Deletion of vanilloid receptor 1–expressing primary afferent neurons for pain control

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Control of cancer, neuropathic, and postoperative pain is frequently inadequate or compromised by debilitating side effects. Inhibition or removal of certain nociceptive neurons, while retaining all other sensory modalities and motor function, would represent a new therapeutic approach to control severe pain. The enriched expression of transient receptor potential cation channel, subfamily V, member 1 (TRPV1; also known as the vanilloid receptor, VR1) in nociceptive neurons of the dorsal root and trigeminal ganglia allowed us to test this concept. Administration of the potent TRPV1 agonist resiniferatoxin (RTX) to neuronal perikarya induces calcium cytotoxicity by opening the TRPV1 ion channel and selectively ablates nociceptive neurons. This treatment blocks experimental inflammatory hyperalgesia and neurogenic inflammation in rats and naturally occurring cancer and debilitating arthritic pain in dogs. Sensations of touch, proprioception, and high-threshold mechanosensitive nociception, as well as locomotor function, remained intact in both species. In separate experiments directed at postoperative pain control, subcutaneous administration of RTX transiently disrupted nociceptive nerve endings, yielding reversible analgesia. In human dorsal root ganglion cultures, RTX induced a prolonged increase in intracellular calcium in vanilloid-sensitive neurons, while leaving other, adjacent neurons unaffected. The results suggest that nociceptive neuronal or nerve terminal deletion will be effective and broadly applicable as strategies for pain management.

Introduction

In patient populations ranging from pediatric to the elderly, control of severe pain with currently available analgesics is frequently less than optimal (1–3). Opioids remain the mainstay of treatment for moderate to severe pain, but they cannot universally provide relief. Some patients with cancer have pain that is refractory to conventional medical therapies including opioid analgesia, and many of them experience aversive, dose-limiting side effects. Despite more than 100 years of searching for new pharmacological modalities to treat severe pain (4–6), only very few have emerged, and the current alternatives to opioids are nonselective chemical or surgical neuroablative interventions and palliative sedation (7–9). Both options are unsatisfactory, however. With neuroablative interventions, neurons responsible for motor, bladder, and bowel function may be indiscriminately destroyed, while with palliative sedation there is loss of consciousness; in both cases, quality of life is potentially compromised (10). Implantable pumps delivering continuous intrathecal administration of drugs may offer relief, but pump placement is expensive, requires a surgical procedure, has its own set of complications, and may not be appropriate or completely effective for advanced stages of cancer (11).

In the present article, we explore selective nociceptive neuronal deletion as an approach to control nociceptive processes in experimental models and naturally occurring clinical conditions. This approach takes advantage of specific gene expression patterns in nociceptive, mainly small unmyelinated C-fiber neurons localized in sensory ganglia. These neurons transduce sensations generated by noxious heat and endogenous inflammatory algic substrates and express high levels of the transient receptor potential cation channel, subfamily V, member 1 (TRPV1; also known as the vanilloid receptor, VR1), a sodium/calcium ion channel, as compared with other sensory neurons in the dorsal root (DRG) or trigeminal ganglia (TG) (12–14). Direct live-cell imaging of primary DRG cultures and ectopically TRPV1-expressing cells has demonstrated that resiniferatoxin (RTX), a potent TRPV1 agonist (15), causes a large, prolonged increase in the free intracellular calcium concentration ([Ca2+]i). The resulting calcium cytotoxicity rapidly compromises and then deletes only the TRPV1-expressing neurons or cells (16). These data provided a mechanistic rationale for therapeutic intervention in which application of RTX in the vicinity of DRG or TG neuronal cell bodies in vivo would delete TRPV1+ neurons, thereby eliminating sensitivity to nociceptive input associated mainly with inflammation, hyperalgesia, and cancer-related pain. Other sensory modalities mediated by neurons not expressing TRPV1 would remain unaffected (e.g., proprioception and high-threshold mechanosensation). We reasoned that the same mechanism applied to nociceptive nerve terminals in the periphery would reversibly inhibit pain transduction but spare neuronal perikarya in the DRG, which are remote from the site of RTX application. Thus, three strategies for pain modulation based on differential application of RTX were investigated: single intranganglionic, multiganglionic (intrathecal), and peripheral nerve terminal. We employed robust, well-characterized nociceptive experimental models in rats, as well as a canine veterinary model presenting with naturally occurring conditions (cancer, osteoarthritis) that have direct relevance.
to humans. To predict efficacy and establish whether the same selectivity would occur in humans, we exposed primary cultures of human DRG neurons to RTX. Live-cell imaging disclosed that only a subpopulation of DRG neurons exhibited activation with RTX; this is consistent with previous results in cultured rat DRG neurons. The procedures outlined should provide analgesia to a variety of disorders that are characterized by severe pain and that may include trigeminal neuralgia, arthritis, peripheral neuromas, and advanced metastatic disease.

Results

Single intraganglionic injection in rat experimental models. Unilateral intraganglionic microinjection of RTX into the TG (Figure 1, A and B) attenuated or eliminated both the afferent (nociceptive) and efferent (neurogenic inflammation) functions supported by C-fiber neurons (Figure 1, C–E). Intraganglionic RTX attenuated plasma extravasation; this is an indicator of neurogenic inflammation and efferent function mediated by C fibers (17, 18). Intravenous Evans Blue dye, which binds to plasma proteins, delineated the areas with intact innervation (Figure 1C), staining them deep blue. The skin remained largely white on the RTX-microinjected side, where trigeminal neurons expressing TRPV1 were deleted. Analgesic actions were observable as soon as 24 hours after injection using a sensitive test for C-fiber function, the eye-wipe response to corneal application of capsaicin (CAP). An intraganglionic dose of 20 ng of RTX nearly eliminated the wiping behavior, and complete suppression was obtained with 200 ng (Figure 1D). The antinociceptive effects from a single RTX injection were maintained for at least 1 year (Figure 1E). The long-duration behavioral effect and loss of TRPV1-immunoreactive (TRPV1-IR) neurons (Figure 1, F and G) suggest that RTX analgesia is the result of permanent cell removal and not the result of prolonged receptor desensitization; the latter phenomenon is observed when CAP is applied to neurons in acute in vitro electrophysiological experiments (19–21). The loss of CAP chemosensitivity did not affect the mechanosensitive aspects of the corneal reflex to the liquid droplet itself. Furthermore, there were no observable alterations in grooming that might indicate the presence of a sensory dysesthesia.

That intraganglionic RTX administration produced extensive deletion of TRPV1-expressing neurons could be seen by immunohistochemistry as early as 1–3 days after RTX treatment (Figure 2, A–D). To establish that another class of sensory ganglion neurons remains intact, we stained for a neurofilament protein (N52) expressed in neurons with large-diameter myelinated axons (22). Both single- and double-stained (N52 and TRPV1) sections were examined. Injected TGs contain many N52+ neurons (684 ± 32, brown in Figure 2, C and D) but very few TRPV1+ neurons (123 ± 36, purple), while contralateral TGs show both N52+ (709 ± 23) and TRPV1+ (604 ± 68) neurons (Figure 2, E and F). The loss of TRPV1+ neurons was significant (P < 0.01). The loss was also evident using RT-PCR; however, a small amount of RNA_{TRPV1} was detectable in the RTX-injected ganglia (Figure 2G), consistent with cell counts showing substantial loss of TRPV1-IR neurons and retention of the N52 neurons not expressing TRPV1 (Figure 2, C and D).

Multiganglionic administration of RTX in rats. Severe pain secondary to advanced metastatic disease is generally more diffuse and not localized to one or two dermatomes. In these cases, multiple ganglia can be treated bilaterally via the cerebrospinal fluid. To test this, RTX was administered by lumbar puncture to target ganglia that innervate the tail and lower limbs. Treatment effectiveness and specificity were assessed by measuring (a) selective attenuation of noxious thermal and inflammatory responses, (b) the degree of spread of the drug by behavioral comparison to the forelimbs, corresponding to the spatial extent of the cell deletion, and (c) retention of locomotor activity and mechanosensation. RTX administered intrathecally (10, 50 ng) produced minor increases in withdrawal latency to a radiant noxious thermal stimulus (23, 24), whereas at doses of 100 and 200 ng, many animals reached the 14-second cutoff for hindpaw and tail stimulation without affecting forepaw latencies (Figure 3A). Intrathecal RTX also blocked carrageenan-induced thermal hyperalgesia, consistent with a substantial role of TRPV1+ neurons in experimental inflammatory conditions (Figure 3B), as shown by targeted disruption of the TRPV1 gene (25, 26). Behavioral effects were reflected at the cellu-
N52 sensory neurons (brown) are retained on the RTX-injected side, whereas TRPV1 neurons (purple) are deleted (C). On the contralateral, noninjected side, both N52 and TRPV1 neurons are intact (D). Quantification shows no significant difference in the number of N52-IR perikarya after RTX, whereas an 80% reduction in TRPV1+ neurons occurs (E and F). Bars in graph represent the average neuron counts in three sections of TG from three to five different rats assessed between 1 and 3 days after injection (*P < 0.01). (G) RT-PCR shows reduction of mRNATRPV1 in two different rats. M, markers; NO, no primer; SC, spinal cord; T, RTX treated; C, contralateral TG. Bars: 0.5 mm (A and B) and 50 μm (C and D).

Figure 2
Selective loss of TRPV1-IR sensory ganglion neurons after RTX (200 ng) microinjection. (A) Immunostaining for TRPV1 shows extensive loss of IR neuronal perikarya after RTX injection compared with contralateral noninjected TG (B). (C–F) Double labeling shows that large myelinated N52+ sensory neurons (brown) are retained on the RTX-injected side, whereas TRPV1+ neurons (purple) are deleted (C). On the contralateral, noninjected side, both N52 and TRPV1 neurons are intact (D). Quantification shows no significant difference in the number of N52-IR perikarya after RTX, whereas an 80% reduction in TRPV1+ neurons occurs (E and F). Bars in graph represent the average neuron counts in three sections of TG from three to five different rats assessed between 1 and 3 days after injection (*P < 0.01). (G) RT-PCR shows reduction of mRNATRPV1 in two different rats. M, markers; NO, no primer; SC, spinal cord; T, RTX treated; C, contralateral TG. Bars: 0.5 mm (A and B) and 50 μm (C and D).

Lar level by decreases in TRPV1 and calcitonin gene-related peptide (CGRP) immunoreactivities in the lumbar ganglia and dorsal horn, respectively, consistent with deletion of TRPV1-expressing neurons (Figure 3, D–G). In contrast, TRPV1+ neurons in cervical ganglia, which are remote from the level of lumbar RTX application, were unaffected (Figure 3C), corresponding to retention of normal forepaw-withdrawal latencies (Figure 3A). Thus, by targeting one or many ganglia, using the intraganglionic or intrathecal routes, respectively, the spatial extent of the therapeutic action may be adjusted to match varied clinical presentations.

Retention of other somatosensory functions and locomotion. Importantl
ed that the effect would be transient, and that TRPV1+ cells would remain intact; both of these predicted results were observed (Figure 3, I–K). Counts of TRPV1-IR neurons from eight sections of L5 ganglia showed 119 ± 23 neurons on the injected side and 113 ± 10 on the contralateral side. Thus, the dose and anatomical site of RTX administration. The same concept of ligand-activated calcium cytotoxicity was applied to the peripheral nerve terminals to obtain a transient reduction of nociceptive transmission. Injection of 100 ng RTX subcutaneously into the hindpaw significantly attenuated thermal nociception for approximately 20 days. The analgesic activity was confined to the injected hindpaw, and no alteration in latency was detected for the contralateral paw or the forepaws (Figure 3I). Since the injection was remote from the cell body, it was predictably assessed by withdrawal from a pinprick (all animals had a withdrawal duration of 0.5 second or less) (27).

No alteration occurred in locomotor function as evaluated with an accelerating Rota-Rod, indicating the presence of intact motor axons and sensory proprioceptive neurons in ganglia exposed to RTX (Figure 3, D and H). Normal locomotion was also evident in the canine subjects (see later). As with the trigeminal microinjections, no indications of sensory neglect or denervation-induced hyperesthesia syndromes (e.g., autotomy behavior) were observed (28, 29). These features reinforce the clinical applicability of RTX, since the pathological, inflammation-associated aspects of pain are eliminated, while mechanonociception, motor function, and other sensory modalities remain intact.

Peripheral RTX administration. The same concept of ligand-activated calcium cytotoxicity was applied to the peripheral nerve terminals to obtain a transient reduction of nociceptive transmission. Injection of 100 ng RTX subcutaneously into the hindpaw significantly attenuated thermal nociception for approximately 20 days. The analgesic activity was confined to the injected hindpaw, and no alteration in latency was detected for the contralateral paw or the forepaws (Figure 3I). Since the injection was remote from the cell body, it was predictably assessed by withdrawal from a pinprick (all animals had a withdrawal duration of 0.5 second or less) (27).

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Veterinary clinical application. The canine model was established to assess the efficacy of primary afferent neuronal deletion in a higher order mammal with clinical problems more closely approximating the human situation. On the basis of attenuation of thermal response determined in an initial cohort, a single intrathecal dose of RTX (1 μg/kg) was administered under general anesthesia via cisternal puncture. Visual analog scale (VAS) ratings of nociceptive responses were maintained at 6 and 10 weeks (9.6 ± 5.3 and 7.5 ± 4.2, respectively) (Figure 4A). The animals initially presented with limb guarding as they walked, but this improved over time and daily activity increased, as was evident from video recordings. In fact, the entire demeanor of the dogs appeared improved following intrathecal RTX treatment. The efficacy of RTX action was further demonstrated by (a) the discontinuation or greatly reduced use of supplementary analgesics (opioids and NSAIDs in all eight dogs)
and (b) the fact that neoplastic advancement did not diminish the RTX-induced analgesia. Comparative histology of rat and canine sensory ganglia demonstrated that RTX eliminated many small-diameter neuronal cell bodies in both species, leaving no visible damage to surrounding neuropil (Figure 4, B–G). The early morphological signs of specific cell deletion were studied in rats sacrificed at 6, 12, and 24 hours. During this period, nuclear envelope ruffling and displacement of the nucleus in specific ganglion neurons were noted, indicating early stages of neuronal damage (Figure 4, B and C). In dogs, satellite cell activation and neuronophagia occurred around the dying neurons 3 weeks after intrathecal RTX treatment, progressing into characteristic nodules of Nageotte, which have been described in human diseases with pathological sensory aspects (Figure 4, D and E) (32). After 1 year, retention of TRPV1-IR neurons (arrows) in LS DRGs from RTX-injected (J) and noninjected (K) hindpaws. Cell counts of TRPV1-IR perikarya showed no significant alteration between left and right lumbar ganglia (see Results). Bars: 50 μm (C and D); 100 μm (J and K); 300 μm (E–G).
Discussion

These data demonstrate the crucial and necessary role that TRPV1+ neurons play in supporting nociception arising from inflammatory hyperalgesia, osteoarthritis, and cancer, and how this can be exploited to produce a new class of pain treatments. The RTX intervention removes the “C-fiber line of communication” but is not equivalent to drugs that block one of the multiple neuroactive substances these neurons release or respond to (e.g., substance P, CGRP, glutamate, bradykinin, prostaglandins). The diversity of transmitters and receptors involved in pain generation suggests that blocking only one element may not yield effective analgesia (6, 12, 35). In contrast, eliminating the participation of TRPV1-expressing neurons at either the nerve ending or the perikarya reveals the algesic profile of these neurons and their potential as pharmacological targets. Furthermore, the present chemical cellular knockout is not equivalent to removal of the TRPV1 molecule in TRPV1 KO mice (25, 26), although the behavioral repertoire seen with RTX-neuronal deletion is quite similar and reinforces the idea that TRPV1 is a key molecule in transduction of nociceptive stimuli.

RTX-induced Ca2+ cytotoxicity is the proposed mechanism for therapeutic cell deletion (16). Experiments in vivo demonstrate the behavioral effectiveness of RTX, and immunohistochemistry confirms that RTX deletes TRPV1+ neurons. Our results are consistent with earlier in vitro observations of CAP/RTX-mediated damage and elimination of cultured rat DRG neurons or cells ectopically expressing TRPV1 and do not appear to support a receptor desensitization mechanism for the analgesic actions (21, 36–39). The potency of RTX yields a duration and degree of intracellular calcium elevation that makes it fundamentally more efficacious for cell deletion in adult animals than CAP, which while highly algesic (40), is a readily reversible agonist of TRPV1 (16, 41, 42).

The data presented here suggest that cellular excision by “molecular neurosurgery” would confer several advantages over neurosurgical, chemical (phenol, ethanol), or radiofrequency neuroablative techniques in terms of selectivity, efficacy, and lack of side effects. RTX removes one class of neurons, whereas the other treatments lack sensory modality selectivity, and, if motor axons are compromised, bring about loss of muscular control. Conventional ablative approaches often yield incomplete relief or evolve into a deafferentation pain syndrome (43). Indiscriminate destruction of ganglionic neurons produces extensive proliferation of glia, inflammatory, and connective tissue cells, leading to scar formation and neuropathic sensory problems (43, 44). RTX excises the TRPV1+ neurons in such a way that formation of abnormal anatomical structures is circumvented. In RTX-treated animals, the lack of chronic behavioral maladaptations also may be due to retention of afferents not expressing TRPV1, incomplete removal of C-fiber neurons, and the fact that C-fiber primary afferents ascend and descend five or more dermatomes (45). All of these influences may prevent trophic disturbances and secondary neuronal alterations due to loss of synaptic contacts. Furthermore, TRPV1 cell deletion is not equivalent to peripheral nerve axotomy. Formerly, researchers had thought that rearrangement of A-beta fibers occurred upon damage to peripheral nerve, which stimulated collateral sprouting of A-beta fibers into the superficial dorsal horn and conferred mechanosensitive allodynia and abnormal behaviors in animals (autotomy) (28, 46).
Recent evidence suggests that sprouting is substantially less than originally reported (47, 48).

Deletion of nociceptors is especially effective in blocking inflammatory hyperalgesia in rats and clinical manifestations of pain in dogs. However, the loss of the TRPV1 nociceptors does not eliminate all types of pain sensation. Many animals remained responsive to thermal stimuli, and not all went to the behavioral cutoff. No significant alteration in response to noxious mechanical sensation was detected, as was also noted with intrathecal administration of CAP (37), indicating the presence of a population of high-threshold mechanoreceptive afferents that do not express TRPV1. No tendency to increased sensitivity to mechanical stimuli (von Frey hairs or withdrawal duration after pinprick) developed, suggesting that TRPV1-neuronal deletion, as performed here, does not induce a neuropathic pain state (49).

The anatomically directed nature of RTX administration allows the treatment to be tailored to the site (or sites) of nociceptive generation using interventional procedures with either a reversible or irreversible outcome as the therapeutic goal. Routes of administration can include single or multiple intraganglionic injections, intrathecal application for disseminated pain problems, or injections into peripheral sites. In all cases, the same principle of receptor-mediated Ca\(^{2+}\) cytotoxicity is operative (50), but the present data show that the distal injections do not eliminate the TRPV1-expressing DRG neurons, and therefore the analgesic effect is transient. The peripheral data support a spatially delimited Wallerian-like degeneration occurring locally and specifically in vanilloid-sensitive fibers (31, 51). Because the damaged ending remains connected to an intact cell body, the ending reinnervates the skin, a process that requires approximately 20 days in the present study. This reinnervation is consistent with studies of topically applied CAP (52) and recent data using the present study. This reinnervation is consistent with studies of topically applied CAP (52) and recent data using RTX (31).

Myelinated fibers are not affected as was reported earlier with CAP (53). The extent of the reversible degeneration correlates with the RTX application. While the intraganglionic and peripheral injections are well localized, the intrathecal and intracisternal routes have the potential to reach the brain via the cerebrospinal fluid. We observed no behavioral abnormalities in RTX-treated rats. In the dogs, even with intracisternal administration, there were no reports of abnormal personality changes. RNA blot (14) and our own immunohistochemical studies have not detected TRPV1 expression in brain regions or immunopositive neurons. The lack of behavioral alterations suggests that, despite reports of CAP actions (54, 55), RTX-binding sites (56), or TRPV1 expression (57), higher brain centers are not substantially affected upon vanilloid treatment. The RTX effect is clearly selective; nonresponding cells (traces 5–7) maintain normal calcium levels. Increase in normalized fluorescence intensity (numbered \(\Delta F/F_0\) traces refer to cells in E and F). Bars: 100 \(\mu m\).

Elements of the approach outlined here are scattered in the literature but have not been integrated into an explicit therapeutic intervention for several possible reasons: (a) early studies used CAP, which is less effective for cell deletion; (b) vanilloid agonists are generally thought of as algesic agents; and (c) the proposed mechanism(s) of action were either vaguely defined or inconsistent to form a basis for human clinical trials (21, 36–38, 40, 41, 52, 58). In contrast, the direct observation of RTX action, in real time, on in vitro DRG neurons or TRPV1-expressing cells provided a visually appreciable mechanism for therapeutic application (16).

The present experiments address an additional key issue in analgesia research, which is the predictive relationship between evaluation of new drugs in experimental models and their eventual performance in human clinical pain, where they are frequently ineffective or only minimally so. The canine model we introduce provides a transitional bridge between nonclinical rodent data and eventual human trials, and reinforces the idea that a treatment will be safe and effective. The canine study also highlights the need for better measurement tools for large-animal studies. The results with RTX were robust enough to be easily rated by the human observers; however, new methods to obtain objective, meaningful measurements should be developed. The selectivity of RTX in live-cell imaging of human DRG neurons provides a second type of translational observation between basic in vitro observations

### Figure 5

Human DRG neurons show selective sensitivity to RTX treatment. (A) H&E section, adult human DRG (*large-sized neurons). (B–D) Green immunofluorescence human TRPV1-IR neurons (arrows), contrasted with satellite cell nuclei (blue). (C) TRPV1-IR (green) is adsorbed by the peptide antigen (D). Selective response of human embryonic DRG neurons to RTX assessed by increases in [Ca\(^{2+}\)], using Fluo-4 AM imaging (E and F). Arrows indicate responding neurons, and arrowheads indicate nonresponding neurons. E shows the baseline fluorescence depicted as near 0 for the first 30 seconds of the normalized data in the graph (G). In F, the transmembrane calcium flux leads to increases in intracellular calcium in specific neurons, which become brightly fluorescent. Magnification ×200. (G) Traces of individual cells showing a substantial and abrupt response in vanilloid-sensitive sensory neurons (traces 1–4). Elevation of [Ca\(^{2+}\)] for a prolonged period of time is suggestive of imminent cell death in cells responding to vanilloid treatment. The RTX effect is clearly selective; nonresponding cells (traces 5–7) maintain normal calcium levels. Increase in normalized fluorescence intensity (numbered \(\Delta F/F_0\)) traces refer to cells in E and F). Bars: 100 \(\mu m\).
and human clinical application. These data represent a progressive series of steps from experimental rodent models to clinical veterinary models, to human cells that explore the clinical potential of selective nociceptive neuronal deletion for pain control. The cell-deletion approach may provide an important alternative to present treatments, and its eventual use will help to improve the quality of life in patients with unrelenting pain.

Methods

**Animals.** Procedures for rat studies followed the NIH Guidelines for the Care and Use of Laboratory Animals, and were approved by the National Institute of Dental and Craniofacial Research Animal Care and Use Committee. A protocol to treat dogs with RTX was approved by the University of Pennsylvania School of Veterinary Medicine Institutional Animal Care and Use Committee prior to the start of the study.

**Trigeminal microinjection.** Male Sprague-Dawley rats (300 g, \( n = 54 \)) were anesthetized with ketamine/xylazine and mounted in a stereotaxic frame. A 10-μl Hamilton syringe (Hamilton Company, Reno, Nevada, USA) was positioned at 2.5 mm posterior and 1.5 mm lateral to the bregma. The skull was drilled, and the needle was advanced until it gently touched the base of the skull; the needle was retracted 0.1 mm, and RTX (20 or 200 ng) was injected in a volume of 2 μl over 10 minutes. A pharmacological-grade preparation of RTX was formulated as follows: PBS, pH 7.2, containing 0.1 μg/μl RTX, 0.05% ascorbic acid, and 7% Tween 80. Vehicle contained no RTX. The Pharmaceutical Development Section, Clinical Center, NIH, performed characterization of this preparation for drug purity, stability, presence of endotoxin, and microbiological contamination.

**Immunohistochemistry.** Tissues were fixed in 10% buffered formalin for 24 hours and processed in paraffin. Five μm sections were stained with H&E or used for immunohistochemistry as described (31). Primary antibodies used were anti-rat TRPV1, 1:1,000 (Affinity Bioreagents Inc., Golden, Colorado, USA), CGRP, 1:2,500, and mouse monoclonal NS2 (1:2,000) (Chemicon International, Temecula, California, USA).

**Histology and immunohistochemistry of adult human DRG.** Tissue for neuroanatomical studies was obtained under a protocol with the Delaware Valley Kidney 1 Human Organ Donor Program. DRGs were removed within 1 hour of cessation of perfusion, and either specimens were paraffin-processed or frozen sections were cut for histochromy or immunohistochemistry, respectively. Human TRPV1-specific antibody (PA1-748; Affinity BioReagents Inc.) was used at a dilution of 1:500 and incubated overnight at 4°C visualized with FITC-conjugated AffiniPure Donkey Anti-Rabbit IgG (711-095-152; Jackson ImmunoResearch Laboratories Inc., West Grove, Pennsylvania, USA) and counterstained with Hoechst 33342 dye (H3570; Molecular Probes Inc., Eugene, Oregon, USA). Control sections for assessment of nonspecific binding were processed in an identical way, except that primary antibody steps contained 2.5 μg peptide antigen/ml.

**Human primary DRG culture.** Human DRG tissue was obtained from Advanced Bioscience Resources Inc. (Alameda, California, USA). Ganglia were collected in cold Lebowitz medium (Invitrogen Corp., Carlsbad, California, USA). After digestion with 0.125% trypsin at 37°C for 25 minutes, cells were seeded on coverslips coated with poly-D-lysine. Culture medium contained DMEM with 20 mM HEPES, 7.5% FBS, 7.5% horse serum, and 50 μg/ml nerve growth factor. After the first day, 5 mg/ml uridine, 2 mg/ml FUDR (Sigma-Aldrich, St. Louis, Missouri, USA) was added to inhibit nonneuronal cell division.

**Fluorescent microscopy and calcium imaging.** Fluorescent imaging was performed with an Olympus IX 70 microscope equipped with the CARV confocal imaging system (Atto Bioscience Inc., Rockville, Maryland, USA). Calcium imaging with Fluo-4 AM (F14201; Molecular Probes Inc.) was performed in an open imaging chamber. Calcium concentration was analyzed using the AttoVision (Atto Bioscience Inc.) and KaleidaGraph (Synergy Software, Reading, Pennsylvania, USA) software packages.

**RNA extraction and RT-PCR.** Dissected ganglia were immediately frozen at –80°C. Total RNA was extracted and treated with RNase-free DNase; 5 μg were then reverse transcribed and amplified over the linear range (26 cycles) using the Access RT-PCR System (Promega, Madison, Wisconsin, USA). The rat TRPV1-specific primers, TG-5’-AAGCCGACCTGAAGTTCTC and TG-3’-CTCCAGTGTCTGCATCCCTC yielded a 100-μb product; the GAPDH-specific primers, 5’-ACACAGTCCATGGCATAC and 3’-TCCAGACCCCTGTCTGTTGA, generated a 452-bp RT-PCR product. DNAs were separated by electrophoresis in 1% agarose gels, stained, and quantified by densitometry.

**Intrathecal injection of rats.** Animals (\( n = 58 \)) were anesthetized with isoflurane, and RTX (10–200 ng in 3 μl over 10 minutes) or vehicle was administered by inserting a 20-gauge guide catheter into the L4/L5 interspace. A PE-10 catheter was advanced 1 cm into the intrathecal space. The criterion for the intrathecal placement was withdrawal of clear cerebrospinal fluid into the catheter.

**Intraplantar RTX injection.** Rats (\( n = 20 \)) were injected with vehicle or RTX (100 μl) over a concentration range of 1, 10, and 100 μg/ml. A dose response was observed, and time course data for the 1-μg/ml dose are shown (Figure 3l). Cell counts were made on three sections ipsilateral and three sections contralateral from the L5 ganglia from three rats.

**Sensory and physiological tests.** The afferent functions of the trigeminal nerve were tested in 30 rats after TG microinjection of RTX using the CAP-induced eye-wipe response. A stock solution of 5% CAP was made in 75% ethanol, 0.05% ascorbic acid (Sigma-Aldrich). After dilution to 0.01%, CAP with saline (50 μl) was dropped into the cornea, and eye wipes were counted for 1 minute. A separate group of seven rats was tested over the course of a year.

**Efferent functions of the trigeminal nerve were tested in rats (\( n = 7 \)) at 7 days after injection. Rats were anesthetized with ketamine/xylazine and hair removed with Nair (Carter-Wallace Inc., New York, New York, USA). After washing thoroughly with water, 5% CAP cream, containing 0.05% ascorbic acid, was applied evenly to the entire head and shoulder region. After 15 minutes, the left femoral vein was catheterized and a 0.5-ml bolus of Evans Blue (Sigma-Aldrich) (20 mg/kg in saline) was injected.

Radiant-heat thermal testing was performed on unrestrained rats (\( n = 58 \)) according to previously published procedures (23, 24). The endpoint recorded was latency (seconds) to paw withdrawal. Thermal intensity was set to produce a withdrawal latency of 8–10 seconds. A 10-second withdrawal latency corresponds to a temperature at the surface of the paw of 45.2°C (23). Mechanical sensory stimulation included a pinch with toothed forceps, the application of graded von Frey filaments, and assessment of pinprick hyperalgesia as described previously (27). Rats treated 3 and 10 days and 1 year (\( n = 4 \) per group) prior to mechanosensory testing and a group of untreated controls (\( n = 4 \)) were examined.
Reduction in inflammatory hyperalgesia was tested by radiant thermal heat 5 days after intrathecal injection of either vehicle or RTX (200 ng; n = 8, four to a group). Rats were lightly anesthetized with isoflurane, and the plantar surface of the left hindpaw was injected with 100 μl 4% carrageenan (C-3889; Sigma-Aldrich), dissolved in saline. Assessment of locomotion and proprioception was performed in the intrathecally injected (vehicle and RTX) rats on an accelerating Rota-Rod (4–40 rpm in 5 min; model 7750; Ugo Basile Biological Research Apparatus, Comerio, Italy).

Intrathecal injection of dogs. We recruited dogs having advanced cancer or osteoarthritis (n = 8), whose owners brought them to the veterinary clinic for evaluation because of pain not well controlled by current medications. This was an open-design study, both for ethical reasons and because of the very prolonged duration of action of a single intrathecal dose of RTX, which we hypothesized would provide sustained improvement in activity and animal personality issues, thereby outlasting a ‘placebo effect’ in third-party observers.

Dogs were maintained under general anesthesia with isoflurane and oxygen. The cisterna magna was penetrated with a 20-gauge 4-cm spinal needle. When the needle is properly placed, the cerebrospinal fluid flows freely. A single dose of 1 μg/kg of RTX was injected over 10 minutes, followed by 0.2 ml of sterile saline. Any arousing effects were controlled with intravenous fentanyl.

VAS ratings. To assess the effect of a single dose of intrathecal RTX, owners were asked to rate their dogs’ pain before and after treatment, using a 100-mm VAS, with zero designated as no pain and 100 the worst pain imaginable. Ratings were obtained before injection, and at 2, 6, and 10 weeks after injection. (The lack of data in several cases results from euthanasia following progression or complications of the main pathology.)

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