Enkephalins Stimulate Leukemia Cell Migration and Surface Expression of CD9

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Abstract

Opioid peptides have been implicated in the regulation of tumor growth and biology; however, little attention has been given to the mechanisms that are involved. In this study we show that physiological concentrations of the endogenous opioid peptide methionine-enkephalin (MET-ENK) and the synthetic enkephalins D-Ala³,Me-Phe⁴,Gly(ol)⁵ and D-Ala²,D-Leu⁵ are stimulants for the in vitro migration of pre-B acute lymphoblastoid leukemia (ALL) cells. Activation of the human pre-B ALL cell lines NALM 6 and LAZ 221 with MET-ENK resulted in both an increase in their migration and an augmentation in the surface expression of the leukemia cell marker CD9. The opiate receptor antagonist naloxone reversed these enkephalin-induced effects on the leukemia cells. When the pre-B ALL cells were preincubated with an anti-CD9 mAb before challenge with MET-ENK their migration to the enkephalin was markedly reduced. These studies show that endogenous and synthetic opioid peptides are stimulants for pre-B ALL cell migration and suggest that CD9 is important in the regulation of leukemia cell motility. (J. Clin. Invest. 1995. 96:1366–1374.) Key words: enkephalins • opioid peptides • cell movement • pre-B cell • leukemia leukocyte-adhesion receptors

Introduction

First identified in brain tissues for their analgesic activity the opioid family of neuropeptides are now recognized as pleiotropic hormones that modulate vital processes in the periphery as well as the CNS. These neuropeptides are important in the regulation of moods, behaviors, higher cognitive processes, reproduction, pain control, inflammatory responses, and a variety of immune functions (1–5). In humans the intravenous use of opioid alkaloids has been associated with an increased risk for infectious and oncogenic diseases (6). Evidence obtained in animal models has documented that use of opioid peptides or alkaloids diminishes resistance to infection (6–8). Studies showing that stress stimulates the endogenous synthesis and release of opioid peptides has fueled speculation that the stress induced-release of these agents is important in disease processes (1, 3). Within the past decade many reports have detailed the in vivo effects of opioid peptides on human and rodent immunocytes; however, the precise mechanism(s) by which these neuropeptides influence health or disease have not been defined.

The best studied activity of opioid peptides is their characteristic effect on pain. This morphomimetic activity is dependent on the NH₂ terminal sequence (Tyr-Gly-Gly-Phe-Met or Leu) of the analgesic peptides and is blocked by classical opiate receptor antagonists such as naloxone (4). Opioid peptides interact with a number of distinct opiate/opioid receptor types (e.g., δ, μ, κ) and subtypes (e.g., μ1, μ2, δ1, δ2, κ1, κ2) that are expressed in the CNS and periphery (3, 4, 9, 10). Peripheral blood leukocytes have been reported to express both classical (naloxone-sensitive) and nonclassical (naloxone-insensitive) opiate/opioid recognition sites (3, 9).

Opioid peptides are potent signals that direct leukocyte motility and one of their key functions in the periphery may be in leukocyte recruitment and migration (11). Studies in rodent and large animal models have shown that the opioid pentapeptide methionine–enkephalin (MET-ENK) enhances lymphocyte trafficking (7, 11–13), stimulates transendothelial migrations, and causes the redistribution of circulating leukocytes (14, 15). In sheep the infusion of MET-ENK into the afferent lymphatics of popliteal nodes causes lymphocyte migrations from the node to the effenter lymph (16). When MET-ENK was injected into the cerebral ventricles of rodents both lymphocytes and macrophages were recruited to the injection site (14). Other studies carried out in vitro have confirmed that MET-ENK is chemotactic for isolated human peripheral blood T cells (11), monocytes (13, 14), and polymorphonuclear leukocytes (17). Opioid peptides, like other leukocyte chemoattractants, have been shown to trigger a number of biochemical and cellular events including transmembrane signaling (18), cell flattening, membrane spreading, and pseudopod extension (19). Despite this impressive number of studies documenting the effects of MET-ENK on leukocyte motility there is little known concerning the molecular processes that regulate the enkephalin-mediated effects.

MET-ENK and other opioid peptides have been reported to bind to certain kinds of tumor cells and to have antiproliferative activity (20). It is known that many growth-modulating factors also regulate cell movement (21–24); however, the effect of opioid peptides on the motility of leukemias, transformed leukocytes or other type tumors has not been addressed. To gain a better understanding of the actions of opioid peptides we have studied the effects of MET-ENK and other enkephalins on two human pre-B acute lymphoblastoid leukemia (ALL) cell lines, NALM 6 and LAZ 221, and the EBV-transformed mature B

1. Abbreviations used in this paper: ALL, acute lymphoblastoid leukemia; DADLE, [D-Ala, D-Leu]–enkephalin; DAMGO, [D-Ala, D-Me-Phe, D-Gly(ol)]–enkephalin; DesTyr-LEU-ENK, H-Gly-Gly-Phe-Leu-OH; MET-ENK, methionine–enkephalin.
cell line LAZ 388. We have used a migration assay and monitored cell morphology (i.e., cell flattening and membrane spreading) using flow cytometric analysis of forward and right angle light scatter for assessing the effects of MET-ENK and other opioid receptor ligands. Previously, other investigators have shown that chemotactic factors modulate the expression and function of cell surface adhesion molecules (e.g., the β2 and β1 integrins) (22-24). Since MET-ENK has been shown to be a stimulus for lymphocyte migration (11) we have investigated the possibility that this enkephalin also modulates the expression of adhesion receptors. For this purpose we used immunofluorescent staining and flow cytometry to measure the effects of MET-ENK on the surface expression of a panel of B cell determinants.

In this report we show that MET-ENK and synthetic enkephalin analogs serve as potent signals for the migration of NALM 6 and LAZ 221 pre-B ALL cells. Exposure of these leukemia cells to MET-ENK results in a discrete and transient increase in CD9 surface expression as well as an enhancement in migration. Preincubation of the leukemia cells with anti-CD9 mAb before challenge with MET-ENK blocks their migration to the enkephalin. These findings suggest that leukemia cell migration to MET-ENK is regulated, at least in part, via CD9 expression. In previous reports CD9 has been implicated in the formation of pre-B ALL cell homotypic aggregations (25, 26) and in the spontaneous locomotion of some kinds of tumor cells (27, 28). Our findings provide the first evidence that CD9 is involved in the migration of lymphoid cells. These studies provide new insights into the actions of opioid peptides and into mechanisms that regulate lymphoid cell motility.

Methods

Cells. The pre-B ALL cell lines NALM 6 and LAZ 221 (CD9, CD10, IgGl slg ) and the EBV-transformed B lymphoblastoid cell line LAZ 388 (CD19, CD20, CD9, CD10) have been described previously (29). The LAZ 221 and LAZ 388 lines were established from donor cells harvested from a patient at the Dana-Farber Cancer Institute, Boston, MA (29) and were obtained for these studies from Dr. Jerome Ritz.

Cells were passaged (1-2 × 10^6/ml) every 2-4 d in RPMI 1640 medium (BioWhittaker, Inc., Walkersville, MD) supplemented with 2 mM glutamine, a solution of 100 U/ml-100 μg/ml of penicillin-streptomycin (Gibco Laboratories, Grand Island, NY), and 10% bovine serum (HyClone Laboratories, Inc., Logan, UT). Cultures were maintained at 37°C in a 5% CO2 tissue culture incubator.

Reagents and antibodies. The naturally occurring pentapeptide MET-ENK (Tyr-Gly-Gly-Phe-Met), and the synthetic peptides [D-Ala2,D-Leu5]-enkephalin (DADLE), [D-Ala2,Me-Phe4,Gly(ol)1]-enkephalin (DAMGO), H-Gly-Gly-Phe-Leu-OH (Des-Tyr-Leu-ENK), and FMLP were purchased from Sigma Chemical Co. (St. Louis, MO). Naloxone (Norcan) was obtained from DuPont Pharmaceuticals (Manchester, PA, Puerto Rico).

Anti-CD9 and -CD10 mAb of the IgGl isotype were from Biodesign International (Kennebunkport, ME) and those of the IgGl isotype were obtained from Amac, Inc. (Westbrook, ME). Anti-CD20 (IgGl), -CD14 (IgG2a), -CD29 (IgGl) and -CD54 (IgGl) mAb were from Biodesign International. Anti-CD72 (IgG2a), and -CD23 (IgGl) mAb were from The Binding Site, Inc., (Birmingham, United Kingdom) and anti-CD22 (IgGl) was from Amac Inc., Anti-CD2 (IgGl), -CD3 (IgGl), -CD11a (IgGl), -CD11b (IgGl), -CD18 (IgGl), -CD45 (IgGl), -CD8 (IgGl), -CD58 (IgGl), -Class I (HLA-A,B,C, IgG2a), and -Class II (HLA-DR, IgG2b) were prepared as ascites developed in Balb/c mice. Hybridoma cells were obtained for the anti-Class II mAb (LB3.1) from Dr. Peter Knudsen and Dr. Jack Strominger, Dana-Farber Cancer Institute, Boston, MA and for anti-CD2 (HB195), -CD3 (CRL 8001), -CD11a (HB 202), -CD11b (HB204), -CD18 (HB203), -CD45 (HB196), -CD58 (HB205) and -Class I (HB 95) from the American Type Culture Collection (Rockville, MD). Anti-CD19 mAb (IgGl) was a gift from Dr. Ken Anderson, Dana-Farber Cancer Institute, Boston, MA. The mAb recognizing VLA’s 1-6 were from the V Human Leukocyte Typing Workshop (subpanel 6) and the DE9N anti-VLA β chain mAb (IgGl) was provided by Dr. Jeff Bergelson, Dana-Farber Cancer Institute, Boston, MA (30). The MOPC 21 myeloma protein (IgGl) that was used as a control was purchased from Organon Teknika Corp. (Durham, NC).

Migration assays. Cell migration was measured using microchemotaxis chambers (NeuroProbe, Inc., Cabin John, MD) as previously described (11). Cells (5 × 10^6/ml) were separated from peptides in the lower wells of the chamber by nitrocellulose filters (Sartorius, 5 μm; Neuproene, Inc.). In some studies the cells were preincubated with 10^-5 M naloxone for 15 h at 37°C or anti-CD9, -CD10, or -VLA 4 mAb for 30 min at 4°C and then incubated in the microchemotaxis chambers for 75-90 min at 37°C. The nitrocellulose filters were fixed and stained with Congo red dye (Sigma Chemical Co.). Cells within the filters were identified by fluorescence microscopy and then enumerated utilizing an optical image analyzer (Optomax V image analyzing system; Analytical Instruments, Hollis, NH). In some studies the cells were identified visually and photographed with the use of a fluorescence microscope.

Flow cytometric analysis. For this we used Flow Cytometry (Becton Dickinson and Co.) Fluorescence was measured as the arithmetic/linear mean relative fluorescence intensity; the range of values was between 1 and 10,000 (10^-4^). Indirect immunofluorescence staining and flow cytometric analysis of cell surface molecules. Cells were resuspended at 10^6/ml in PBS and then incubated for periods of 5-120 min at 37°C with MET-ENK (0-10^-5 M, final concentration). Incubations were terminated by transferring the samples to an ice bath and then pelleting the cells by centrifugation at 1,800 rpm for 5 min at 4°C.

Flow cytometric analysis was performed using a FACScan® Flow Cytometer (Becton Dickinson and Co.) equipped with a LP9000 Series computer. Data were collected as histograms depicting the fluorescence intensity for 5,000-10,000 cells per sample; arithmetic/linear mean relative fluorescence intensities were obtained using the Lyss System software analysis package. Arithmetic/linear values (range, 1-10,000) were used for all sample to sample comparisons. The nonbinding IgGl MOPC myeloma protein as well as isotype matched nonreactive mAb were used as controls for measuring the autofluorescence of B cells.

Results

Methionine–enkephalin stimulates a naloxone-sensitive alteration in the morphology of NALM 6 cells. An initial set of studies were carried out to determine whether cultured human
B cells responded to the naturally occurring opioid pentapeptide MET-ENK. Flow cytometric analysis of forward and right angle light scatter was used for monitoring the effects of MET-ENK on the morphology of NALM 6 cells. As indicated (Fig. 1), treatment of the NALM 6 cells with MET-ENK provoked a rapid change in cell morphology (i.e., an increase in right angle and forward light scatter). Physiological doses (i.e., \( \approx 0.1\) nM) of MET-ENK initiated this morphological response since enkephalin doses as low as 0.01 nM elicited a near maximum response (Fig. 1). These results provided evidence that the NALM 6 pre-B ALL cell line was responsive to MET-ENK. Consistent with the specificity expected for a classical opiate/opioid effect the morphological response to MET-ENK was inhibited when the NALM 6 cells were preincubated with the opiate receptor antagonist naloxone (Fig. 1). Whereas the application of MET-ENK to the NALM 6 cells enhanced both right angle and forward light scatter the preincubation in naloxone, alone caused a modest negative deflection in the baseline (Fig. 1).

**CD9 expression on pre-B acute lymphoblastoid leukemia cells is modulated by methionine–enkephalin.** Our studies showing that MET-ENK provoked physical changes in the NALM 6 cell membrane (Fig. 1) prompted us to investigate the possibility that this enkephalin also induced a rearrangement in membrane proteins. As indicated, (Figs. 2 and 3 and Table 1) a panel of mAb was used to assess the effects of MET-ENK on the expression of B cell surface determinants; samples of NALM 6, LAZ 221, and LAZ 388 cells were incubated with and without the enkephalin and the expression of surface markers measured with the use of immunofluorescent staining and flow cytometry. The incubation of NALM 6 or LAZ 221 cells with MET-ENK resulted in a transient increase in their expression of CD9 (Figs. 2 and 3). A brief incubation of the pre-B ALL cells with MET-ENK (i.e., \( \approx 10\) min) was sufficient to cause the enhancement in CD9 expression (Fig. 3 A). This enhanced level of CD9 expression was sustained for periods of 60 min but declined to near baseline by 120 min (Fig. 3 A). The induction of CD9 surface molecules by MET-ENK was dose dependent (Fig. 3 B). CD9 expression was increased, albeit modestly, by the low MET-ENK dose of 0.01 nM, further increased with the greater 100 nM dose, and still further increased, with the maximum dose (10,000 nM) (Fig. 3 B). Whereas MET-ENK caused an augmentation in the CD9 molecules displayed on the surfaces of pre-B ALL cells the enkephalin had no effect on their expression of CD10 (Figs. 2 and 3 A). These findings show that the MET-ENK-stimulated enhancement in CD9 expression is specific and not solely the result of an enkephalin-mediated increase in cell surface area.

When the pre-B ALL cells were incubated in a mixture of MET-ENK and a 100-fold excess of naloxone the induction of CD9 by MET-ENK was totally blocked indicating the specificity of the enkephalin response (Fig. 3 C).

Immunofluorescent staining for other adhesion molecules

![Figure 1. Methionine–enkephalin induces a morphological change in NALM 6 cells. NALM 6 cells (10^5/ml) were preincubated in PBS alone or containing 10^-3 M naloxone for 15 min on ice, stimulated with the indicated concentrations of MET-ENK, and then incubated for an additional 5 min at 37°C. Analysis of forward and right angle light scatter was performed using a Becton Dickinson FACScan® flow cytometer as described in Methods. Histograms of light scatter were obtained by analyzing 10,000 cells per sample.](https://doi.org/10.1172/JCI118171)

![Figure 2. Methionine–enkephalin causes a discrete, transient up-regulation in the CD9 molecules expressed on the surface of NALM 6 cells. NALM 6 cells (10^5/ml) were incubated at 37°C with MET-ENK (10^-3 M) for 0 (A and B) or 30 min (C and D). Cells were then transferred to an ice bath, pelleted by centrifugation, and stained as described in Methods. Data are shown as histograms depicting the distribution of 10,000 cells labeled with anti-CD9 (A and C) or CD-10 (B and D) mAb. The anti-CD9 and anti-CD10 mAb used for these experiments were isotype matched (IgG1).](https://doi.org/10.1172/JCI118171)
Figure 3. Methionine–enkephalin causes a rapid and dose-dependent increase in the CD9 but not CD10 molecules expressed on pre–B leukemic cells. Data are presented as the arithmetic/linear mean relative fluorescence intensities measured for 10,000 cells labeled with anti-CD9 or CD10 mAb. Cell samples were prepared and stained for immunofluorescent analysis as described for Fig. 2 and in Methods. In panel A, NALM 6 cells were incubated with MET-ENK (10^-7 M) for the times indicated. In panel B, LAZ 221 cells were incubated with MET-ENK at the doses indicated for 5 min at 37°C. In panel C, LAZ 221 cells were incubated at 37°C for 15 min with MET-ENK (10^-7 M) (— — —) or with both MET-ENK and naloxone (10^-5 M) (○—○). The anti-CD9 and anti-CD10 mAb used for these experiments were of the IgG2a isotype.

### Table 1. Methionine-Enkephalin Effects on the Surface Expression of Lymphocyte Determinants

<table>
<thead>
<tr>
<th>Cell determinant</th>
<th>Antibody isotype</th>
<th>Fluorescence*</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>NALM 6</td>
</tr>
<tr>
<td></td>
<td>(-)</td>
<td>(+)</td>
</tr>
<tr>
<td>CD9</td>
<td>IgG1</td>
<td>140.8</td>
</tr>
<tr>
<td>CD10</td>
<td>IgG1</td>
<td>37.1</td>
</tr>
<tr>
<td>Class II (HLA-DR)</td>
<td>IgG1</td>
<td>14.0</td>
</tr>
<tr>
<td>Class I (HLA-A,B,C)</td>
<td>IgG1</td>
<td>225.9</td>
</tr>
<tr>
<td>CD11a (LFA1α)</td>
<td>IgG1</td>
<td>ND^a</td>
</tr>
<tr>
<td>CD11b (Mac 1)</td>
<td>IgG1</td>
<td>1.0</td>
</tr>
<tr>
<td>CD18 (Integrin β chain)</td>
<td>IgG1</td>
<td>1.1</td>
</tr>
<tr>
<td>CD54 (ICAM-1)</td>
<td>IgG1</td>
<td>9.3</td>
</tr>
<tr>
<td>CD44 (homing receptor)</td>
<td>IgG1</td>
<td>1.0</td>
</tr>
<tr>
<td>CD19</td>
<td>IgG1</td>
<td>ND</td>
</tr>
<tr>
<td>CD58 (LFA3)</td>
<td>IgG1</td>
<td>1.0</td>
</tr>
<tr>
<td>CD22</td>
<td>IgG1</td>
<td>2.3</td>
</tr>
<tr>
<td>CD20</td>
<td>IgG1</td>
<td>1.5</td>
</tr>
<tr>
<td>CD72</td>
<td>IgG2a</td>
<td>1.6</td>
</tr>
<tr>
<td>CD29 (VLA-β chain)</td>
<td>IgG1</td>
<td>8.4</td>
</tr>
<tr>
<td>VLA-β chain; DE9N)</td>
<td>IgG1</td>
<td>13.6</td>
</tr>
<tr>
<td>VLA-1</td>
<td>IgG1</td>
<td>1.1</td>
</tr>
<tr>
<td>VLA-3</td>
<td>IgG1</td>
<td>1.1</td>
</tr>
<tr>
<td>VLA-5</td>
<td>IgG1</td>
<td>5.2</td>
</tr>
<tr>
<td>CD49b (VLA-α2 chain)</td>
<td>IgG1</td>
<td>1.0</td>
</tr>
<tr>
<td>CD49f (VLA-α6 chain)</td>
<td>IgG1</td>
<td>1.8</td>
</tr>
<tr>
<td>CD49d (VLA-α4 chain)</td>
<td>IgG1</td>
<td>3.9</td>
</tr>
<tr>
<td>CD23</td>
<td>IgG1</td>
<td>1.8</td>
</tr>
<tr>
<td>CD45 (T200)</td>
<td>IgG2a</td>
<td>1.1</td>
</tr>
</tbody>
</table>

* Cells were labeled for analysis as described for Figs. 2 and 3 and in Methods. Fluorescence values were measured from histograms collected for samples of 5,000 or 10,000 cells. Data are expressed as ratios: [arithmetic-linear mean relative fluorescence intensity measured for indicated mAb]/[(arithmetic-linear mean relative fluorescence intensity measured for isotype matched control)]. ^ Cells were incubated for 30 min at 37°C in PBS alone (—) or PBS containing 10^-7 M MET-ENK (+). ^ ND = not done. ^ Two anti-VLA-β1 (CD29) mAb were used for these studies; the data obtained for these mAb are listed for anti-CD29 (VLA-β chain) and VLA-β; DE9N.

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stained with Congo red dye as previously described (11). Cells which migrated into the filter were enumerated utilizing fluorescence microscopy and an Optimax V image analyzing system. 10 high power fields (HPF) were measured at 10-μm distances through the filter. Data are presented as the means ± SD for 30 values (i.e., 10 readings × triplicate assays). Note: the scales for the y-axis are different in panels A and B.

including VLA 4 (CD49d) or -5 (β-1 integrins) and the common VLA β1 chain (CD29) was equivalent on NALM 6 cell samples incubated with (+) or without (−) MET-ENK (Table I). With the pre-B ALL cells the fluorescence intensity measured for other β-1 integrins (VLA-1, VLA-3, CD49b, or CD49f) or for β-2 integrins (CD11a, CD11b, and CD18) was near that of the IgG1 isotype control (i.e., between 1 and 1.5 times the control value) regardless of whether the cells were incubated with (+) or without (−) MET-ENK (Table I).

As expected, immunofluorescent staining of the LAZ 388 cells revealed a phenotype distinct from that of the pre-B ALL lines and characteristic of mature EBV-transformed B cells (Table I). With the LAZ 388 cells the level of cell surface staining for CD9 and the β-1 and β-2 integrins as well as other determinants was equivalent for the MET-ENK treated (+) and non-treated (−) samples (Table I).

**Enkephalins stimulate a naloxone-reversible enhancement in the migration of pre-B acute lymphoblastoid leukemia cells.**

MET-ENK was a potent stimulus for the migration of NALM 6 and LAZ 221 (pre-B ALL), but not LAZ 388 (EBV-transformed) cells (Fig. 4 and Table II). The number of NALM 6 or LAZ 221 cells which migrated to MET-ENK was greater than that which responded to the assay buffer, alone (spontaneous locomotion) (P = 0.01) (Fig. 4 and Table II). The effect of MET-ENK on the pre-B ALL cell migration was dose dependent (Fig. 4). MET-ENK was stimulatory over a four log dose range (0.01–100 nM); the low MET-ENK dose of 0.01 nM resulted in a modest response which was increased with the greater 1 nM dose and further increased with the maximum 100 nM enkephalin dose (Fig. 4).

Like MET-ENK the synthetic enkephalins DADLE and DAMGO enhanced the migration of pre-B ALL cells (P ≤ 0.01); the potency of these synthetic enkephalins was near to that measured for the naturally occurring pentapeptide MET-ENK (Table II). Unlike the enkephalins, the neutrophil chemotactic agent FMLP and the nonanalgesic peptide DesTyr-Leu-ENK had no effect on the migration of pre-B ALL cells (P, NS ≥ 0.05).

The migration of LAZ 388 cells to MET-ENK, DADLE, DAMGO, DesTyr-Leu-ENK, or FMLP was equivalent to their spontaneous locomotion (response to assay buffer, alone) (P, NS ≥ 0.05) (Table II).

Naloxone inhibited the migration of NALM 6 cells to MET-ENK (Table III). Spontaneous locomotion resulting from ran-

**Table II. The Pre-B Acute Lymphoblastoid Leukemia Cell Lines Nalm 6 and LaZ 221 Migrate to Natural and Synthetic Enkephalins**

<table>
<thead>
<tr>
<th>Cells</th>
<th>Spontaneous locomotion</th>
<th>FMLP (nonanalgesic)</th>
<th>DesTyr-Leu-ENK (nonanalgesic)</th>
<th>MET-ENK</th>
<th>DAMGO</th>
<th>DADLE</th>
</tr>
</thead>
<tbody>
<tr>
<td>NALM 6</td>
<td>2190</td>
<td>3198 (P, NS; ≥ 0.05)</td>
<td>3085 (P, NS; ≥ 0.05)</td>
<td>6952 (P = 0.01)</td>
<td>5070</td>
<td>6240</td>
</tr>
<tr>
<td>LAZ 221</td>
<td>1886</td>
<td>2906 (P, NS; ≥ 0.05)</td>
<td>2010 (P, NS; ≥ 0.05)</td>
<td>6199 (P = 0.01)</td>
<td>4602</td>
<td>4405</td>
</tr>
<tr>
<td>LAZ 388</td>
<td>1938</td>
<td>1914 (P, NS; ≥ 0.05)</td>
<td>2244 (P, NS; ≥ 0.05)</td>
<td>1986 (P = 0.01)</td>
<td>2043</td>
<td>2283</td>
</tr>
</tbody>
</table>

* Assays were as described for Fig. 4. 1 Migration was scored as described for Fig. 4. The numbers presented are cell counts obtained by adding the values measured at 10-μm intervals through the filters. 1 The statistical significance of differences between the treatments was analyzed by a Dunnett's multiple comparison test.
Table III. Naloxone Inhibits the Migration of NALM 6 Cells to Methionine-Enkephalin

<table>
<thead>
<tr>
<th>Spontaneous locomotion</th>
<th>MET-ENK (1 nM) Without</th>
<th>Control</th>
<th>MET-ENK (100 nM) Without</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cell counts(^{*})</td>
<td>1,647</td>
<td>1,350</td>
<td>2,925</td>
</tr>
<tr>
<td>P values(^{\dagger})</td>
<td>NS; (\geq 0.05)</td>
<td>0.01</td>
<td>(0.01)</td>
</tr>
<tr>
<td>Percent response(^{\ddagger})</td>
<td>82</td>
<td>82</td>
<td>42</td>
</tr>
</tbody>
</table>

\(*\) NALM 6 cells were cultured in RPMI 1640 medium supplemented with 2% FBS, 100 \(\mu\)g/ml of penicillin/streptomycin, and 2 mM glutamine or in the culture medium containing 10\(^{-5}\) M naloxone for 15 h at 37°C in a 5% CO\(_2\) atmosphere. Viability of the cells incubated with or without naloxone was equivalent (\(\geq 96\%\)) as judged from vital staining with trypan blue. NALM 6 cells treated with naloxone were added to the upper wells of the migration chamber in assay buffer that contained naloxone (10\(^{-5}\) M). Migration was allowed to proceed for 75 min at 37°C. \(^{\dagger}\) Without = cells that were incubated in assay buffer alone; with = cells that were incubated in 10\(^{-5}\) M naloxone. \(^{\ddagger}\) Migration was scored as described for Fig. 4. The numbers presented are cell counts obtained by adding the values measured at 10-μm intervals through the filters. \(^{\dagger\dagger}\) The statistical significance of differences between treatments was analyzed by a Student's \(t\) test. Percent = ([Sum for cells incubated with naloxone]/[Sum for cells incubated without naloxone]) \(\times 100\%\).

Table IV. Migration of NALM 6 and LAZ 221 Cells Is Inhibited by Preincubation with Anti-CD9 Monoclonal Antibody

<table>
<thead>
<tr>
<th>MET-ENK (nM)</th>
<th>Control</th>
<th>Monoclonal antibody</th>
</tr>
</thead>
<tbody>
<tr>
<td>(Buffer, alone)</td>
<td>anti-CD10 IgG2a</td>
<td>anti-CD10 IgG1 IgG2a</td>
</tr>
<tr>
<td>NALM 6 cells</td>
<td>0.01</td>
<td>135 160 175</td>
</tr>
<tr>
<td>LAZ 221 cells</td>
<td>100</td>
<td>463 367 448</td>
</tr>
</tbody>
</table>

\(*\) Cells (10\(^{7}\)/ml) were preincubated in the migration assay buffer or the buffer containing anti-CD10, -CD29 (DE9N), or -Class II (LB 3.1), Class I (W632) or -CD9 mAb for 30 min at 4°C, then pelleted by centrifugation and washed twice in assay buffer before their addition to the upper wells of the chemotaxis chamber. The mAb doses used for these studies (1/100 dilution of ascites) were saturating as predetermined by immunofluorescent staining and flow cytometry. \(^{\dagger}\) Assays were as described for Fig. 4. Cells which migrated 40 \(\mu\)m from the origin were identified and scored visually with the use of a fluorescence microscope. The numbers presented are the cell counts determined for 10 high power fields \(\times\) triplicate wells.

Discussion

Most cancer-related deaths result from the dissemination and metastasis of tumor cells; therefore, it is important to identify the signals that regulate the migration of malignant cells. In this study factors which regulate the in vitro migration of leukemia cells were identified. The naturally occurring opioid pentapeptide MET-ENK as well as the synthetic enkephalin analogs DADLE and DAMGO were shown to be stimulants for the migration of NALM 6 and LAZ 221 pre-B ALL cells (Fig. 4 and Table II). In a previous study (11) we showed that opioid peptides were potent signals for the migration of normal lymphocytes causing both an enhancement in their directional (chemotaxis) and random movement (chemokinesis). We found, as previously reported for the normal lymphocytes (11), that activation of the pre-B ALL cells with MET-ENK caused an increase in both their nondirectional and directional migration (data not shown). These findings identify opioid peptides as signals for the in vitro migration of cultured human leukemia cells.

The pre-B ALL cells responded to physiological doses of MET-ENK (Figs. 1 and 4). Circulating and local tissue levels of the enkephalins have been difficult to quantitate because of their rapid hydrolysis (4); however, in brain these peptides have been shown to reach concentrations of at least 0.1 nM (32). The level measured in brain samples (0.1 nM) is 10 times the MET-ENK dose (0.01 nM) that induced near maximum activation of the morphological response (Fig. 1) and that caused significant increases in the pre-B ALL cell migration (Fig. 4).

It is of note that the LAZ 221 and NALM 6 pre-B ALL cells express enkephalinase (CD10; neutral endopeptidase) on their surfaces (see Fig. 2). This enzyme hydrolyzes and inactivates MET-ENK (33) and may be important in down-regulating responses to MET-ENK by either decreasing the enkephalin concentration in the surrounding medium or by degrading cell-associated MET-ENK. The migration of NALM 6 and LAZ...
221 cells to MET-ENK was dose dependent; whereas the 0.01 nM dose stimulated a modest increase in migration, the response was increased when the dose was titered to 10 nM, and still further increased with the maximum enkephalin dose of 100 nM (Fig. 4). This dose dependency may have resulted, at least in part, from endogenous enkephalinase activity and MET-ENK hydrolysis.

In our studies we measured the effects of MET-ENK on cell morphology using flow cytometric analysis of forward and right angle light scatter (Fig. 1). In a previous report (19) the effects of opioid peptides and other chemotactic factors on the shape of granulocytes were monitored by light and electron microscopy. The opioid peptides, like other chemotactic factors, were seen to activate a morphological response before the onset of cell migration (19). This response was characterized by membrane spreading as well as cell elongation and flattening. Our studies showed that only a brief exposure (≤5 min) of the pre-B ALL cells to low doses of MET-ENK (0.01 nM) was sufficient to cause the morphological response (i.e., change in forward and right angle light scatter) (Fig. 1). These enkephalin effects were probably related to the spreading and flattening that cells undergo in preparation for movement. The opiate receptor antagonist naloxone inhibited the enkephalin-stimu-

lated reshaping of the pre-B ALL cells (i.e., the increases in light scatter) (Fig. 1). However, when applied alone, the antagonist caused a modest deflection in the baseline light scatter (Fig. 1). The antagonist was not directly cytocidal because the viability that we measured (i.e., trypan blue exclusion) was equivalent (≥96%) for naloxone-treated and nontreated NALM 6 cells. Other investigators have reported that naloxone has growth inhibitory effects on some tumor cells (20, 34) and the naloxone effect on light scatter that we observed may be related to such antiproliferative activity.

MET-ENK was a potent stimulant for the migration of NALM 6 and LAZ 221 pre-B ALL cells but not for the EBV-transformed, mature LAZ 388 cell line (Table II). The responsive LAZ 221 and unresponsive LAZ 388 cells have the identical genetic background since they were derived from one donor (33); however, as expected these cell lines were found to be dissimilar with respect to phenotype (Table I). The responsive LAZ 221 cells exhibited the characteristic phenotype of pre-B ALL cells (e.g., CD9+, CD10+, CD18+) whereas the unresponsive LAZ 388 cells displayed the surface markers (e.g., CD11a+, CD18+, CD44+) of an EBV-transformed cell line (Table I). Other investigators (9) have been unable to demonstrate the presence of classical, naloxone-sensitive MET-ENK
receptors on EBV-transformed lymphocytes and it is likely that the LAZ 388 cells are unresponsive to MET-ENK simply because of a deficiency in enkephalin binding sites. It is known that EBV transformation renders cells unresponsive to normal activation signals (35) and it is possible, therefore, that the LAZ 388 cells are insensitive to MET-ENK by virtue of their viral transformation.

Activation of NALM 6 or LAZ 221 cells with MET-ENK correlated with an increase in migration (Fig. 4) and an enhancement in surface expression of CD9 (Figs. 2 and 3) but not other adhesion molecules (e.g., CD4, CD44, β1, or β2 integrins) (Table I). Interestingly, a similar relationship has been described by Letarte et al. (25) for pre-B ALL cell aggregations and the expression of adhesion receptors. These investigators (25) reported that the size and number of homotypic aggregates formed by various pre-B ALL cell lines correlated with the surface level of CD9, but not other adhesion molecules including CD11a, CD18, CD19, CD44, or CD54. Our studies with human pre-B ALL cells have linked CD9 expression with an important leukocyte function (migration).

The precise mechanism(s) by which CD9 promotes or strengthens adhesions has not, as yet, been identified. CD9 is a 24–27-kD glycoprotein that belongs to the novel superfamily of proteins having four transmembrane domains (TM4SF) (36–39). CD9 as well as other proteins belonging to the same superfamily (i.e., TAPA-1 and M33) form multimeric complexes with other adhesion molecules (40–44) and it is possible that such multimeric complexes are important for cell adhesion and migration.

Preincubation of NALM 6 or LAZ 222 cells with anti-CD9 but not anti-CD10, anti-

References


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