

Acquisition and Synthesis of Folates by Obligate Intracellular Bacteria of the Genus *Chlamydia*

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Abstract

We undertook studies focused on folate acquisition by *Chlamydia trachomatis* L2, *Chlamydia psittaci* 6BC, and *C. psittaci* francis. Results from in situ studies, using wild-type host cells, confirmed that *C. trachomatis* L2 and *C. psittaci* 6BC are sensitive to sulfonamides whereas *C. psittaci* francis is resistant. In addition *C. trachomatis* L2 and *C. psittaci* francis were inhibited by methotrexate in situ whereas *C. psittaci* 6BC was not. In contrast to *C. trachomatis*, neither *C. psittaci* strain was affected by trimethoprim. Surprisingly our results indicate that all three strains are capable of efficient growth in folate-depleted host cells. When growing in folate-depleted cells *C. psittaci* francis becomes sensitive to sulfonamide. The ability of all three strains to carry out de novo folate synthesis was demonstrated by following the incorporation of exogenous [^3H]pABA into intracellular folates and by detecting dihydropteroate synthase activity in reticulate body crude extract. Dihydrofolate reductase activity was also detected in reticulate body extract. In aggregate the results indicate that *C. trachomatis* L2, *C. psittaci* francis, and *C. psittaci* 6BC can all synthesize folates de novo, however, strains differ in their ability to transport preformed folates directly from the host cell. (*J. Clin. Invest.* 1992;90:1803–1811.) Key words: parasite • dihydropteroate synthase • dihydrofolate reductase • sulfonamide • methotrexate

Introduction

Chlamydiae are obligate intracellular bacterial parasites that infect a wide range of host cells and are the causative agents of a variety of human, nonhuman mammal, and avian diseases (1–5). Chlamydiae display a unique developmental cycle consisting of an infectious extracellular metabolically inert elementary body (EB)¹ and a noninfectious intracellular metabolically active reticulate body (RB). The function of the EB is to survive transit in the extracellular environment until a host is

encountered. Once inside a host cell EBs differentiate to RBs, which divide by binary fission within the confines of a membrane bound cytoplasmic vacuole. Chlamydiae have an absolute nutritional dependency on the host cell to provide a wide variety of intermediates of metabolism. After multiple rounds of division RBs differentiate back to EBs, which are subsequently released from the host cell to begin a new infection cycle.

The genus chlamydiae is currently divided into three species, *Chlamydia trachomatis*, *Chlamydia psittaci*, and *Chlamydia pneumoniae* (5–7). Classically, the species have been differentiated by inclusion morphology (diffuse vs. compact), presence or absence of glycogen within the inclusion (as determined by iodine staining), and differing sensitivity to sulfa drugs. *C. trachomatis* is sensitive to sulfonamides and they develop diffuse glycogen containing inclusions. In contrast *C. psittaci*, with the exception of strain 6BC, is resistant to sulfa drugs and they give rise to dense inclusions that lack glycogen. *C. pneumoniae* is also resistant to sulfonamides and they yield dense inclusions that do not contain glycogen.

Sulfonamides are structural analogues and competitive antagonists of para-aminobenzoic acid (pABA), and thus prevent normal bacterial use of pABA for the de novo synthesis of folic acid (8). More specifically, sulfonamides are competitive inhibitors of the bacterial enzyme dihydropteroate synthase (DHPS), which catalyzes the incorporation of pABA into dihydropteroic acid, the immediate precursor of folic acid. As such, microorganisms that are sensitive to sulfonamides must synthesize their own folates and those that can use preformed folates are resistant to sulfa drugs. Mammalian cells are not affected by sulfonamides because they must obtain preformed folates from dietary sources.

The biologically active form of folate is tetrahydrofolate (H_4folate), which functions as a one-carbon unit carrier in a variety of biosynthetic reactions, including methionine biosynthesis, thymidylate synthesis, and purine biosynthesis (9, 10). Thymidylate synthesis is unique among the biosynthetic reactions that employ H_4folate as cofactor in that it involves not only the transfer of a one-carbon moiety but also the oxidation of the carrier (11). The dihydrofolate (H_2folate) formed is converted back to H_4folate by the enzyme dihydrofolate reductase (DHFR). H_4Folate is again converted to a cofactor by the addition of a one-carbon unit as catalyzed by serine hydroxymethyltransferase. Together these reactions form the thymidylate cycle as represented schematically in Fig. 1.

Both *C. psittaci* and *C. trachomatis* have been shown to contain folates different from those present in their host cells (12, 13). In addition several studies have shown that chlamydiae cannot use medium-supplied thymidine (14–17), however, they can incorporate exogenously supplied uridine into parasite DNA (15–17). Taken together these results imply that chlamydiae must contain a thymidylate synthase. Recently we have shown that *C. trachomatis* does encode a thymidylate

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Received for publication 6 February 1992 and in revised form 21 May 1992.

1. Abbreviations used in this paper: CHO, Chinese hamster ovary; DHFR, dihydrofolate reductase; DHPS, dihydropteroate synthase; EB, elementary body; H_2folate , dihydrofolate; H_4folate , tetrahydrofolate; ID_{50} , antimetabolite concentration required to reduce incorporation of radiolabel into DNA by 50%; pABA, para-aminobenzoic acid; RB, reticulate body.

ately after infection with chlamydiae the cell monolayer was rinsed with Hanks' buffered saline, then 15 ml of DME H-21 medium supplemented with 10% dialyzed fetal bovine serum, 0.3 mM proline, 0.3 mM glycine, 30 μ M hypoxanthine, 1 μ g/ml cycloheximide, and 30 μ Ci [3 H]pABA was added to each flask. The cultures, both mock- and chlamydiae-infected, were incubated at 37°C for 24 h and then the cells were harvested and intracellular folates were extracted as previously described (23). Briefly, the monolayers were washed five times with ice-cold PBS then the cells were harvested in 1 ml of PBS by scraping the surface of the flask with a rubber policeman. The cells were heated at 100°C for 1 min in 3% sodium ascorbate, pH 6.0, and 3% 2-mercaptoethanol and then the cell debris were removed by centrifugation. The cell supernatant was treated with 0.5 ml of partially purified hog kidney polyglutamate hydrolase, prepared according to the method of McMartin et al. (24), at 37°C for 30 min to convert all folates to monoglutamates. After an additional boiling with ascorbate and 2-mercaptoethanol, the folates were extracted into methanol using a C-18 cartridge (Sep-Pak; Waters Chromatography Division, Milford, MA) and concentrated under a steady stream of nitrogen. The dried sample was dissolved in 100 μ l of 5 mM PIC A (Waters Chromatography Division) and the individual folates were resolved by HPLC using a C-8 μ Bondapak column (12.5 cm; Whatman International, Clifton, NJ) under isocratic conditions; the mobile phase consisted of 22.5% methanol and 77.5% 5 mM PIC A, pH 5.5. Isotope incorporation into individual folates was determined by in-line radioactive flow detection (171 detector; Beckman Instruments, Fullerton, CA). The identity of the radioactive peaks was confirmed by simultaneously monitoring the A_{290} (1066 UV detector; Beckman Instruments) of known unlabeled folate standards that were coinjected with each sample. Data were collected and processed with an IBM PC 50 using Beckman System Gold software.

Assay of DHPS activity in vitro. DHPS was assayed by previously described procedures (25) with the following modifications. The DHPS assay mix contained, in a final volume of 100 μ l, 100 mM Tris-HCl (pH 8.5), 5 mM NaF, 10 mM MgCl₂, 10 μ M H₂PtCH₂OPP, 1 μ M [3 H]pABA (10 μ Ci/ml), and 5 mM dithiothreitol. The reaction was initiated by the addition of 150 μ g RB extract protein as a source of enzyme and then was allowed to proceed at 37°C for 60 min. The reaction was terminated by the addition of 100 μ l of 3% ascorbate/3% 2-mercaptoethanol followed by boiling for 1 min. The resulting precipitate was removed by centrifugation (14,000 g for 10 min) and then 50 μ l of the supernatant was spotted onto 3 \times 30-cm strips of 3MM chromatography paper (Whatman International). The strips were developed in a descending chromatography tank using a mobile phase buffer of 0.1 M KH₂PO₄, pH 7.0. Once the buffer front had traveled 20 cm, the paper strip was removed from the chromatography tank, the origin containing the labeled product was cut from the strip, dried, and placed in a scintillation vial containing 10 ml cocktail (Universol; ICN Biomedicals, Inc., Costa Mesa, CA). The vial was left at room temperature for 16 h and then it was counted in a liquid scintillation counter (LS 5000; Beckman Instruments).

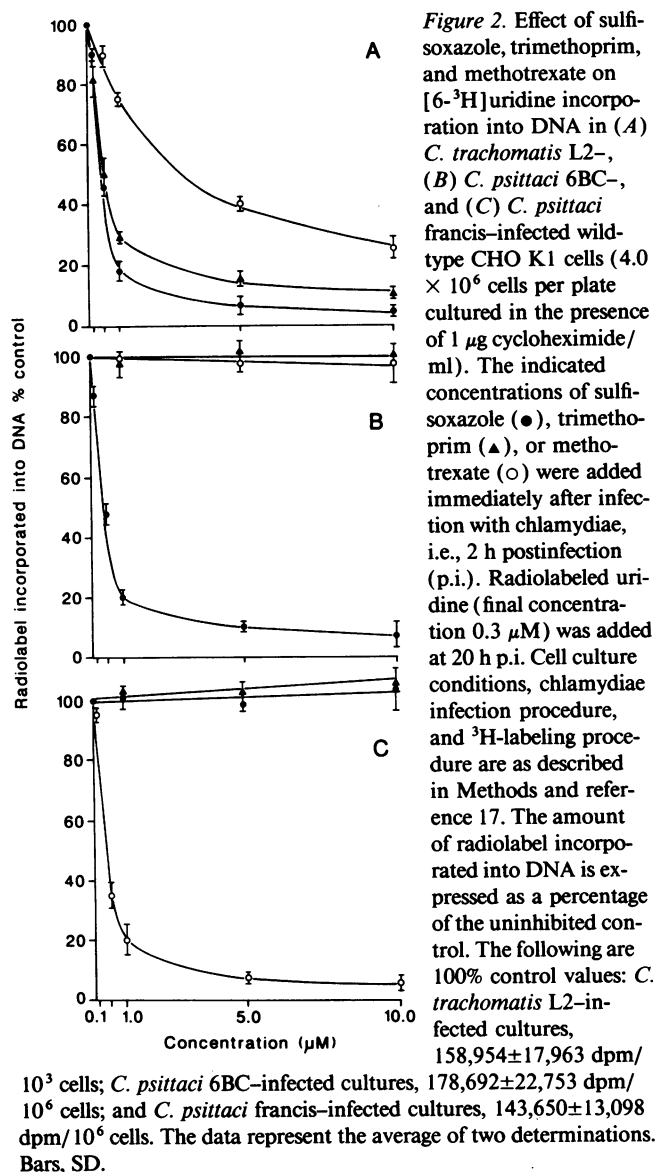
Assay of DHFR activity in vitro. DHFR assays were carried out essentially as described by Baccanari et al. (26). The complete reaction mixture contained, in a total volume of 100 μ l, 50 mM Tris-HCl (pH 7.5), 1 mM dithiothreitol, 200 μ M NADPH₂, 100 μ M [3 H]H₂folate (10 μ Ci/ml) or [3 H]folate (10 μ Ci/ml), and 150 μ g of RB extract protein as a source of enzyme. The reaction was allowed to proceed at 30°C for 10 min and then the reaction was terminated by the addition of 100 μ l of 3% ascorbate/3% 2-mercaptoethanol followed by boiling for 1 min. Precipitated protein was removed by centrifugation and the radiolabeled folic acid, H₂folate, and H₄folate present in the supernatant were resolved by HPLC using a C-18 μ Bondapak column (12.5 cm; Whatman International) under isocratic conditions; the mobile phase consisted of 5 mM PIC A, 10 mM (NH₄) H₂PO₄ (pH 7.3), 20% methanol, and 5% acetonitrile. The identity of the radioactive folate peaks was confirmed by simultaneously monitoring the A_{290} of known folate, H₂folate, and H₄folate standards coinjected with each sample. Data were collected and analyzed as described above.

Results

Effect of various inhibitors of folate metabolism on chlamydiae growth. Initially we wanted to determine the effects of various inhibitors of folate metabolism on the growth of *C. trachomatis* and *C. psittaci*. Chlamydial growth was monitored by measuring the incorporation of [3 H]uridine into DNA in the presence of the eucaryotic protein synthesis inhibitor cycloheximide (16). For historical reasons we used the commonly studied *C. trachomatis* strain L2 as well as *C. psittaci* psittacosis strain 6BC and *C. psittaci* meningopneumonitis strain francis (frequently referred to as *C. psittaci* Cal-10). Three drugs that target folate metabolism were tested. Sulfisoxazole, a competitive inhibitor of dihydropteroate synthase, inhibits de novo folate synthesis (8); trimethoprim, an inhibitor of bacterial DHFR that enters cells by simple diffusion (27); and methotrexate, an aminopterin analogue that inhibits both mammalian and bacterial DHFR (28, 29). In vivo methotrexate is only effective against cells that have a transport system(s) for folates (28, 30).

Results of experiments determining the effect of various concentrations of these three inhibitors on chlamydial growth in wild-type CHO K1 cells are shown in Fig. 2. In keeping with earlier findings (12, 18–20), sulfisoxazole was an effective inhibitor of both *C. trachomatis* strain L2 and *C. psittaci* strain 6BC growth. The concentration of sulfisoxazole required to inhibit DNA synthesis by 50% (ID₅₀) was 0.4 μ M for *C. trachomatis* L2 and 0.5 μ M for *C. psittaci* 6BC. Also in agreement with previous reports (5, 31) we found that sulfisoxazole had no effect on *C. psittaci* francis DNA synthesis. Trimethoprim was effective against *C. trachomatis* L2, having an ID₅₀ of 0.5 μ M. In contrast neither of the *C. psittaci* strains were sensitive to trimethoprim. Both *C. trachomatis* L2 and *C. psittaci* francis were inhibited by methotrexate, having ID₅₀ values of 3.2 and 0.3 μ M, respectively. Growth of *C. psittaci* 6BC was unaffected by methotrexate even at concentrations as high as 100 μ M (data not shown). Since the chlamydial ID₅₀ values for methotrexate are much higher than the ID₅₀ values for mammalian cell lines (28, 29) it is difficult to determine whether methotrexate inhibits chlamydiae directly or indirectly via an effect on the host cell line. This is particularly relevant when one considers that methotrexate inhibits de novo purine biosynthesis in mammalian cells (28, 29) and chlamydiae are auxotrophic for purine ribonucleotides (4, 5). To determine whether methotrexate directly affects chlamydiae replication, we used a DHFR-deficient CHO cell line as a host to support parasite growth. As a result of the DHFR deficiency this cell line is unable to regenerate H₄folate from H₂folate and is unaffected by methotrexate (17, 22). Methotrexate was an effective inhibitor of chlamydial growth in this cell line (Fig. 3). The concentration of methotrexate required to inhibit *C. trachomatis* L2 and *C. psittaci* francis DNA synthesis activity by 50% in this cell line was 4.8 and 2.0 μ M, respectively. As was the case with wild-type cells as host, *C. psittaci* 6BC growth was unaffected by methotrexate in the DHFR-deficient cell line (data not shown).

Growth of chlamydiae in host cells depleted of folates and pABA. To evaluate the requirement of chlamydiae for exogenous folates we tested the ability of the parasite to grow in wild-type CHO K1 cells with depleted intracellular folate pools. To achieve maximal folate depletion we grew the CHO K1 cells for 10 passages in folate- and pABA-free medium sup-



plemented with hypoxanthine, proline, glycine, and thymidine. Results presented in Table I indicate that all three chlamydial strains grew as well in CHO K1 cells extensively starved for folates and pABA as they did in host cells that had been previously cultured in complete medium. The observation that *C. trachomatis* L2 and *C. psittaci* 6BC could grow in folate-depleted host cells is in keeping with their sulfa sensitivity and further supports the suggestion that these two strains can synthesize folates de novo. However, given that *C. psittaci* francis was resistant to sulfonamide (a result that suggested that it could obtain preformed folates from the host) we were surprised that it could grow so well in host cells depleted of folates. To help clarify this paradox we checked the sulfonamide sensitivity of *C. psittaci* francis growing in host cells depleted of folates. The results clearly showed that, in contrast to the findings with folate-replete cells, *C. psittaci* francis was highly susceptible to sulfisoxazole inhibition when grown in folate-starved cells (Fig. 4). With *C. psittaci* francis-infected folate-starved cells the ID_{50} for sulfonamide was $1.0 \mu\text{M}$.

Incorporation of [³H]pABA into chlamydial folates. To directly test if chlamydiae could synthesize folates de novo we determined the ability of all three strains to incorporate radiolabeled pABA into their folate pools. For these studies all chlamydial strains were grown in folate-depleted CHO K1 cells in the presence of [³H]pABA. Fig. 5 shows typical elution profiles obtained after HPLC separation of radiolabeled folates extracted from *C. trachomatis* L2-, *C. psittaci* 6BC-, and *C. psittaci* francis-infected folate-starved CHO K1 cells. As expected, CHO K1 cells were unable to use [³H]pABA for the synthesis of folates (data not shown). All three chlamydial strains incorporated [³H]pABA into their folate pools; however, there are obvious differences in the elution profiles obtained for reduced folates when *C. trachomatis* and *C. psittaci* are compared. The major reduced folates produced by *C. trachomatis* L2 were H₄folate and 10-CHO-H₄folate; variable amounts of 5-CH₃-H₄folate or 5,10-CH₂-H₄folate (the two peaks coeluted) were also routinely detected. In contrast, the predominant reduced folate produced by *C. psittaci* strain 6BC was 10-CHO-H₄folate; with variable amounts of H₄folate, 5-CHO-H₄folate, and 5-CH₃-H₄folate also being detected. *C. psittaci* strain francis produced variable amounts of 10-CHO-H₄folate, H₄folate, and 5-CH₃folate and/or 5,10-CH₂-H₄folate. Sulfisoxazole ($10 \mu\text{M}$) was effective in preventing the incorporation of [³H]pABA into folates by all three chlamydial strains.

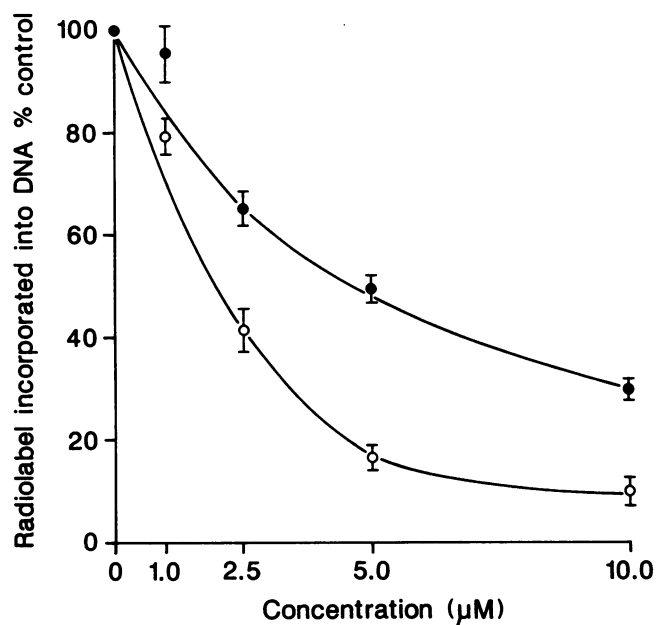


Table 1. Effect of Exogenous Folate on the Growth of Chlamydiae in Chinese Hamster Ovary K1 Cells

Cell line	Culture medium*	DNA synthesis†			
		Mock infected	<i>C. trachomatis</i> L2	<i>C. psittaci</i> 6BC	<i>C. psittaci</i> francis
CHO K1	Folate containing	0.9±0.1	162.6±16.8	197.9±10.7	189.4±9.2
CHO K1	Folate free	0.7±0.2	151.2±12.1	191.0±16.1	197.4±16.8

* Before chlamydial infection, CHO K1 cells were cultured in complete medium containing 2.2 μ M folate or were depleted of intracellular folates by passage in folate- and pABA-free medium. For details, see methods and text. † The effect of exogenous folate on chlamydiae growth was assessed by measuring [6- 3 H]uridine incorporation into DNA at 20 h p.i. Folate-replete or folate-depleted CHO K1 cells were either mock- or chlamydiae-infected confluent monolayers (3.0×10^6 cells per plate cultured in the presence of 1 μ g cycloheximide/ml). For details see Methods and reference 17. Each value represents the mean±SD from two experiments. Results are expressed in 10^3 dpm per 10^6 cells.

Detection of in vitro DHPS activity in chlamydial extracts. To conclusively show that chlamydiae contain DHPS, we prepared extracts from highly purified *C. trachomatis* L2 and *C. psittaci* strains 6BC and francis RBs and then assayed for DHPS activity in vitro. DHPS activity was measured by following the synthesis of dihydropteroate from [3 H]pABA and 6-hydroxymethyl-7,8-dihydropterin pyrophosphate. We consistently detected DHPS activity using RB extract prepared from any one of the three chlamydial strains as a source of enzyme. RB extracts prepared from *C. trachomatis* L2, *C. psittaci* 6BC,

and *C. psittaci* francis catalyzed the synthesis of 3.1 ± 0.5 , 6.5 ± 1.6 , and 2.8 ± 0.3 pmol dihydropteroate product/min per mg protein, respectively. The DHPS activity detected from all strains was inhibited $\geq 90\%$ by 10 μ M sulfisoxazole.

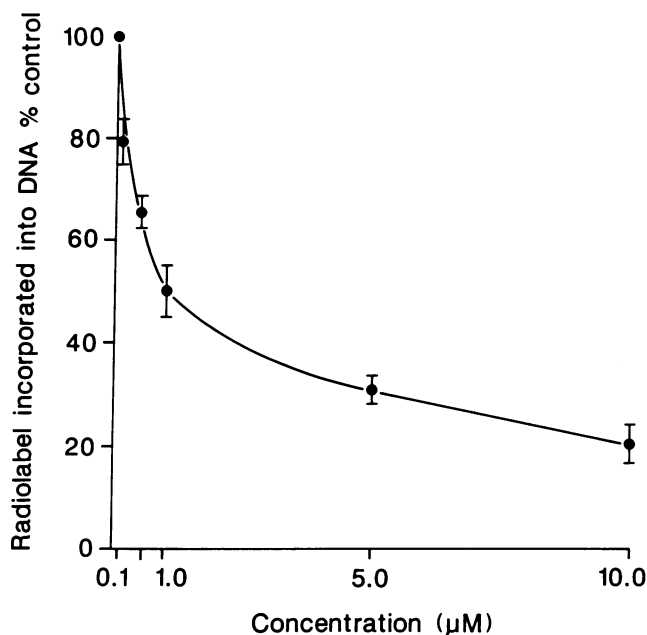


Figure 4. The effect of sulfisoxazole on [6- 3 H]uridine incorporation into DNA in *C. psittaci* francis-infected folate- and pABA-depleted wild-type CHO K1 cells (4×10^6 cells per plate cultured in folate- and pABA-free medium supplemented with proline, glycine, and hypoxanthine in the presence of 1 μ g cycloheximide/ml). The indicated concentrations of sulfisoxazole were added to the culture medium at 2 h p.i. Radiolabeled uridine was added at 20 h p.i. Cell culture conditions, chlamydiae infection procedure, and 3 H-labeling procedure were as described in Methods and reference 17. The amount of radiolabel incorporated into DNA is expressed as a percentage of the uninhibited control. The following is the 100% control value: *C. psittaci* francis-infected cultures, $169,094 \pm 15,386$ dpm/ 10^6 cells. The data represent the average of two determinations. Bars, SD.

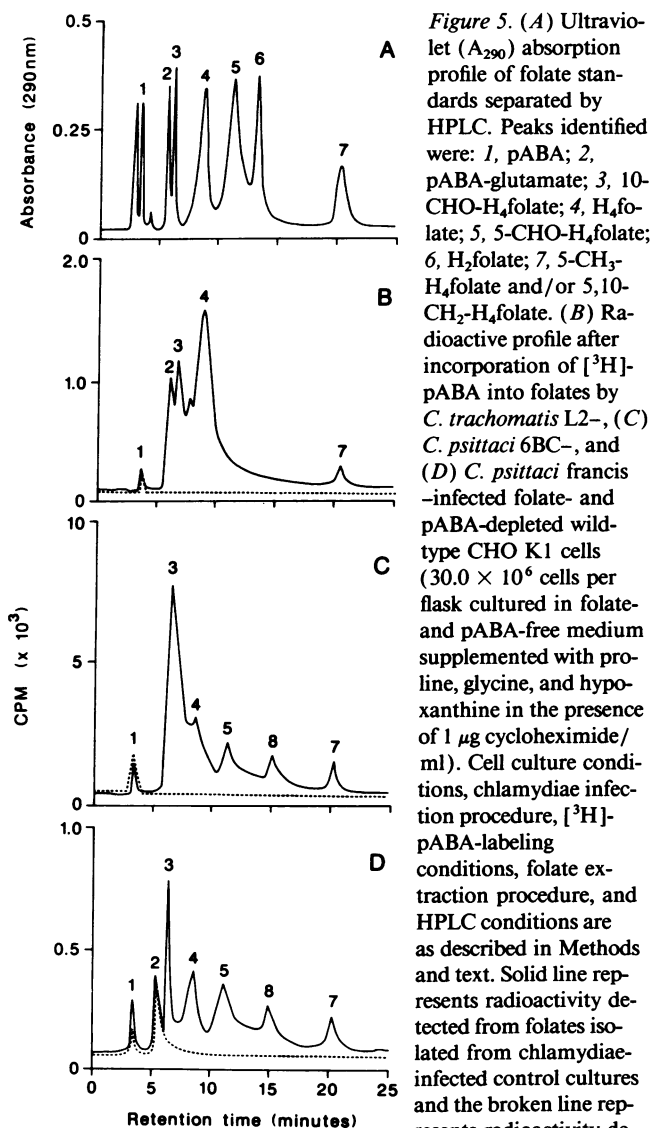


Figure 5. (A) Ultraviolet (A_{290}) absorption profile of folate standards separated by HPLC. Peaks identified were: 1, pABA; 2, pABA-glutamate; 3, 10-CHO- H_4 folate; 4, H_4 folate; 5, 5-CHO- H_4 folate; 6, H_2 folate; 7, 5-CH $_2$ - H_4 folate and/or 5,10-CH $_2$ - H_4 folate. (B) Radioactive profile after incorporation of [3 H]-pABA into folates by *C. trachomatis* L2-, (C) *C. psittaci* 6BC-, and (D) *C. psittaci* francis-infected folate- and pABA-depleted wild-type CHO K1 cells (30.0×10^6 cells per flask cultured in folate- and pABA-free medium supplemented with proline, glycine, and hypoxanthine in the presence of 1 μ g cycloheximide/ml). Cell culture conditions, chlamydiae infection procedure, [3 H]-pABA-labeling conditions, folate extraction procedure, and HPLC conditions are as described in Methods and text. Solid line represents radioactivity detected from folates isolated from chlamydiae-infected control cultures and the broken line represents radioactivity detected from folates extracted from chlamydiae-infected cultures treated with 10 μ M sulfisoxazole. Radioactive peak 8 in the *C. psittaci* chromatograms was not identified.

Reversal of sulfisoxazole inhibition by pABA and folates. It has been shown with numerous experimental systems that the inhibitory action of sulfa drugs can be antagonized by pABA (8). Results presented in Fig. 6 indicate that, with folate- and pABA-depleted CHO K1 cells as host, the growth inhibition caused by 1 μ M sulfisoxazole (Fig. 6 A–C, *hatched bars*) on all three strains of chlamydiae can be completely reversed by 0.1 μ M pABA (Fig. 6 A–C, *cross-hatched bars*). With *C. psittaci* francis, 10 μ M folic acid completely reversed the inhibition caused by 1 μ M sulfisoxazole (Fig. 6 C, *square-checked bar*). Even 1 μ M folic acid was sufficient to reverse 1 μ M sulfisoxazole-induced inhibition (data not shown). In contrast, folic acid was much less effective at reversing the effects of sulfa on *C. trachomatis* L2 and *C. psittaci* 6BC, showing essentially no antagonism at 10 μ M (Fig. 6 A and B, *square-checked bars*) and only partial reversion at 100 μ M (data not shown). We found that the inhibitory effects of 1 μ M sulfisoxazole on *C. trachomatis* L2 and *C. psittaci* francis could be partially and completely reversed, respectively, by 1 μ M 5-CHO-H₄folate (Fig. 6 A and C, *dotted bar*). At a concentration of 10 μ M,

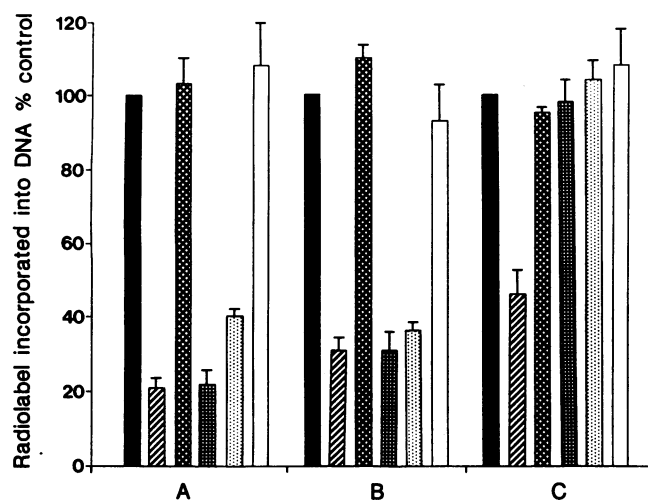


Figure 6. (A) Effect of exogenous pABA and folates on the sulfisoxazole induced inhibition of *C. trachomatis* L2, (B) *C. psittaci* 6BC, and (C) *C. psittaci* francis DNA synthesis. Chlamydiae-infected folate and pABA-depleted wild-type CHO K1 cells (4.0×10^6 cells per plate cultured in folate- and pABA-free medium supplemented with proline, glycine, and hypoxanthine in the presence of 1 μ g cycloheximide/ml) were incubated in the absence or presence of sulfisoxazole, pABA, and/or folates. The indicated components were added at 2 h p.i., and then at 20 h p.i. the cultures were pulsed with radiolabeled uridine. Cell culture conditions, chlamydiae infection procedure, and ³H-labeling conditions are as described in Methods and reference 17. The amount of radiolabel incorporated into DNA is expressed as a percentage of the uninhibited controls. The 100% control values are as follows: *C. trachomatis* L2-infected cultures, $138,034 \pm 15,984$ dpm/ 10^6 cells; *C. psittaci* 6BC-infected cultures, $165,428 \pm 19,683$ dpm/ 10^6 cells, and *C. psittaci* francis-infected cultures, $167,592 \pm 20,849$ dpm/ 10^6 cells. The data represent the average of two determinations. Bars, SD. Chlamydiae-infected control cultures, (*solid bars*); chlamydiae-infected cultures plus 1.0 μ M sulfisoxazole, (*hatched bars*); chlamydiae-infected cultures plus 1.0 μ M sulfisoxazole and 0.1 μ M pABA, (*cross-hatched bars*); chlamydiae-infected cultures plus 1.0 μ M sulfisoxazole and 1.0 μ M 5-CHO-H₄folate, (*dotted bars*); and chlamydiae-infected cultures plus 1.0 μ M sulfisoxazole and 10.0 μ M 5-CHO-H₄folate, (*open bars*).

5-CHO-H₄folate completely reversed the inhibitory effects of 1 μ M sulfisoxazole on *C. trachomatis* L2 (Fig. 6 A, *open bar*). Surprisingly, even though methotrexate did not inhibit the growth of *C. psittaci* strain 6BC (Fig. 2), we found that 10 μ M 5-CHO-H₄folate could reverse the effects of 1 μ M sulfisoxazole (Fig. 6 B, *open bar*).

Our commercial preparation of folic acid was \approx 98% pure, therefore it was possible that a small amount of contaminating pABA may have been present in our folate preparations. Since pABA was \geq 100 times more effective at antagonizing sulfa activity compared with 5-CHO-H₄folate it was possible that the reversion brought about by folates was really caused by contaminating pABA. To eliminate this possibility we tested the ability of folic acid to reverse the inhibitory action of trimethoprim/sulfisoxazole against *C. trachomatis* L2. The results clearly show that 5-CHO-H₄folate can antagonize the combined activity of the DHFR inhibitor trimethoprim and the DHPS inhibitor sulfisoxazole (Table II). As expected folic acid could not reverse trimethoprim inhibition of *C. trachomatis* L2 growth (data not shown).

Detection of in vitro DHFR activity in chlamydial extracts. To directly demonstrate that chlamydiae encode DHFR we conducted in vitro assays for DHFR using extract prepared from highly purified RBs as a source of enzyme (Table III). As a control experiment we conducted DHFR assays with crude extract prepared from logarithmically growing wild-type mouse L cells. We consistently detected DHFR activity in extracts prepared from *C. trachomatis* L2 as well as *C. psittaci* 6BC and francis RBs. The formation of tetrahydrofolate was dependent on the presence of RB extract, NADPH₂, and H₂folate (data not shown). No activity was detected if folic acid was used as substrate. Mock infected mouse cell extract had essentially no DHFR activity.

Similar to in situ results we found that trimethoprim was a highly effective inhibitor of *C. trachomatis* L2 DHFR activity in vitro, however, it was less effective against *C. psittaci* 6BC

Table II. Effect of 5-CHO-H₄folate on Trimethoprim/Sulfisoxazole-induced Inhibition of *C. trachomatis* Growth

Addition to growth medium*			DNA synthesis [‡]	
Sulfisoxazole	Trimethoprim	5-CHO-H ₄ folate	<i>C. trachomatis</i> infected cells	Percent activity [§]
μ M				
—	—	—	145.2 \pm 20.4	100
1.0	1.0	—	19.0 \pm 8.5	13
1.0	1.0	1.0	67.4 \pm 8.6	46
1.0	1.0	10.0	147.4 \pm 12.3	102

* The indicated concentration of sulfisoxazole, trimethoprim, and/or 5-CHO-H₄folate was added to the culture medium immediately after infection (2 h p.i.) with *C. trachomatis* L2. [‡] The effect of the various agents on chlamydiae growth was assessed by measuring [6-³H]-uridine incorporation into DNA at 20 h p.i. Confluent monolayers of CHO K1 cells (3.0×10^6 cells per plate cultured in the presence of 1 μ g cycloheximide/ml) were infected with *C. trachomatis* L2. For details see Methods and reference 17. Each value represents the mean \pm SD from two experiments. Results are expressed in 10^3 dpm per 10^6 cells. [§] The effect of the various agents on the incorporation of radiolabel into DNA is expressed as the percentage of the uninhibited control.

Table III. Dihydrofolate Reductase Activity in Crude Extracts Prepared from Logarithmically Growing Host Cells and Purified Chlamydiae Reticulate Bodies

Substrate*	Inhibitor†	Source of enzyme‡									
		Log growing mouse cells		Mock-infected mouse cells		<i>C. trachomatis</i> L2		<i>C. psittaci</i> 6BC		<i>C. psittaci</i> francis	
		DHFR activity	%	DHFR activity	%	DHFR activity	%	DHFR activity	%	DHFR activity	%
H ₂ folate	—	3.65±0.56	100	<0.01	100	2.34±0.56	100	0.37±0.05	100	1.87±0.39	100
	TMP	4.18±0.71	114	ND [¶]	ND	0.05±0.02	2	0.25±0.04	65	1.53±0.11	82
	MTX	0.45±0.06	12	ND	ND	1.26±0.15	54	0.03±0.02	8	1.03±0.12	55
Folic acid	—	0.26±0.05	7.1	ND	ND	<0.01	<1	<0.01	<1	<0.01	<1

* The complete DHFR reaction mix contained either 100 μ M H₂folate or 100 μ M folic acid as the substrate. For details of assay conditions, see Method. † To assess the effect of trimethoprim (TMP) and methotrexate (MTX) on DHFR activity, a complete reaction mix minus substrate was incubated in the presence of 10 nM TMP or 1 nM MTX for 10 min at 4°C. The reaction was initiated by the addition of H₂folate substrate and incubation was at 30°C. ‡ Crude extracts were prepared from the various sources as described in reference 17. || Reactions were carried out at 30°C for 10 min. DHFR activity is expressed as nmol H₄folate formed/mg protein per min. Each value represents the mean±SD from two experiments. ¶ ND, not determined.

and francis DHFR activity in vitro (Table III). In agreement with previous observations that methotrexate is an effective in vitro inhibitor of DHFR activity from most bacterial and mammalian sources (32), we found that it was active against in vitro DHFR activity of all three chlamydial strains.

Discussion

Although there was no conclusive evidence until recently (17), it has been assumed that chlamydiae require folates for the generation of thymidine nucleotides (5, 15). Furthermore, it has generally been accepted that all *C. trachomatis* strains are sensitive to sulfonamides whereas all *C. psittaci* strains, with the exception of 6BC, are resistant to sulfa action (1, 4, 5, 12, 18–20). A reasonable explanation for these findings was that *C. trachomatis* strains and *C. psittaci* 6BC were capable of synthesizing folates de novo whereas the remainder of the *C. psittaci* strains were not. Unlike the simple interpretation required to explain the action of sulfonamides against chlamydiae it has proven difficult to interpret results obtained using antifolates that target DHFR (5, 19, 33). One must be cautious when comparing various results obtained with DHFR inhibitors because many different host cell systems, i.e., chicken embryo and tissue culture cell lines from different mammalian species, have been employed and it has recently been shown that variations in methodology markedly influence chlamydial antimicrobial susceptibility results (34).

In agreement with earlier in situ observations (12, 18), we found that *C. trachomatis* L2 and *C. psittaci* 6BC were sensitive to sulfonamides. Furthermore, *C. psittaci* francis was resistant to sulfonamide, so long as folates were present in the culture medium. In addition our results indicate that trimethoprim was active against *C. trachomatis* L2 in situ but had no effect against either *C. psittaci* strain. Methotrexate inhibited the growth of *C. trachomatis* L2 and *C. psittaci* francis but did not effect *C. psittaci* 6BC growth. Previously Morgan (19) reported that *C. psittaci* 6BC was sensitive to aminopterin. We have consistently found that the growth of *C. psittaci* 6BC is resistant to a wide variety of antifolates, including aminopterin (data not shown), methotrexate, and trimethoprim. It is sur-

prising that *C. psittaci* 6BC was completely resistant to methotrexate especially since we found that 5-CHO-[H₄]folate could reverse sulfa inhibition of *C. psittaci* 6BC growth. One possible explanation for this finding is that *C. psittaci* 6BC may only be capable of transporting reduced folates. This would also explain why folic acid was much more effective at reversing sulfa action against *C. psittaci* 6BC than was folic acid. It is of interest to note that *Pediococcus cerevisiae* is resistant to aminopterin and methotrexate but it requires 5-CHO-[H₄]folate for growth and has a specific transport system for reduced folates (35).

Since *C. trachomatis* L2 and *C. psittaci* 6BC are sensitive to sulfa action it has long been assumed that they must be capable of de novo folate synthesis. We have confirmed this by showing that: (a) both these strains readily grow in folate-depleted CHO K1 cells, (b) both incorporate exogenous [³H]pABA into folates, and (c) extracts prepared from highly purified RBs of both strains contain DHPS activity. Most interestingly our results clearly indicate that, when growing in folate-depleted CHO K1 cells, the normally sulfonamide-resistant *C. psittaci* francis becomes sensitive to the drug. The ability to incorporate [³H]pABA into folates and the detection of in vitro DHPS activity provides conclusive evidence for the existence of a de novo synthesis pathway in *C. psittaci* francis. As expected, sulfisoxazole prevented the in situ incorporation of [³H]pABA into folates in all three chlamydial strains.

With folate-starved CHO K1 cells as host we found that sulfonamide inhibition of all chlamydial strains could be reversed by the addition of exogenous pABA. 5-CHO-H₄Folate was able to antagonize sulfonamide activity in all three strains, a result that supports the suggestion that all strains have the capacity to transport reduced folates. Interestingly, even though it was less effective on a molar basis than 5-CHO-H₄folate, folic acid could also effectively reverse sulfa inhibition of *C. psittaci* francis growth but was much less effective at antagonizing sulfa activity against *C. trachomatis* L2 or *C. psittaci* 6BC. Although many interpretations are possible, we believe that this result likely reflects the fact that the host cell folate transporter has a lower affinity for folic acid than 5-CHO-[H₄]folate (30) and that *C. psittaci* francis is more efficient at ob-

taining both reduced and nonreduced forms of folates from the host cell than are *C. trachomatis* L2 or *C. psittaci* 6BC. This hypothesis is also supported by the observation that when folates are present in the culture medium *C. psittaci* francis does not depend on de novo folate synthesis (as indicated by sulfa resistance) whereas *C. trachomatis* L2 and *C. psittaci* 6BC do (as indicated by sulfa sensitivity).

We consistently found that there was a difference in the composition of the intracellular folate pools between *C. trachomatis* and *C. psittaci* species. Although reduced folates were predominant in both chlamydial species, the *C. trachomatis* L2 folate pool was dominated by H₄folate whereas the *C. psittaci* 6BC folate pool was dominated by reduced folates carrying a one-carbon unit (i.e., 10-CHO-H₄folate). At the present time the significance of this difference is not known. However, it is interesting that, using the classical microbiological assay with *Lactobacillus casei* and *Pediococcus cerevisiae*, Colon and Moulder (31) also detected a difference in the composition of chlamydial species folate pools.

Our ability to detect DHFR activity in RB extracts from all three chlamydial strains confirm that the parasite does encode a DHFR. Results of in vitro DHFR assays indicate that the enzyme from all three strains is sensitive to methotrexate. In agreement with in situ results, we found that trimethoprim was a good inhibitor of *C. trachomatis* L2 DHFR activity in vitro. Both strains of *C. psittaci* were resistant to trimethoprim in situ and the in vitro DHFR activities of these two strains were less sensitive to trimethoprim than was *C. trachomatis* L2 DHFR activity in vitro. However, the difference in in vitro sensitivity between the species was not as great as might have been expected given the large difference in trimethoprim sensitivity in situ. This raises the possibility that there could be differences in the way *C. trachomatis* and *C. psittaci* metabolize trimethoprim or in their intrinsic permeability to the drug.

It is evident from the results presented that no simple concluding statement can be made with regard to folate metabolism in chlamydiae. The vast majority of free-living bacteria, both pathogenic and nonpathogenic, lack transport system(s) for preformed folates and thus depend on de novo synthesis. Recent studies on a variety of parasitic protozoa have shown that both de novo synthesis and salvage pathways for folates exist in eucaryotic intracellular parasites (36–40). Intracellular parasites spend most of their lives within host cells rich in nutrients. To obtain nondiffusible nutrients from their host, intracellular parasites must evolve (or obtain) suitable transport systems. Once a parasite has acquired the ability to obtain complex nutrients from its host it can afford to lose the capability to synthesize the given nutrient de novo. There would likely be a period of time when both capacities overlap and in some instances it may be necessary for the parasite to retain both pathways.

We believe that folate metabolism in chlamydiae is currently at this stage in evolution. All strains have an absolute dependence on folates for de novo thymidine synthesis. Originally this need was likely fulfilled via de novo folate synthesis as suggested by the ability of all strains tested to incorporate exogenous pABA into folates. More recently chlamydiae has obtained the necessary genetic information to allow them to acquire preformed folates from their host. The current status of the folate transport system(s) appears to vary from strain to strain. At one extreme *C. psittaci* francis fulfills its needs for folate strictly by transporting preformed host folates but does

retain the capacity to synthesize de novo. At the other extreme *C. psittaci* 6BC appears to depend almost exclusively on its ability to synthesize folates de novo, however, it also has the capacity to transport reduced folate to a limited extent. Much of the discrepancy in the literature regarding the effectiveness of antifolates against chlamydiae, both in situ and clinically, probably results from the parasites variable dependence on the two folate acquisition pathways.

Acknowledgments

We thank C. Allegra for providing us with H₂PtCH₂OPP and A. Andersen for performing the monoclonal antibody typing.

This research was supported by grants provided from the Manitoba Health Research Council (G. McClarty) and Medical Research Council of Canada (G. McClarty and R. Brunham). Grant McClarty is a recipient of a Manitoba Health Research Council Scholarship and Huizhou Fan is a recipient of a Manitoba Health Research Council Studentship.

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