

Altered Aldose Reductase Gene Regulation in Cultured Human Retinal Pigment Epithelial Cells

Douglas N. Henry,^{**} Monte Del Monte,[§] Douglas A. Greene,[‡] and Paul D. Killen^{||}

The Departments of ^{*}Pediatrics and Communicable Diseases, [§]Ophthalmology, [‡]Internal Medicine and ^{||}Pathology, and the Michigan Diabetes Research and Training Center, University of Michigan, Ann Arbor, Michigan 48109

Abstract

Aldose reductase (AR2¹), a putative "hypertonicity stress protein" whose gene is induced by hyperosmolarity, protects renal medullary cells against the interstitial hyperosmolarity of antidiuresis by catalyzing the synthesis of millimolar concentrations of intracellular sorbitol from glucose. Although AR2 gene induction has been noted in a variety of renal and nonrenal cells subjected to hypertonic stress in vitro, the functional significance of AR2 gene expression in cells not normally exposed to a hyperosmolar milieu is not fully understood. The physiological impact of basal AR2 expression in such cells may be limited to hyperglycemic states in which AR2 promotes pathological polyol accumulation, a mechanism invoked in the pathogenesis of diabetic complications. Since AR2 overexpression in the retinal pigment epithelium has been associated with diabetic retinopathy, the regulation of AR2 gene expression and associated changes in sorbitol and *myo*-inositol were studied in human retinal pigment epithelial cells in culture. The relative abundance of aldehyde reductase (AR1) and AR2 mRNA was quantitated by filter hybridization of RNA from several human retinal pigment epithelial cell lines exposed to hyperglycemic and hyperosmolar conditions in vitro. AR2 but not AR1 mRNA was significantly increased some 11- to 18-fold by hyperosmolarity in several retinal pigment epithelial cell lines. A single cell line with a 15-fold higher basal level of AR2 mRNA than other cell lines tested demonstrated no significant increase in AR2 mRNA in response to hypertonic stress. This cell line demonstrated accelerated and exaggerated production of sorbitol and depletion of *myo*-inositol upon exposure to 20 mM glucose. Therefore, abnormal AR2 expression may enhance the sensitivity of cells to the biochemical consequences of hyperglycemia potentiating the development of diabetic complications. (*J. Clin. Invest.* 1993. 92:617–623.) Key words: diabetic retinopathy • aldehyde reductase • *myo*-inositol • sorbitol • osmolyte

Introduction

Hyperglycemia has been strongly implicated in the development of the chronic complications of diabetes, but the specific pathogenetic pathways and their possible modulation by genetic and environmental factors remain the subject of speculation and controversy (1). Metabolism of excess tissue glucose to sorbitol by the enzymes aldehyde and aldose reductase (AR1¹ [EC 1.1.1.2] and AR2 [EC 1.1.1.21], respectively) has recently received widespread consideration as a potentially important tissue-specific mechanism of glucose toxicity (2, 3). Some of the potential adverse effects of increased glucose flux through the sorbitol pathway have been ascribed to secondary alterations in intracellular *myo*-inositol (MI) metabolism (4) perhaps due to depletion of specific intracellular MI pools (5, 6). The mechanisms leading to depletion of intracellular MI is poorly understood, but may be due to the action of sorbitol and MI as alternative non-ionic intracellular osmolytes (7). Accumulation of intracellular sorbitol may be accompanied by reciprocal decreases in other intracellular osmolytes, such as MI, betaine, or glycerophosphorylcholine in order to maintain osmotic equilibrium. Other mechanisms by which increased flux through the sorbitol pathway might cause tissue damage include perturbation in the NADP⁺/NADPH and NAD⁺/NADH redox couples linked to the AR and sorbitol dehydrogenase oxidoreductase enzymes, respectively (8, 9), and/or direct osmotic effects of accumulated intracellular sorbitol (10).

Osmotic stress has recently emerged as a major regulator of the genes for AR2 and the Na⁺-MI cotransporter in the renal medulla, where sorbitol and MI function physiologically as intracellular osmolytes that compensate for the high interstitial osmolarity required for antidiuresis (7, 11). Under these circumstances, elevated extracellular osmolarity is thought to increase AR2 and MI-transporter gene expression, protein synthesis, and activity (11, 12). Increased AR2 gene expression, immunoreactivity and enzymatic activity in a variety of tissues and cells have also been associated with diabetes and its chronic complications in man and animals (13–15). AR2 immunoreactivity is increased in the retinal pigment epithelial (RPE) cell layer in eyes obtained at autopsy from diabetic subjects with and without proliferative diabetic retinopathy, with a greater increase in the former (13). The RPE cell layer exhibits sorbitol accumulation, MI depletion, and impaired Na⁺/K⁺-ATPase activity in diabetic rabbits (16, 17). Moreover, electroretinographic impairment in diabetic pigmented rats is ascribed to dysfunction of the RPE cell layer and is corrected by either aldose reductase inhibitors (ARI) or by MI supple-

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Address reprint requests to Douglas A. Greene, M.D., Division of Endocrinology and Metabolism, Department of Internal Medicine, University of Michigan Medical Center, 3920 Taubman Center, Box 0354, 1150 East Medical Center Drive, Ann Arbor, MI 48109-0354.

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1. *Abbreviations used in this paper:* ARI, aldose reductase inhibitors; AR1, aldehyde reductase; AR2, aldose reductase; MI, *myo*-inositol; RPE, retinal pigment epithelial.

mentation (18). Therefore, cultured RPE cells from a variety of mammalian species including humans have been examined as an in vitro model of biochemical and functional defects that may contribute to the pathogenesis of some diabetic complications in vivo (19, 20). Human RPE cells in culture demonstrate glucose-induced, ARI-sensitive MI depletion, which is associated with a reversible reduction in the rate of uptake of ^{125}I -labeled human retinal rod outer segments, an important function of the RPE cell (20). The supposition that altered AR2 (and/or AR1) gene expression might predispose diabetic patients to chronic tissue-specific complications such as retinopathy prompted this study of AR1 and AR2 gene modulation by osmolarity in RPE cells. The results demonstrate that aberrant AR2 gene expression is associated with accelerated sorbitol accumulation and MI depletion in response to pathophysiological levels of glucose, suggesting that dysregulation of the AR2 gene may be a predisposing factor to glucose-induced metabolic abnormalities in RPE cells and, by extrapolation, in other tissues prone to diabetic complications.

Methods

Reagents and supplies. Unless otherwise indicated, culture media, antibiotics, and trypsin were obtained from GIBCO BRL (Life Technologies Inc.), Gaithersburg, MD; fetal bovine and calf serum from Hyclone Labs, Logan, UT; culture dishes, wells, and flasks from Falcon (Becton-Dickinson and Co., Lincoln Park, NJ); chemicals and reagents (highest available purity) from Sigma Chemical Co., St. Louis, MO; and radioisotopes were from Amersham Corp., Arlington Heights, IL.

Cell culture techniques and experimental design. Primary cultures of RPE cells were established and maintained by the method of Del Monte and Maumenee (21) as recently modified (20). In brief, RPE cells were recovered with fire-polished Pasteur pipettes under direct observation with a dissecting microscope from surgically removed, sagittally bisected eyes, obtained from the Michigan Eye Bank within 24 hr post-mortem, and cultured in Ham's F12 nutrient medium containing 16% FBS, 2 mM glutamine, 100 U/ml penicillin, 100 $\mu\text{g}/\text{ml}$ streptomycin, and 26 mM sodium bicarbonate in a 37°C, humidified 5% CO_2 atmosphere. Colony-forming primary cultures were rinsed with Hanks' balanced salt solution and passaged with 0.5 mg/ml trypsin and 0.53 mM EDTA in normal saline at 37°C for 10 min, centrifuged for 5 min at 50 g , resuspended in new culture medium and replated (21). The established RPE cell lines were passaged at a density of 40–100,000 cells/ cm^2 in 25 cm^2 or 75 cm^2 flasks in MEM with 2 mM L-glutamine ([MI] = 11.1 μM) containing 20% calf serum ([MI] = 90–120 μM) and 5 mM glucose (20). All experiments were performed between passages 19 and 24 of 4 RPE cell lines described in Table I. Passaged cells were plated to yield near-confluent cultures at the end of 1- and 7-d experiments (20). Six-well plates (9.62 cm^2) or 175 cm^2 dishes were used for metabolic studies or RNA isolation, respectively. The freshly plated cells were allowed to attach in standard growth medium for 24 h and then incubated for various times in MEM containing 5% calf serum and various supplements (see below). RPE

cells (lines 125, 45 and 91) were incubated in medium containing 300 mM glucose for 0, 2, 4, 8, 16 or 24 h to determine when AR2 mRNA induction was maximal. Subsequent experiments (RPE cell lines 125, 0308, 45, and 91) compared the effect of 300 mM glucose, 3-*O*-methylglucose or mannitol on AR2 mRNA levels attained at each time point. The effect of various concentrations of glucose on AR1 and AR2 mRNA were examined after 24 h. These studies were performed in triplicate using RPE 125 or 45 cells. Further experiments compared the response of 4 RPE cell lines to 20 mM or 300 mM glucose at 24 h or 7 d. 20 mM glucose simulated physiological hyperglycemia (20) where flux through the polyol pathway would be stimulated primarily by mass action (8). 300 mM glucose was considered a combined osmotic stimulus in the presence of maximal substrate concentrations for the AR reactions. Cultures were routinely examined by phase contrast microscopy and survival estimated by trypan blue exclusion. Neither cell morphology nor survival was detectable altered by the experimental conditions employed in these studies.

Preparation of AR1 and AR2 cDNA from RPE mRNA by PCR amplification. Total RNA was purified from cultured human RPE cells by a modification of the method of Stallcup and Washington (22). Oligo-dT primed cDNA was prepared from 20 μg of total RPE cell RNA and reverse transcribed with Superscript (Bethesda Research Laboratories, Gaithersburg, MD) according to the vendor's protocol. AR1 and AR2 cDNAs were selectively PCR amplified using Amplitaq (Perkin-Elmer Corp., Norwalk, CT), the vendor's buffer and synthetic oligonucleotide primers for human hepatic AR1 (5'TATAAGCTTGCTACGACTCCACCCACTA3' and 5'ATATCTAGATGHCTTCATCTCTTCTGGGCT3') and placental AR2 (5'TATAAGCTT-AGAAGCTCAGGGAGCAGGT3' and 5'ATATCTAGAGGTCAC-CACGATGCCTTTG3') cDNA (23), anchored with either HindIII or XbaI sites. Oligonucleotides were synthesized by the DNA Synthesis Core Facility, a part of the University of Michigan's Biomedical Research Core Facilities, on automated DNA synthesizers (Applied Biosystems, Inc., Foster City, CA) employing β -cyanoethyl phosphoramidite chemistry on controlled pore glass support. After 30 cycles (95°C, 1.5 min denaturation; 54°C, 1.5 min annealing; and 72°C, 1.5 min extension) a single major amplification product was identified for each primer pair. The subcloned amplification products obtained with AR1 or AR2 primers each exhibited 100% sequence identity with human hepatic AR1 and placental AR2 cDNAs (23), respectively, but had only 35% nucleotide identity with each other. The purified cDNA inserts for RPE AR1 and AR2 were labeled with random primers and [^{32}P]dCTP (3000 Ci/mM; Amersham) to a specific activity of 10⁹ dpm/ μg (24).

Filter hybridization of RNA from RPE cells. 10 μg of denatured total RNA was resolved on 2.2 M formaldehyde 1% agarose gels, transferred by capillary blotting to Zetabind filters (Cuno, Inc., Meriden, CT) stained with methylene blue and photographed. Because of the large differences in the abundance of AR2 mRNA in many of the samples, quantitation was performed with serial dilutions (64- to 128-fold) of RNA applied with a vacuum manifold to Zetabind filters. The filters were fixed in vacuo at 80°C for 2 h, and prehybridized at 65°C in 0.5 M NaH_2PO_4 pH 7.0, 1 mM EDTA, 7% SDS, and 1% BSA for 4–6 h before adding the denatured AR2 cDNA probe (10⁶ dpm/ml) (25). After 18 h, filters were washed twice with 40 mM NaH_2PO_4 pH 7.0, 1 mM EDTA, 7% SDS at 65°C, and 4 times with 40 mM NaH_2PO_4 pH

Table I. Human Retinal Pigment Epithelial Cell Lines Used in These Studies

Cell line	Male/Female	Age	Cause of death	Hours to enucleation	Hours to culture	Passage number
125	Male	12	Gunshot	8	22	24
0308	Male	15	MVA	5	14	23
45	Female	50	CVA	5.75	21	21
91	Female	54	CVA	2.5	21	19

7.0, 1 mM EDTA, 1% SDS at 65°C, and exposed to preflashed photographic film (X-Omat; Eastman-Kodak, Rochester, NY) at -80°C in cassettes with intensifying screens. Exposures of the dot blot filters within the linear range of the film were quantitated by densitometry. Filters were subsequently stripped, reexposed, and, when free of residual probe, sequentially rehybridized with AR1, chicken β -actin (26), or human vimentin cDNA probes (27). Densitometry results from three replicate samples obtained in three independent experiments were normalized for the abundance of β -actin mRNA, and statistical analysis was performed using analysis of variance with repeat measurements design (28).

Measurement of sorbitol and MI in cultured RPE cells. Confluent monolayers of RPE cells in six-well plates were rinsed with 3 ml of serum-free MEM containing 3.5 mg/ml BSA, lysed in 1 ml distilled H₂O for 15 min at 37°C, and scraped with a rubber policeman into 3-ml plastic tubes, sonicated, and deproteinized with balanced aliquots of 5% ZnSO₄ and Ba(OH)₂ followed by centrifugation. Aliquots of the supernatant were combined with the internal standard α -D-mannoside, lyophilized, derivitized with a 10:2:1 solution of dried pyridine-hexamethylsilane-trimethylchlorosilane (Pierce, Rockford, IL), evaporated to dryness under N₂, resolubilized in hexane and injected into a gas-liquid-chromatograph (3400; Varian Instruments, Palo Alto, CA) fitted with a flame ionization detector and a 30-m DB-Megabore column (J&W Sci., Folsom, CA) with helium as a carrier, and programmed from 160 to 210°C as previously described (20). Peak areas were digitally quantitated with a Varian 402 data system, corrected for the internal standard, and transformed to mass measurements on the basis of daily standard curves for α -D-mannoside, sorbitol, and MI (20). Statistical analysis was performed using analysis of variance with repeat measurements design (26).

Results

Induction of AR2 mRNA but not AR1 mRNA in RPE cells in vitro. The increase in AR2 mRNA induced by glucose in RPE cell lines 125, 0308, and 45 was time and concentration dependent (Figs. 1 and 2). Increased AR2 mRNA was first evident after 8 h exposure to 300 mM glucose, achieving near maximal steady-state levels after 16 h (Fig. 1). 7 d exposure to 300 mM glucose resulted in only a slight (30%) decline in AR2 mRNA from the maximal level noted at 24 h (data not shown). Increased AR2 mRNA was evident after 24 h exposure to medium containing 150 mM glucose (440 mosmol/kg), however no significant increase in AR2 mRNA was noted at lesser glu-

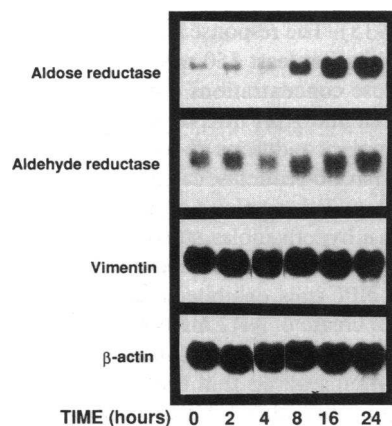
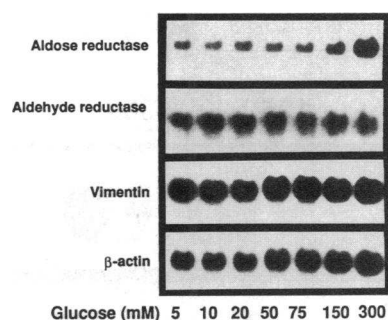


Figure 1. Time course of AR2 mRNA induction by 300 mM glucose. RNA was isolated from RPE cell line 125 after 0, 2, 4, 8, 16, and 24 h exposure to 300 mM glucose. A Northern blot was sequentially hybridized with AR1, AR2, β -actin, and vimentin cDNA probes. Increased AR2 mRNA was first noted after 8 h exposure and was near maximal by 16 h. No significant change in AR1, vimentin, or β -actin mRNA was evident.



ing exposure to 150 mM glucose and was near maximal at 300 mM glucose. No significant change in AR1, β -actin, or vimentin mRNA was apparent at any glucose concentration.

cos concentrations (Fig. 2) even after 7 d (data not shown). No significant change in AR1, vimentin or β -actin mRNA was noted at any glucose concentration at any time (Figs. 1 and 2). The effect of 300 mM glucose was reproduced by an equimolar concentration of the nonmetabolizable glucose analogue, 3-O-methylglucose, or by mannitol, an osmotically active non-ionic compound that does not undergo facilitated transport (Fig. 3).

Constitutive AR2 gene expression in RPE 91 cells. An effect of glucose on the expression of AR2 mRNA was significant only in RPE 125, 0308, and 45 cells ($P < 0.02$). Exposure to medium supplemented with 300 mM glucose (590 mosmol/kg) resulted in a marked increase (17.7-, 17.7-, and 11.3-fold, respectively) in AR2 mRNA after 24 h (Figs. 4 and 5). In agreement with previous data, 20 mM glucose has no effect on AR2 mRNA levels in any cell line ($P > 0.05$). The mRNA encoding AR1, another oxido-reductase capable of metabolizing glucose to sorbitol, was not significantly changed by this treatment, and no significant change in vimentin or β -actin mRNA was noted (Fig. 4). In contrast, the basal level of AR2 mRNA in RPE 91 cells (isotonic medium, 5 mM glucose) was 14- to 17-fold higher than the basal AR2 mRNA level in RPE 125, 0308, or 45 cells ($P < 0.001$), and there was no significant increase in AR2 mRNA after exposure to 300 mM glucose for 24 h or 7 d (Figs. 4-6).

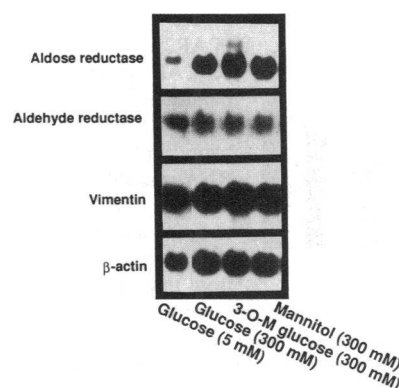


Figure 3. Induction of AR2 mRNA by glucose, 3-O-methylglucose, and mannitol. RNA was isolated from RPE cell line 45 after 24 h exposure to 5 mM glucose, 300 mM glucose, 300 mM 3-O-methylglucose, and 300 mM mannitol. A Northern blot was sequentially hybridized with AR1, AR2, β -actin, and vimentin cDNA probes. Glucose, 3-O-methylglucose, a nonmetabolizable glucose analogue, and mannitol, an osmotically active, non-ionic compound that does not undergo facilitated transport, were similarly effective in increasing AR2 mRNA levels. No significant change in AR1, vimentin, or β -actin mRNA was evident.

RPE CELL LINE

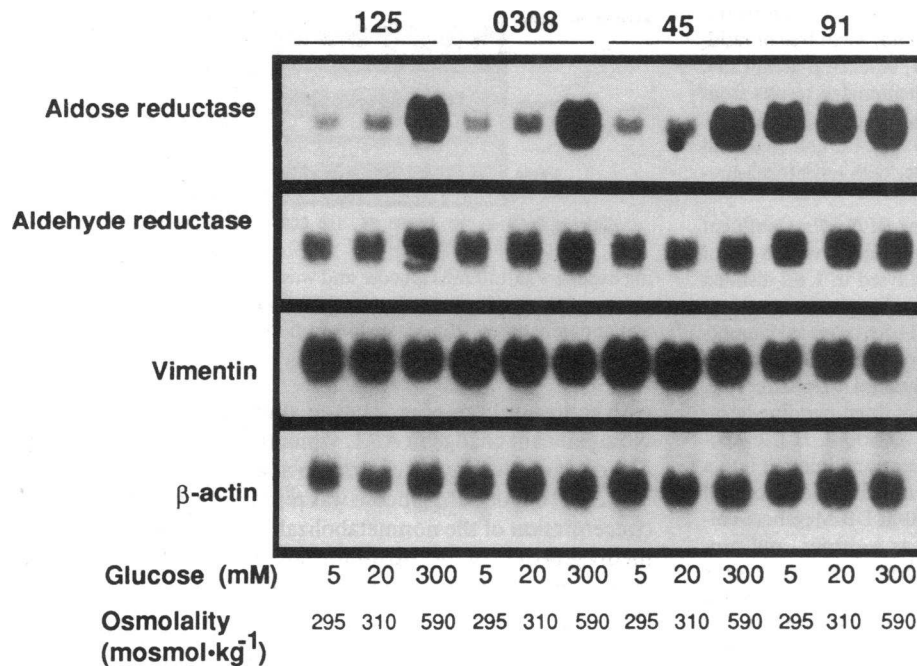


Figure 4. Osmotic induction of AR2 mRNA in different human RPE cell lines. RNA was isolated from four different RPE cell lines after 24 h exposure to 5, 20, or 300 mM glucose. A Northern blot was sequentially hybridized with AR1, AR2, β -actin, and vimentin cDNA probes and autoradiograms were prepared. AR1 and AR2 mRNA were normalized to β -actin as described in Methods. RPE cell lines 125, 0308, and 45 had marked increases in AR2 mRNA in response to 300 mM glucose, but not to pathophysiologic hyperglycemia (20 mM glucose). In contrast, RPE cell line 91 had a high basal level of AR2 mRNA and demonstrated little increase in AR2 mRNA in response to 300 mM glucose. No significant change in AR1 was detected in any of these conditions when normalized to β -actin mRNA. No change in vimentin or β -actin mRNA was evident in any cell line under any condition.

Metabolic correlates of constitutive vs. regulated AR2 gene expression. The metabolic correlates of regulated or constitutive high level AR2 gene expression were assessed by comparing time-dependent changes in sorbitol and MI content in RPE 45 and 91 cells upon exposure to hyperglycemic (20 mM glucose, 310 mosmol/kg) (20) and hyperosmolar concentrations of glucose (300 mM glucose, 590 mosmol/kg) (Fig. 7). RPE 45 cells, with regulated AR2 gene expression, demonstrated little or no sorbitol accumulation and inconsistent MI depletion 24 h after exposure to either 20 or 300 mM glucose, and significant sorbitol accumulation (164.6 nmol/mg protein) and consistent MI depletion (32.4 nmol/mg protein to 8.3 nmol/mg protein, $P < 0.001$) only after 7 d in 300 mM glucose. In con-

trast, modest sorbitol accumulation (42.8 nmol/mg protein) was detectable in RPE 91 cells within 24 h of exposure to 20 mM glucose, and marked sorbitol accumulation and MI depletion were evident (sorbitol, 428.7 nmol/mg protein, $P < 0.001$; MI, 29.6 nmol/mg protein to 11.4 nmol/mg protein, $P < 0.001$) in 300 mM glucose. After 7 d, sorbitol accumulation (468.6 nmol/mg protein, $P < 0.001$) and MI depletion (44.0 nmol/mg protein to 1.7 nmol/mg protein, $P < 0.001$) was clearly exaggerated in RPE 91 cells compared with RPE 45 cells at 20 mM and 300 mM glucose.

Discussion

RPE cell lines from three human donors (RPE cell lines 125, 0308, and 45) exhibited low basal levels of AR2 mRNA when cultured in isotonic medium and an 11- to 18-fold increase in AR2 mRNA in response to 300 mM glucose. These results are similar to those reported in a variety of renal and nonrenal cells, including rat glomerular mesangial cells, dog kidney endothelial cells, dog and rat lens epithelial cells, and Chinese hamster ovary cells, (7, 12, 29–35). The response required 8 h, was maximal at 16 h, and required at least 150 mM glucose (445 mosmol/kg). Lesser glucose concentrations did not increase AR2 mRNA in any RPE cell line. AR1 mRNA was not significantly altered by hyperosmolar conditions up to 590 mosmol/kg, suggesting that AR1, an oxido-reductase capable of metabolizing glucose to sorbitol is unlikely to play a significant role in the cellular adaptation to a hyperosmolar stress. However, a fourth cell line (RPE 91), isolated and cultured under conditions identical to the other RPE cells, exhibited high basal AR2 gene expression and little increase of AR2 mRNA in response to hyperosmolarity produced by glucose, 3-*O*-methylglucose, or mannitol. This observation suggests that the expression of AR2 gene in RPE 91 cells is unregulated. Further support for

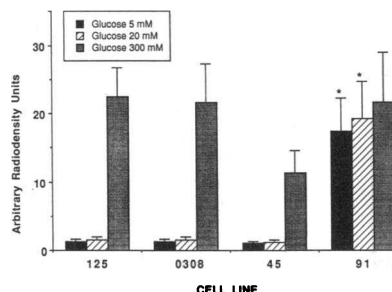


Figure 5. Quantitation of AR2 mRNA in RPE cells. AR2 mRNA was quantitated in total cellular RNA by dot blot hybridization of total RNA isolated 24 h after exposure of RPE cells to 5, 20, and 300 mM glucose. The results from three independent

experiments were quantitated using a laser densitometer. The radiodensity obtained was corrected for differences in β -actin abundance before statistical analysis. There was no difference in AR2 mRNA abundance between 5 and 20 mM glucose in any cell type. An effect of glucose on AR2 mRNA was evident only in RPE cell lines 125, 0308, and 45 ($P < 0.02$). The level of AR2 mRNA in RPE 91 cells exposed to 5 or 20 mM glucose was significantly higher than any other cell line ($*P < 0.001$). The basal level of AR2 mRNA in RPE 91 cells was 14- to 17-fold higher than that in any other cell type.

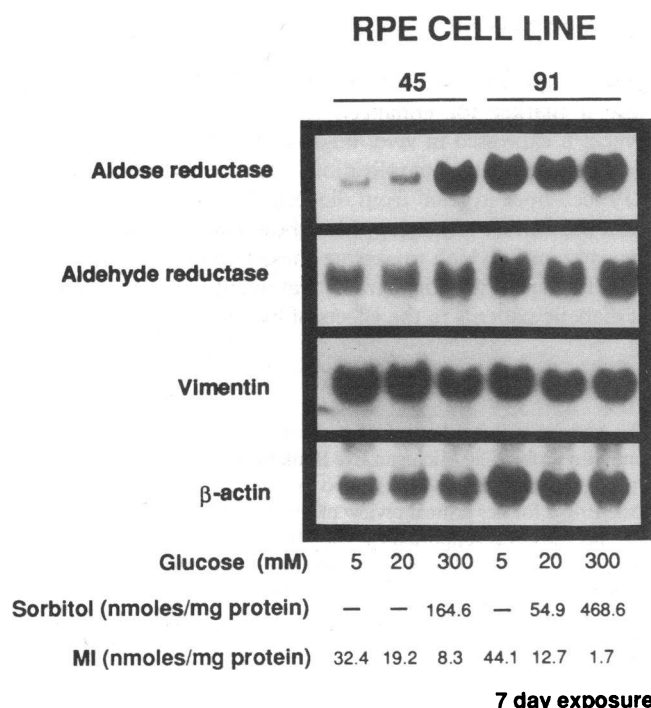


Figure 6. Effect of chronic exposure to glucose. RPE cell lines 45 and 91 were exposed to 20 mM or 300 mM glucose for 7 d before RNA isolation. A Northern blot was sequentially hybridized with AR1, AR2, β -actin, and vimentin cDNA probes. Twenty mM glucose did not increase AR2 mRNA in RPE 45 even after 7 d exposure. 300 mM glucose increased AR2 mRNA in RPE 45 cells but the steady-state level after 7 d was 30% of that noted at 24 h. RPE 91 had a 17-fold higher basal level of AR2 mRNA than the basal level of RPE 45 cells. No significant change in AR1, vimentin, or β -actin was evident.

this hypothesis derives from studies of the dynamic effects of 7-d exposure to 300 mM glucose. After an acute increase in AR2 mRNA, RPE 45 cells subsequently demonstrated a 30% decrease in AR2 mRNA levels between 1 and 7 d, which was prevented by treatment with the aldose reductase inhibitor, sorbinil (36). The decline in AR2 mRNA in RPE 45 cells may be due to the “feedback” effect of metabolites of the enzymatic reaction catalyzed by AR2 (e.g., sorbitol accumulation), as has been proposed for renal medulla (37). Despite the presence of markedly elevated sorbitol levels in RPE 91 cells (nearly three-fold greater than RPE 45), AR2 mRNA levels were unchanged after 7 d exposure to 300 mM glucose. These data would suggest that AR2 gene expression in RPE 91 cells is unresponsive to both hyperosmotic stimuli and to putative feedback inhibition by polyol pathway intermediates, consistent with the hypothesis that the AR2 gene is essentially unregulated in RPE 91 cells.

The regulation of the AR2 gene expression has been studied best in the renal medulla, where sorbitol is one of several alternative non-ionic organic intracellular osmolytes produced in response to the physiological extracellular hyperosmolarity associated with antidiuresis (7). This “nonperturbing” class of intracellular organic osmolytes, which includes sorbitol, MI, betaine, and glycerophosphorylcholine (7, 11, 32, 33), maintains intracellular osmolarity without disrupting the intracellular ionic environment (7). Extracellular hyperosmolarity has been shown to increase AR2 gene transcription and AR2 mRNA stability, leading to an increased steady-state level of AR2 mRNA (38) and subsequently AR2 protein (34). Similar mechanisms likely underlie the expression of relevant transporters and/or enzymes for the alternative nonperturbing osmolytes (11, 12, 35) in renal medullary tubular cells. Osmotic induction of AR2 and MI transport has also been described in

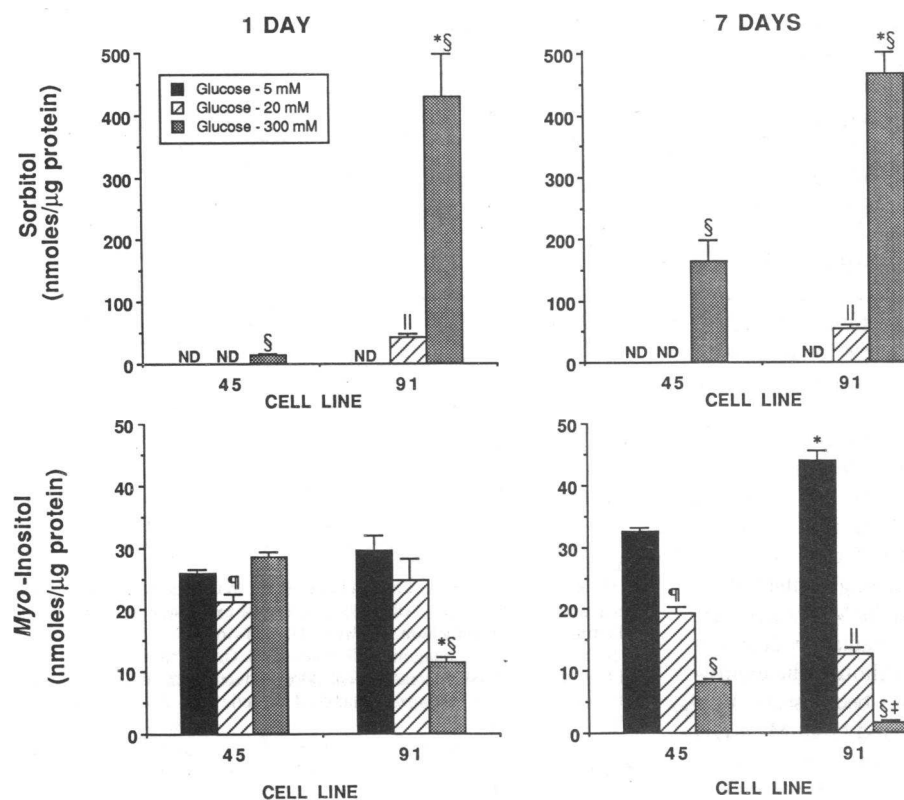


Figure 7. Metabolic consequences of high constitutive AR2 expression. Levels of sorbitol and myo-inositol (nmol/mg protein) were determined in RPE cell lines 45 (regulated AR2 expression) and 91 (constitutive high level AR2 expression) after 1 and 7 d exposure to 5, 20, or 300 mM glucose. RPE 91 demonstrated accelerated sorbitol accumulation and a profound depletion of MI at both hyperglycemic (20 mM glucose) and hyperosmolar (300 mM) glucose conditions. RPE 45 had sorbitol detected only in 300 mM glucose and MI depletion only after 7 d exposure to hyperglycemic or hyperosmolar conditions. *Significantly different from respective condition in RPE 45 ($P < 0.001$). †Significantly different from respective condition in RPE 45 ($P < 0.02$). §Significantly different from 5 and 20 mM glucose ($P < 0.001$). ‡Significantly different from 5 and 300 mM glucose ($P < 0.001$). §Significantly different from 5 and 300 mM glucose ($P < 0.05$).

extramedullary renal cells and in nonrenal cells rarely or never exposed to significant physiologic hyperosmolarity in vivo (31, 39). Thus, the AR2 gene is likely expressed at basal levels in most mammalian cells unless induced by degrees of hyperosmolarity only rarely encountered outside the renal medulla. It would follow that physiologically significant polyol pathway activity in nonrenal tissues is for practical purposes restricted to pathologically elevated glucose concentrations of diabetes that stimulate AR2 activity through mass action rather than gene induction.

Constitutive high AR2 gene expression in RPE 91 cells in 20 mM glucose (20), was associated with exaggerated sorbitol accumulation and MI depletion. Indeed, RPE 91 cells exhibit greater sorbitol accumulation and MI depletion than RPE 45 cells even after 7 d in 300 mM glucose despite comparable AR2 mRNA levels. This presumably reflects the delayed accumulation of the AR2 protein in the RPE 45 cells, which may have not yet reached steady-state levels even after 7 d due to the prolonged metabolic half-life of the AR2 protein (40). If constitutive overexpression of the AR2 gene occurs in vivo, it would probably exaggerate those metabolic consequences of hyperglycemia mediated by the sorbitol pathway, such as MI depletion. MI depletion by substrate-induced (rather than osmotically induced) activation of the polyol pathway in diabetes may itself represent a compensatory shift of alternative intracellular organic osmolytes in an isoosmotic environment (32). Since substrate-induced, iso-osmotic sorbitol accumulation would be unaccompanied by osmotic induction of MI accumulation, MI levels could reach levels sufficiently low as to limit phosphatidylinositol synthesis (20). This degree of MI depletion has been produced in RPE 91 cells with high constitutive AR2 gene expression by brief exposure to 20 mM glucose, and such alterations in phosphoinositide metabolism have been invoked in the pathogenesis of some diabetic complications (41).

Pathways of glucose metabolism whose flux varies with prevailing hyperglycemia are often invoked as potential mediators of the chronic diabetic complications whose cumulative duration and severity parallel that of hyperglycemia (2, 42). Yet these metabolic pathways, such as nonenzymatic glycation and the polyol pathway, are rarely invoked to explain the wide variation of diabetic complications among patients with hyperglycemia of similar degree and duration (1). Fragmentary but nevertheless provocative data now link the tissue-specific catalytic capacity or protein content of aldose reductase-like enzymes with the prevalence and/or severity of diabetic complications (13, 15). The availability of cDNA sequences for RPE AR1 and AR2, and the delineation of the structure of the human AR2 gene (43) provide highly specific tools to independently study normal and abnormal regulation of these genes.

The mechanism underlying constitutive high level AR2 gene expression in RPE 91 cells is unknown, but defects in transcriptional regulation and/or mRNA stability represent likely candidates. This defect may represent only one of a broad range of osmoregulatory defects that might affect other "hypertonicity stress proteins" (11) such as the MI transporter. The possibility of a more generalized osmoregulatory defect in RPE 91 cells has not yet been excluded. Although the expression of the MI transporter gene has not yet been assessed directly in RPE 91 cells, the absence of elevated basal MI levels (20) and the rapid fall in MI induced by exposure to hyperglycemic or hyperosmolar concentrations of glucose certainly

would argue against its constitutive overexpression, and would suggest a more limited defect, perhaps confined to the AR2 gene. It is possible that elucidation of this defect(s) might provide a marker for enhanced sensitivity to hyperglycemia, which, if expressed in vivo, might constitute a significant risk factor for diabetic complications. The ready availability of RPE cell lines isolated from diabetic donors by new and efficient cell culture technology (44) should provide an important new system in which to explore these possibilities and potentially to identify genetic markers that might sensitize subgroups of diabetic patients to the effects of hyperglycemia.

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