of APOA5 (Figure 1). By binding to chylomicrons or VLDL at 1 site and to endothelial proteoglycans and LPL at the other, APOA5 stabilizes the endothelial lipolytic system and thereby augments LPL-mediated plasma triglyceride hydrolysis. In addition, APOA5 can stabilize dimeric LPL or, less likely, modulate the shape of triglyceride-rich lipoproteins, making them a better substrate for LPL. Direct interaction of APOA5 with LPL is with the presence of a previously described a regulator of plasma triglycerides. Based on these data, we propose the following model for the function of APOA5 (Figure 1). By binding to chylomicrons or VLDL at 1 site and to endothelial proteoglycans and LPL at the other, APOA5 stabilizes the endothelial lipolytic system and thereby augments LPL-mediated plasma triglyceride hydrolysis. In addition, APOA5 can stabilize dimeric LPL or, less likely, modulate the shape of triglyceride-rich lipoproteins, making them a better substrate for LPL. Direct interaction of APOA5 with LPL is

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