

J. A. Nolta, X.-J. Yu, I. Bahner, and D. B. Kohn.
The Journal of Clinical Investigation. Volume 90, No. 2, August 1992.

Page 344.

Due to a printer's error, Figure 2 was printed incorrectly. The correct version appears below.

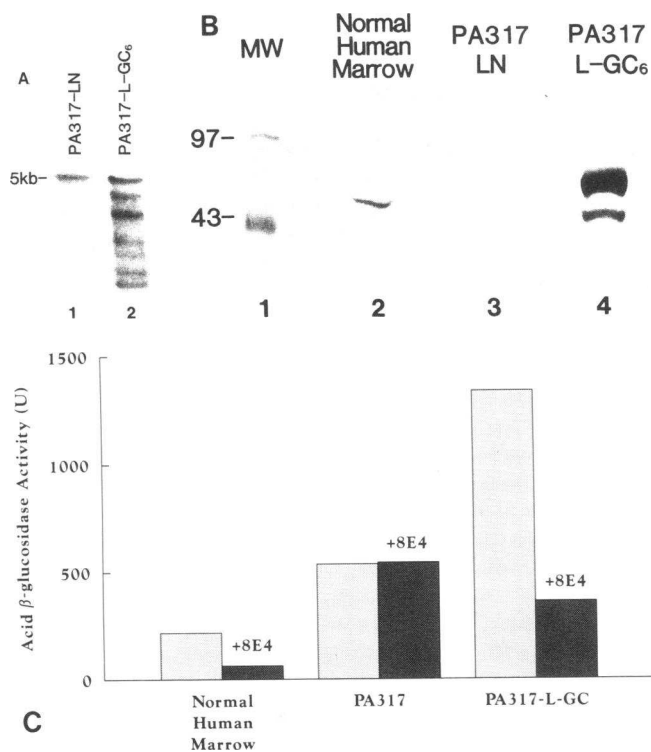


Figure 2. Characterization of an amphotropic PA317 clone producing the L-GC retroviral vector. (A) Quantitation of proviral copy number by Southern blot analysis. Genomic DNA from PA317/LN (lane 1) and PA317/L-GC₆ (lane 2) cells was digested with EcoRI, resolved on a 1% agarose gel, transferred to nylon membrane, and hybridized with the human GC cDNA probe. The band at 5 kb represents the endogenous murine GC gene. The PA317/L-GC₆ clone contains an additional six bands, which represent vector proviral integrants. (B) Production of vector-derived human GC detected by Western blot analysis. 30 μ g of total cellular protein from nonadherent cells harvested from long-term culture was electrophoresed in a 10% SDS-polyacrylamide gel, transferred to nitrocellulose, and incubated with the 8E4 monoclonal antibody, which is specific for human GC. (C) Quantitation of human GC enzymatic activity by selective immunoprecipitation. GC enzymatic activity was measured by incubation with the synthetic fluorogenic substrate 4-methyl-umbelliferyl- β -D-glucopyranoside (4-MU; Sigma Chemical Co.) followed by fluorometry. Units of GC activity are defined as nanomoles 4-MU produced per minute per milligram of protein. GC activity was measured before (solid bars) and after (shaded bars) immunoprecipitation with the monoclonal antibody 8E4 and *Staphylococcus aureus* protein A beads (Calbiochem-Behring Corp., La Jolla, CA).