 Perspectives

Neutrophil-activating Peptide-1/Interleukin 8, a Novel Cytokine That Activates Neutrophils

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Introduction
Neutrophil accumulation in a tissue is characteristic of inflammation and is observed in a variety of pathological conditions as disparate as infection, trauma, ischemia, and cancer. The process of tissue infiltration is best understood in bacterial infection, where neutrophils are selectively attracted in large numbers to phagocytose and kill the invaders. In other conditions neutrophils are presumably recruited as scavengers of damaged tissue or unwanted extracellular deposits like immune complexes or fibrin. Phagocytosis is accompanied by the release of granule enzymes, superoxide, H2O2, and a variety of bioactive lipids. Several of these products are required for the killing and digestion of microorganisms. They also induce inflammation and tissue damage, however, which is normally observed after neutrophil accumulation.

Several neutrophil chemoattractants have been characterized in recent years; the best known are the anaphylatoxin C5a (1), formylmethionyl peptides of bacterial origin (2), plateletactivating factor (PAF; 3),1 and leukotriene B4 (LTB4; 4). These stimuli have different origins and modes of formation, and their occurrence in disease must thus be expected to vary in accord with the underlying pathophysiological process. C5a is formed upon complement activation via the classical pathway after interaction of microorganisms with antibodies or the formation of immune complexes, or via the alternative pathway after the nonimmune recognition of foreign materials. In bacterial infections, on the other hand, formylmethionyl peptides (which are released by the microorganisms) are likely to be the major attractants. PAF and LTB4 are of special interest because they can be generated by the neutrophils themselves and may thus function as auto- or paracrine amplifiers of the responses elicited by other stimuli (5). It has been shown that C5a, formylmethionyl peptides, PAF, and LTB4 act via unrelated receptors, suggesting that neutrophil recruitment can result from the concerted action of multiple stimuli.

Here we review recent findings demonstrating the existence of a novel tissue-derived neutrophil-activating peptide (NAP) secreted by several types of cells in response to inflammatory stimuli. The new peptide was called MDNCF for monocyte-derived neutrophil chemotactic factor (6), MONAP for monocyte-derived NAP (7), and NAF for neutrophil-activating factor (8) by the groups initially involved in this work. Still other names were used in later reports. The terms NAP-1 and IL-8 were subsequently proposed (9, 10), and NAP-1/IL-8 will thus be used.

Neutrophil activation by chemotactic agonists
Neutrophils exposed to chemotactic stimuli marginate, adhere to the endothelial cells, and migrate into the extravascular space. Several functional responses including shape change, adherance, directed movement, enzyme secretion, and the respiratory burst are observed in vitro after stimulation. All responses are initiated by the binding of the agonist to its receptor and the subsequent interaction of the agonist-receptor complex with a GTP-binding protein (11). This event stimulates phosphatidylinositol hydrolysis by a phospholipase C that yields inositol phosphates and diacylglycerol (12), and leads to a transient rise in cytosolic free calcium (13) and the activation of protein kinase C (14). The entire reaction cascade is considered to be part of the signal transduction process.

Structure of NAP-1/IL-8
Purification and sequencing of the material obtained in the culture supernatants of stimulated human blood mononuclear phagocytes showed that NAP-1/IL-8 has no significant homology with other cytokines produced by these cells, including IL-1, tumor necrosis factor (TNF), colony-stimulating factors, and IFNs (6, 15). NAP-1/IL-8 is generated as a 99-amino acid precursor with a characteristic leader sequence of 22 amino acids (16, 17). Several mature forms have been identified (15, 18, 19), suggesting that NAP-1/IL-8 is processed by repeated amino-terminal cleavage. The major form, which consists of 72 amino acids, has been obtained by recombinant methodology using a synthetic gene expressed in Escherichia coli (15).

NAP-1/IL-8 has considerable sequence homology with peptides from platelet alpha granules such as platelet basic protein (PBP) (20), connective tissue activating peptide III (21), and platelet factor 4 (22). In addition, NAP-1/IL-8 has structural and biological similarities with a recently discovered macrophage inflammatory protein 2 (MIP-2) (23) which could represent the murine homologue to NAP-1/IL-8. As shown in Fig. 1, the relative positions of the four cysteine residues of these peptides are virtually identical. After release from the alpha granules, PBP and connective tissue activating peptide

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1. Abbreviations used in this paper: B-TG, beta thromboglogulin; MIP, macrophage inflammatory protein; NAF, neutrophil-activating factor; NAP, neutrophil-activating peptide; PAF, platelet-activating factor; PBP, platelet basic protein; TNF, tumor necrosis factor.

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III are cleaved to yield beta thromboglobulin (B-TG) (24) and, apparently, even smaller derivatives. A 70-amino acid cleavage product of PBP was recently isolated from cultures of stimulated human blood mononuclear cells and called NAP-2 because of its neutrophil-activating properties which are similar to those of NAP-1/IL-8 (25). Two proteins obtained from human cell lines, gamma-IP-10 (26) and gro (27) or melanoma growth stimulatory factor (28) also belong to this family of peptides on the basis of their structural homology. Their biological activities, however, have not been elucidated in detail. It has been shown that the cysteine residues of NAP-1/IL-8 form disulfide bridges that are essential for activity (29). Like B-TG, NAP-1/IL-8 probably has a two-loop configuration resulting from the linkage of the first to the third and the second to the fourth cysteine (24). In all peptides described, the two first cysteines are separated by one amino acid. This feature distinguishes them from a group of homologues with two adjacent cysteines comprising the murine MIP-1 (30, 31), and recently reported peptides that are chemotactic for human monocytes (32, 33) and thus appear to have biological properties that differ significantly from those of NAP-1/IL-8.

Biological properties of NAP-1/IL-8

Comparative studies have shown that the biological profile of activity of NAP-1/IL-8 is very similar to that of the classical chemotactic peptides C5a and f-Met-Leu-Phe. As summarized in Table I, NAP-1/IL-8 induces the full pattern of responses observed in chemotactically stimulated neutrophils, i.e., activation of the motile apparatus and directional migration, expression of surface adhesion molecules, release of storage enzymes, and production of reactive oxygen metabolites. Real time recordings show that the responses are transient, as with other chemotactic agonists (29, 34). The similarity extends to the mechanism of signal transduction. NAP-1/IL-8 elicits a rapid and transient rise in cytosolic free calcium (29, 34). All NAP-1/IL-8-mediated responses, including the calcium changes, are prevented by pretreatment of the cells with Bordetella pertussis toxin, indicating that signal transduction depends on a GTP-binding protein (34). The respiratory burst induced by NAP-1/IL-8 is inhibited by staurosporine, implying a role for protein kinase C, and by wortmannin (34), a fungal metabolite that blocks certain agonist responses by interfering with a calcium-independent signal transduction pathway (35). Desensitization experiments with C5a, f-Met-Leu-Phe, PAF, and LTβ suggest that NAP-1/IL-8 acts via a selective receptor (7, 29). It was recently reported that neutrophils have on average 20,000 NAP-1/IL-8 receptors with a $K_d$ of 8 x 10$^{-10}$ M (36). Studies by Besemer et al. (37) indicate the existence of ~5,000 high-affinity ($K_d = 1-3 \times 10^{-11}$ M) and 30,000-40,000 low-affinity ($K_d = 6-8 \times 10^{-10}$ M) binding sites per cell. Similar densities but lower affinities were reported previously for f-Met-Leu-Phe (38, 39) and C5a (40, 41) receptors.

The effects of C5a, f-Met-Leu-Phe, PAF, and LTβ are not restricted to neutrophils. Monocytes and other granulocytes respond to these stimuli as well, and PAF has an even wider range of activities. By comparison, NAP-1/IL-8 appears to be more selective: it does not stimulate mononuclear phagocytes or platelets (34), and has only borderline effects on eosinophils (7) and basophils from normal and atopic individuals (42). NAP-1/IL-8 was reported to be chemotactic for lymphocytes (9). Human blood lymphocytes show no cytosolic free calcium changes upon stimulation with NAF/NAP-1 (34). Low levels of specific binding of $^{125}$I-labeled NAF/NAP-1 are found with blood lymphocytes and monocytes separated by elutriation. These cell preparations, however, are not completely pure, and binding by contaminating cells cannot be excluded (37).

NAP-1/IL-8 is not species specific, and its effects in vivo could thus be studied in several laboratory animals. Injection into the skin of rabbits results in plasma exudation and a massive neutrophil infiltration. The effect is long lasting and no other cells beside neutrophils can be detected, underlying the target selectivity of the stimulus (43). Similar, but less severe and more transient lesions are obtained with f-Met-Leu-Phe or E. coli LPS. Massive neutrophil infiltration is also observed upon intradermal injection of NAP-1/IL-8 in rats, mice, guinea pigs, and dogs (Zwahlen, R., L. Colditz, and M. Baggiolini, manuscript in preparation). The long duration of action could be due to the remarkable resistance of NAP-1/IL-8 to inactivation by plasma peptides and its low rate of degradation by proteases (29).

NAP-1/IL-8 is a product of many cells

As several independent laboratories originally described NAP-1/IL-8 as a product of mononuclear phagocytes (6-8, 29, 44), it was interesting to find that the same peptide is expressed in wide variety of cells after appropriate stimulation. Fibroblasts (45), epithelial cells (46), and hepatocytes (46) express NAP-1/IL-8 mRNA in response to IL-1-alpha, IL-1-beta, and

![Figure 1. Sequences of NAP-1/IL-8, the PBP cleavage product beta thromboglobulin (B-TG), platelet factor 4 (PF-4), and MIP-2 aligned according to their four cysteines (*). Hyphens above symbols indicate amino acids that correspond to those of the NAP-1/IL-8 sequence. The largest mature NAP-1/IL-8 form (77 amino acids) is shown. The amino-terminal serine of the main form (72 amino acids) is indicated (>). PBP is 13 residues longer than B-TG. The amino terminus of NAP-2 is indicated above the B-TG sequence. (>).](http://www.jci.org)
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