

Platelet-derived Growth Factor Receptors on Macrovascular Endothelial Cells Mediate Relaxation via Nitric Oxide in Rat Aorta

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Abstract

The effects of platelet-derived growth factor (PDGF) were studied in isolated rings of rat aorta contracted submaximally to phenylephrine. The BB isoform of PDGF elicited relaxation in rings with endothelium and further contraction in rings without endothelium. Both the endothelium-dependent relaxation and endothelium-independent contraction occurred at concentrations known to induce PDGF receptor-mediated responses in cultured cells. Furthermore, the relaxation was isoform specific. This conclusion is supported by the unique ability of PDGF-BB to induce endothelium-dependent relaxations, as well as by studies showing isoform specific, concentration-dependent desensitization of PDGF-BB relaxation. The relaxation induced by PDGF-BB was prevented by $N\omega$ -nitro-L-arginine. It was also observed that endothelium-independent contractions to the AB and AA isoforms of PDGF were less than those to PDGF-BB. Contrary to the widely held view that PDGF receptors are not present on the endothelium of macrovessels, these studies provide evidence for an endothelium-dependent, nitric oxide mediated relaxation of rat aorta caused by PDGF via PDGF $\beta\beta$ -receptors. (*J. Clin. Invest.* 89:878-882.)
Key words: $\beta\beta$ -receptor • endothelium • nitric oxide • platelet-derived growth factor • rat aorta

Introduction

Platelet-derived growth factor (PDGF)¹ is a peptide dimer known to exist in three isoforms (BB, AB, and AA) that exerts its biological effects via three different receptor types ($\beta\beta$, $\alpha\beta$, and $\alpha\alpha$). It is a potent mitogen and has been implicated in the vascular response to injury and the pathogenesis of atherosclerosis and hypertension (1-3). These conditions are associated with cell proliferation and/or migration as well as abnormal vasomotor responses (1-7). PDGF is expressed in the walls of normal arteries under circumstances where cell turnover is low, suggesting that PDGF may have important nonmitogenic functions (8). Based on studies showing that PDGF causes endothelium-independent contractions of vascular strips in vitro,

it is thought that PDGF may have a role in the modulation of vascular tone (9).

Recent studies of cultured microvascular endothelial cells have shown that PDGF receptors are present on these cells and that treatment with PDGF results in phosphorylation of the PDGF receptor, increases in glucose and basic amino acid uptake and stimulation of DNA synthesis (10, 11). However, it is a widely held view that PDGF receptors are not present on the endothelium of large vessels (10-18). Also, it is not known if the interaction of PDGF with endothelial cell receptors is capable of eliciting changes in vascular tone. The purpose of this study was to determine possible endothelium-dependent vasoactive effects of PDGF in macrovessels and to characterize these responses.

Methods

Aortic rings. Descending thoracic aortae were excised from male Wistar rats (276-300 g) killed by exsanguination after anesthesia with ketamine (80 mg/kg i.m.) and xylazine (14 mg/kg i.m.) and anticoagulation with sodium heparin (100 U i.v.). The aortae were cleaned of adherent connective tissue and cut into transverse rings 4 mm in length. The endothelium was removed from some rings by inserting small forceps into the lumen and gently rolling the ring on moistened filter paper (19).

Vasoactive effects of PDGF isoforms. To study possible changes of isometric circumferential force in response to PDGF, rings with and without endothelium were suspended from strain gauges in organ baths containing 7 ml of physiological salt solution (PSS) of the following composition (mM): NaCl 118.3, KCl 4.7, MgSO₄ 1.2, KH₂PO₄ 1.2, CaCl₂ 2.5, NaHCO₃ 25, Na₂-EDTA 0.026, and glucose 11.0. Once mounted, the rings were maintained at 37°C and gassed with 95% O₂/5% CO₂. The rings were stretched stepwise to a resting tension of 5 g, allowed to equilibrate for 60 min and then contracted with the α -adrenergic agonist phenylephrine (PE) to ~ 40% of a maximal contraction to potassium (120 mM) for all studies. Mechanical denudation of the endothelium had no significant effect on either maximal contractions to potassium (120 mM), or the concentration of PE required to achieve 40% of the potassium contraction (data not shown). The effect of each PDGF isoform (4-1,200 pM) on stable PE-induced tone was then determined by increasing the organ chamber concentration of the isoform by cumulative half-log increments after a maximal response was reached to each increment. Following the PDGF concentration response, the rings were washed with three volumes of fresh PSS and allowed to equilibrate for 30 min. The rings were once again contracted with PE and responses to cumulative concentrations of acetylcholine (Ach, 10⁻⁸ to 10⁻⁵ M) were obtained to assess the functional integrity of the endothelium (20). Rings that relaxed more than 75% of PE-induced tone in response to Ach were considered to have an intact endothelium. Rings that relaxed less than 10% of PE-induced tone to Ach were considered endothelium-denuded.

Effect of incubation with PDGF isoforms on PDGF-BB relaxations. Endothelium-dependent relaxations caused by PDGF-BB were studied after incubation of rings mounted in organ baths under 5 g of tension for 3 h at 37°C in PSS containing: (a) no additions, (b) PDGF-BB (4,

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1. Abbreviations used in this paper: Ach, Acetylcholine; NNA, $N\omega$ -nitro-L-arginine; PDGF, platelet-derived growth factor; PE, phenylephrine; PSS, physiological salt solution.

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40, or 400 pM), (c) PDGF-AB (400 pM), or (d) PDGF-AA (400 pM). PSS was replaced once every hour during the incubation period with fresh PSS containing the same additions. After the incubation period, the chambers were washed out with three volumes of fresh PSS and allowed to equilibrate for ~ 20 min in the absence of PDGF before beginning PDGF concentration response curves.

Effect of cyclooxygenase and nitric oxide inhibition on PDGF-BB relaxations. The effect of cyclooxygenase inhibition on the endothelium-dependent relaxation to PDGF-BB was studied by adding indomethacin (1.0 μ M) to the organ chamber 30 min before beginning the PDGF-BB concentration response. To test the ability of an inhibitor of nitric oxide synthesis to prevent relaxation to PDGF-BB, some rings were treated for 30 min with the arginine analog *N* ω -nitro-L-arginine (NNA, 30 μ M). After the 30 min treatment, the PSS containing NNA was replaced with PSS that did not contain NNA. The treated rings were allowed to equilibrate for ~ 20 min before beginning the PDGF concentration response. To test the ability of L-arginine (100 μ M) to reverse the effects of NNA, this amino acid was added to the chamber for the 20-min equilibration period following NNA treatment and was present throughout the PDGF concentration response.

Drugs. Human recombinant PDGF was used throughout (PDGF-BB and PDGF-AA from PeptoTech Inc., Rocky Hill, NJ; PDGF-AB from Upstate Biotechnology, Inc., Lake Placid, NY). Concentrations of PDGF were calculated using a calculated average molecular weight of 25 kD for all three PDGF isoforms. Acetylcholine chloride, L-phenylephrine hydrochloride, indomethacin, and *N* ω -nitro-L-arginine were obtained from Sigma Chemical Co. (St. Louis, MO).

Data analysis. All responses are expressed as a percentage of PE-induced tone. Data are presented as means \pm SEM. For all experiments, *n* equals the number of rats from which rings of aorta were taken. Significant differences between concentration-response curves and responses induced by a single concentration of agonist were determined by the Student *t* test for unpaired observations. Contractile concentration-response curves to PDGF were compared by analysis of variance. Results were considered statistically significant when *P* < 0.05.

Results

Responses of rings with and without endothelium to PDGF. In rings denuded of endothelium, all three isoforms of PDGF caused slight contractions, although contractions caused by PDGF-BB (*n* = 4) were significantly larger than those elicited by either PDGF-AB (*n* = 6) or PDGF-AA (*n* = 5, *P* < 0.01; Fig. 1, *a* and *c*). The maximal degree of contraction caused by PDGF-BB (1,200 pM) was modest, averaging 0.76 ± 0.18 g or $13.6 \pm 2.8\%$ of the maximal tone induced by depolarization with potassium (120 mM, 5.4 ± 0.4 g, *n* = 4). In contrast, in rings that had an intact endothelium, PDGF-BB caused concentration-dependent relaxations (Fig. 1, *b* and *d*). At similar concentrations, PDGF-AB produced mild contractions and PDGF-AA had virtually no effect (Fig. 1 *d*).

Effect of incubation with PDGF isoforms on PDGF-BB relaxations. Cumulative data showing the effects on PDGF-BB relaxation of 3 h incubation without or with each of the three PDGF isoforms are shown in Fig. 2 *a*. Incubation in the absence of PDGF had no effect on PDGF-BB induced relaxations (*n* = 12, Fig. 2 *a*). Incubation with 400 pM PDGF-BB completely blocked the endothelium-dependent relaxations to PDGF-BB (*n* = 8, Fig. 2 *a*). Incubation with either PDGF-AB (400 pM, *n* = 9) or PDGF-AA (400 pM, *n* = 8) had no significant effect on the subsequent PDGF-BB induced relaxations (Fig. 2 *a*). Relaxations to the endothelium-dependent vasodilator, Ach, following the concentration-response to PDGF-BB

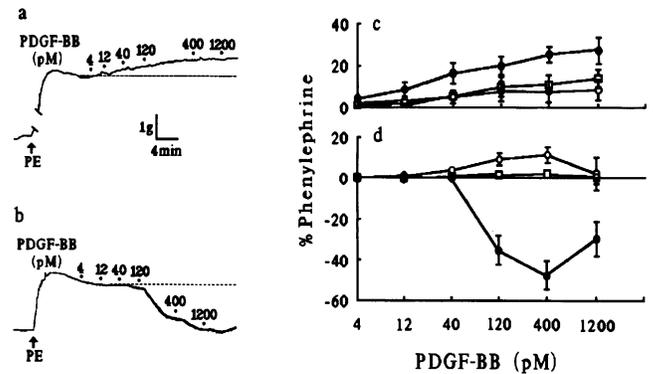


Figure 1. Representative tracings and cumulative data for responses to PDGF in rings of Wistar rat aorta with and without intact endothelium. Tracings show contractions (*a*) and relaxations (*b*) in response to increasing concentrations of PDGF-BB in rings without (*a*) and with intact (*b*) endothelium, respectively. For all studies, aortic rings were contracted with phenylephrine before beginning responses to PDGF. Cumulative data (*c*) for PDGF-BB (\bullet , *n* = 4), PDGF-AB (\circ , *n* = 6) and PDGF-AA (\square , *n* = 5) in endothelium denuded rings showing that PDGF-BB caused significantly greater contractions than either PDGF-AB or PDGF-AA (*P* < 0.01). Cumulative results (*d*) obtained from rings with endothelium showing responses to PDGF-AB (\circ , *n* = 7), PDGF-AA (\square , *n* = 7), and PDGF-BB (\bullet , *n* = 23). Data are expressed as a percent of the phenylephrine contraction. Vertical bars (*c* and *d*) indicate the SEM.

were not affected by any of the incubation conditions (Table I). Incubation with increasing concentrations of PDGF-BB (4 pM, *n* = 5; 40 pM, *n* = 5; 400 pM, *n* = 4) produced concentra-

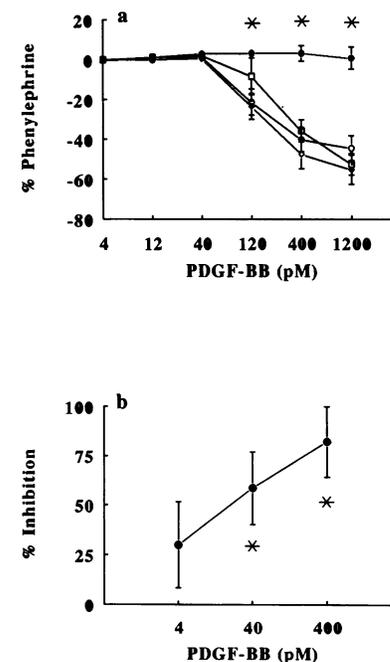


Figure 2. Cumulative data from studies incubating aortic rings having an intact endothelium with each of the three PDGF isoforms. (*a*) Results showing the inhibitory effect of 3 h incubation with PDGF-BB (\bullet , *n* = 8, 400 pM) on subsequent PDGF-BB responses. Neither PDGF-AB (\circ , *n* = 9), nor PDGF-AA (\square , *n* = 8) at a concentration of 400 pM had a significant effect when compared to rings incubated without PDGF (\diamond , *n* = 12). Significant inhibition is shown by asterisks. (*b*) Cumulative data showing concentration-dependent inhibition of relaxations to PDGF-BB following a 3 h incubation at 37°C with PDGF-BB (4 pM, *n* = 5; 40 pM, *n* = 5; 400 pM, *n* = 4). Significant inhibition (*P* < 0.05), shown by asterisks, resulted from incubation with 40 and 400 pM PDGF-BB. Data are expressed as percent inhibition of responses to PDGF-BB 120 pM in rings with endothelium incubated with PDGF-BB as compared to the response of rings simultaneously incubated without PDGF. Vertical bars indicate the SEM.

Table 1. Effect of 3 h Incubation with PDGF Isoforms on Relaxations to Acetylcholine

	Acetylcholine concentration			n
	0.01 μ M	0.1 μ M	1.0 μ M	
Control	-16.83 \pm 4.44	-63.43 \pm 7.94	-85.47 \pm 6.15	11
PDGF-BB	-7.61 \pm 3.46	-48.90 \pm 8.43	-78.57 \pm 7.98	8
PDGF-AB	-23.25 \pm 7.86	-70.47 \pm 10.08	-89.70 \pm 6.14	6
PDGF-AA	-12.40 \pm 3.83	-57.00 \pm 6.13	-86.63 \pm 2.90	8

Values are means \pm SEM. Data are relaxations expressed as a percent of the PE contraction. Following the incubation studies described in Fig. 2 a, rings were contracted with PE and relaxations to cumulative concentrations of acetylcholine recorded. Statistical comparison showed that none of the incubation conditions had a significant effect on relaxations to acetylcholine as compared to the incubated control. n indicates the number of incubated aortae in which acetylcholine responses were studied.

tion-dependent inhibition of relaxations to PDGF-BB (120 pM; Fig. 2 b).

Effect of inhibitors on relaxations to PDGF-BB. PDGF-BB induced relaxations were not significantly affected by indomethacin (1.0 μ M, n = 5; data not shown). However, treatment with NNA (30 μ M), a stereospecific inhibitor of nitric oxide synthase, blocked the endothelium-dependent relaxation to PDGF-BB (n = 13, Fig. 3). Relaxations to sodium nitropruside (1.0 μ M) were not different in rings treated with NNA as compared to control (-100.58 \pm 1.65 vs. -97.17 \pm 1.73, respectively, expressed as a percent of phenylephrine contraction). Following treatment with NNA, PDGF-BB caused contractions that were comparable to those obtained in response to PDGF-BB in rings without endothelium (0.77 \pm 0.24 g). The addition of L-arginine (100 μ M) following NNA treatment, ~ 20 min before the PDGF concentration response, significantly reversed the inhibitory effects of NNA (n = 7, Fig. 3).

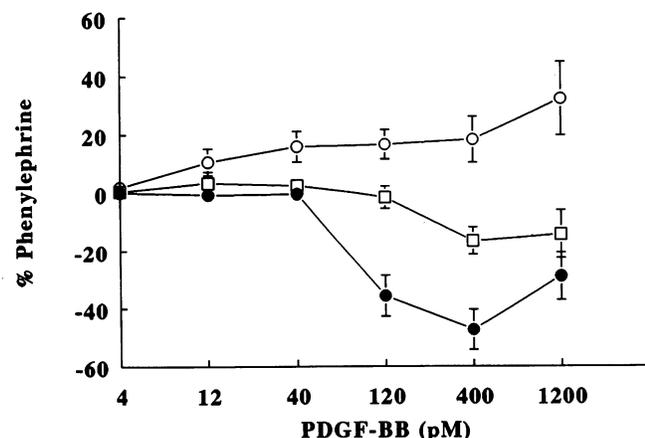


Figure 3. Cumulative data showing that 30 min incubation of aortic rings with endothelium in 30 μ M NNA (o, n = 13) prevented the PDGF-BB induced relaxations observed in control rings (●, n = 23). The inhibitory effect of NNA was significantly reversed by treatment with 100 μ M L-arginine (□, n = 7, P < 0.05). Vertical bars indicate the SEM.

Discussion

Berk et al., in the original report describing a vasoactive effect of PDGF, found that PDGF-induced contractions are unaffected by removal of the endothelium (9). This, in addition to their finding that incubation with PDGF had no effect on Ach-induced relaxation of intact rat aortic strips, led them to the conclusion that PDGF exerts its vasoactive effects in an endothelium-independent manner. This conclusion is in agreement with studies of cultured cells that show macrovascular endothelial cells do not respond to PDGF, presumably because they lack PDGF receptors (10–18). The observation of endothelium-independent responses to PDGF-AB and AA in the present study lends support to this view. However, the demonstration of endothelium-dependent relaxations in response to PDGF-BB suggests that PDGF can act on the endothelium of large vessels, but that such interactions may be isoform specific. The isoform-specific endothelium-mediated effects of PDGF would not have been apparent in the studies by Berk and colleagues as the PDGF preparation used was a mixture of PDGF isoforms (9, 20). Block et al. have also described endothelium-independent contractions to PDGF, but these and other studies were conducted without first inducing active vascular tone (9, 21, 22). These differences from the present study could account for the absence of an effect of the endothelium in earlier reports. A third possibility is that the agonist used to constrict the vessels in the present study stimulated the expression of endothelial cell PDGF receptors, a phenomenon that has been described in cultured vascular smooth muscle cells (23). Even this explanation presumes a capacity for de novo synthesis of PDGF receptors by the aortic endothelium. It is however, still an unsatisfactory explanation of the endothelium-dependent response described, given the short period of time between exposure to the contractile agonist and the PDGF concentration response. A less likely possibility is that in the present study PDGF-BB caused endothelium-dependent relaxations indirectly by first acting on PDGF $\beta\beta$ -receptors in the underlying smooth muscle. Thus, the discovery of an isoform-specific, endothelium-dependent relaxation of rat aortic rings to PDGF-BB is the first evidence for an endothelium-mediated effect of PDGF on vascular smooth muscle.

PDGF-BB initiated endothelium-dependent relaxations at concentrations approximately 10-fold higher than those required for vasoconstriction (Fig. 1). It has been noted by others that PDGF induces contractions of vascular strips at concentrations lower than those required for stimulating either maximal increases in intracellular calcium or growth (9, 21). In the present study, endothelium-dependent relaxation to PDGF-BB occurred in a concentration range that includes the K_d of 140 pM for PDGF receptors and the EC_{50} of 300 pM for half-maximal PDGF receptor autophosphorylation reported by Beitz et al. in cultured microvascular endothelial cells (12). Thus, the concentrations at which relaxations to PDGF-BB occurred are consistent with those known to elicit PDGF receptor-mediated responses in cultured microvascular endothelium.

It is known that PDGF interacts with PDGF receptors in an isoform-specific manner such that $\alpha\alpha$ -receptors bind all three PDGF isoforms, $\alpha\beta$ -receptors bind only PDGF-AB and PDGF-BB, and $\beta\beta$ -receptors bind only PDGF-BB (24). PDGF-BB was the only isoform to cause endothelium-dependent relaxations. This result can be interpreted to indicate that the

relaxations were due to the interaction of PDGF-BB with endothelial cell $\beta\beta$ -receptors.

The isoform-specificity of the endothelium-dependent relaxation to PDGF is further supported by incubation studies conducted with each PDGF isoform. Reports from studies of cultured cells have shown that incubation with PDGF under conditions similar to the ones employed in this study is sufficient to cause isoform-specific downregulation of PDGF receptors and a consequent reduction in PDGF binding (24–26). PDGF-BB was the only isoform to inhibit the endothelium-dependent relaxation to PDGF following incubation, consistent with an isoform specific downregulation of $\beta\beta$ -receptors. Failure of incubation with either PDGF-AB or PDGF-AA to block the endothelium-dependent relaxation is presumably due to the inability of these isoforms to bind at $\beta\beta$ -receptors. The inhibitory effect of incubation with PDGF-BB was concentration-dependent, and even a concentration of PDGF-BB that did not elicit relaxation (40 pM) was able to significantly inhibit subsequent endothelium-dependent relaxation to PDGF-BB. Furthermore, inhibition occurred without affecting endothelium-dependent responses to Ach. These results suggest that the effects of incubation with PDGF-BB were specific for PDGF-induced relaxation and not due to nonspecific desensitization of the endothelium or the underlying smooth muscle. These findings are consistent with inhibition that results from PDGF receptor downregulation. Also, the ready availability of PDGF from platelets and the ability of vascular smooth muscle and endothelial cells to produce and secrete PDGF (2, 21) suggests that the ability of the endothelium to respond to PDGF and exert effects on the smooth muscle may be subject to autocrine or paracrine modulation.

Endothelium-dependent vasodilators act by causing the release of vasoactive prostanoids (27) or the endothelium-derived relaxing factor, nitric oxide (28). The apparent insensitivity of the endothelium-dependent relaxation to the cyclooxygenase inhibitor indomethacin indicates that the relaxation to PDGF-BB was probably not due to endothelium-derived prostanoids. It is now recognized that endothelium-derived nitric oxide, an activator of soluble guanylate cyclase, is synthesized from L-arginine and that certain L-arginine analogs act as competitive inhibitors of nitric oxide synthesis (28–31). In this study, the ability of the arginine analog NNA to completely block the relaxation of PDGF-BB and unmask contractions is comparable to those seen in rings denuded of endothelium, and strongly suggests that the endothelium-dependent relaxation to PDGF-BB is the result of nitric oxide release. The observation that relaxations to sodium nitroprusside were not affected by NNA, coupled with the partial restoration of relaxation to PDGF-BB by supplementation of NNA treated rings with L-arginine, indicate the specificity of NNA for nitric oxide synthase (30). That L-arginine was capable of only partially restoring the endothelium-dependent relaxation is not unexpected, given the potency of this particular arginine analog (31). In separate studies, PDGF-BB also caused endothelium-dependent, NNA-inhibitable, relaxations of porcine coronary artery, but not of rabbit thoracic aorta (data not shown).

This study demonstrates that PDGF-BB causes an isoform specific, endothelium-dependent relaxation of isolated rat aorta. Further evidence showing that arginine-derived nitric oxide mediates this relaxation suggests the presence of previously unrecognized, functional PDGF $\beta\beta$ -receptors on macrovascular endothelium. PDGF stimulation of endothelial cells

may be responsible for the routine replacement and/or repair of damaged or aging vascular endothelial cells. Alternatively, the importance of PDGF- $\beta\beta$ receptor-mediated effects on endothelial cells may rest in their ability to stimulate nitric oxide release. The vascular endothelium may have an important role in regulating the growth of the underlying smooth muscle (32), possibly by releasing nitric oxide (33) that may serve to counteract the direct proliferative effect that PDGF has on the smooth muscle. The loss of this internal control early in vascular diseases featuring abnormal endothelium-dependent responses may help explain the subsequent proliferation of smooth muscle cells that often occurs as a part of the vascular response and which characterizes the pathogenesis of these diseases.

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