

Charge of Circulating Immune Complexes as a Factor in Glomerular Basement Membrane Localization in Mice

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ABSTRACT The effect of the charge of circulating immune complexes on glomerular localization was studied in a model of passive serum sickness. Preformed immune complexes of heterogeneous or restricted charge, prepared *in vitro* from isoelectrically focused or chemically modified proteins, were injected intravenously into mice. The distribution of immune complexes in the kidney was compared by immunofluorescence and electron microscopy. Cationic but not anionic or electrophoretically heterogeneous immune complexes gave rise to diffuse localization in the glomerular basement membrane. The binding in subepithelial and subendothelial sites correlated with the known distribution of structural anionic sites. The observations suggest that electrostatic interactions between fixed anionic sites and immune complexes may be an important factor in glomerular trapping. Alternative mechanisms based on initial localization of excess free cationic antigen cannot be completely excluded and are also considered.

INTRODUCTION

Granular deposits of immunoglobulin (Ig) demonstrable by immunofluorescence in the glomeruli of man and animals with glomerulonephritis are generally regarded as evidence of an immunologically mediated disease resulting from the deposition of immune complexes (IC).¹ Deposits are believed to result from two mechanisms: glomerular trapping of circulating soluble IC (1-4), or *in situ* formation of immune deposits by fixation of antibody (Ab) to structural (5-9) or planted

(10, 11) glomerular basement membrane antigens (Ag). Some diseases are characterized by deposits only in subepithelial sites, while in others, subendothelial and/or mesangial deposits predominate. The factors that determine the sites of IC localization are poorly understood. Repeated studies in experimental animals given intravenous injections of preformed soluble IC (passive serum sickness) have shown that IC lodge predominantly in the glomerular mesangium and occasionally in subendothelial sites (12-20). Localization in subepithelial sites of the glomerular basement membrane is exceptional (21). On the other hand, injection of Ab to renal tubular Ag in the model of passive Heymann nephritis has resulted in deposition of Ab only in subepithelial sites immediately after injection (5, 6, 8) and *in vitro* perfusion of the kidney (7). These observations have suggested that subepithelial deposits, such as those found in membranous glomerulonephritis in man and animals, formerly believed to result from the deposition of circulating IC, result instead from *in situ* binding of Ab to Ag.

Previous studies have documented the presence in the glomerular basement membrane of fixed negatively charged sites (22, 23) which facilitate the penetration and binding of positively charged macromolecules while retarding the penetration of negatively charged ones (24, 25). In the present study, the hypothesis that charge-dependent factors might influence the glomerular basement membrane localization of intravenously injected IC on a similar basis was tested in a model of passive serum sickness. The results demonstrate that cationic, but not anionic or heterogeneous IC localize in subepithelial and subendothelial sites.

METHODS

Preparation of antibody. Rabbits were hyperimmunized with bovine gamma globulin (BGG) (Pentex, Fraction II) in complete Freund's adjuvant (Difco Laboratories, Detroit, Mich.).

Isoelectric focusing. Rabbit anti-BGG antiserum was heat inactivated at 56°C for 30 min and an Ig-containing fraction

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¹Abbreviations used in this paper: Ab, antibody; Ag, antigen; BGG, bovine gamma globulin; IC, immune complex; PBS, 0.01 M phosphate-0.15 M NaCl, pH 7.0-7.5; RGG, rabbit gamma globulin.

was prepared by precipitation in 50% saturated $(\text{NH}_4)_2\text{SO}_4$ followed by dialysis against first 0.01 M phosphate-0.15 M NaCl (PBS) and then 0.1% ampholytes, pH 2–11 (Brinkmann Instruments, Inc., Westbury, N. Y.) in H_2O . The Ig was added to an aqueous slurry of Sephadex G-75 superfine made 2.5% (vol/vol) in ampholytes. Preparative isoelectric focusing was carried out in a Desaga/Brinkmann double chamber for thin layer electrophoresis at 400 V for 20 h, then 800 V for 3 h. pH determinations were made at 1 cm intervals; 1 cm strips were mixed with 10 ml of saline, and the protein was harvested by suctioning through scintered glass filters. Fractions were separately dialyzed against normal saline and stored at -20°C . The amount of Ab in each pH range was determined by quantitative precipitation with the particular BGG to be used to prepare the IC (see below).

Differently charged fractions of BGG were prepared by isoelectric focusing in the same manner and were tested for the presence of IgG1 and IgG2 by double immunodiffusion with subclass-specific antisera kindly provided by Dr. John E. Butler of the University of Iowa College of Medicine.

Chemical modification of BGG antigen. Anionization (26) of BGG was done by dissolving 500 mg in 50 ml PBS, pH 7.0. Maleic anhydride (Fisher Scientific Co., Inc., Pittsburgh, Pa.), 1 gm, was added at 0°C ; the pH was maintained at 7.0 with 2 N NaOH for 10 min. The solution was dialyzed against multiple changes of the same buffer, pH 7.5. For cationization (27) of BGG, 200 mg was dissolved in 2.5 ml of 0.01 M NaCl at room temperature. The solution was added to 4.4 ml of 2 M ethylenediamine (Fisher Scientific Co., Inc) and adjusted to pH 7.0 with 2 N HCl. While stirring, 400 mg of 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide-HCl (Pierce Chemical Co., Rockford, Ill.) was dissolved; the pH was maintained at 7.0 with 0.2 N HCl for 2 h. The reaction mixture was kept overnight at room temperature in a stoppered flask, and then dialyzed against several changes of normal saline.

Gel electrophoresis. The relative electrophoretic mobilities of the antigens and antibodies used to prepare IC were compared at pH 8.6 in agarose gels (Fig. 1).

Preparation of immune complexes. The equivalence point of each fraction of rabbit Ab was determined by quantitative precipitation (28) for each preparation of BGG Ag. $E_{280}^{1\%} = 14$ was used to calculate the amounts of Ag and Ab. Since it was not possible to solubilize immune precipitates at equivalence by adding excess Ag and maintain solubility at neutral pH, soluble IC were formed by slowly adding Ab to the calculated amount of Ag (fivefold excess) while constantly stirring at room temperature. The solutions were adjusted to pH 7.2 with HCl or NaOH, left overnight at 4°C , and centrifuged at 12,317 g maximum for 10 min just before injection to remove small amounts of precipitate.

10 types of soluble IC were prepared from the various forms of Ag and Ab (Table I): (1) native electrophoretically heterogeneous Ag (unfocused, unmodified BGG) combined with the $(\text{NH}_4)_2\text{SO}_4$ fraction of native unfocused Ab, (2) chemically modified anionic Ag and focused anionic Ab (Ab isoelectric point (pI) 2.5–5.5), (3) focused anionic Ag and Ab (both Ag and Ab pI 2.5–5.5), (4) chemically modified cationic Ag and focused cationic Ab (Ab pI 8–10), (5) chemically modified cationic Ag and focused neutral Ab (Ab pI 6.5–7.0), (6) chemically modified cationic Ag and unfocused electrophoretically heterogeneous Ab, (7) focused cationic Ag and Ab (both Ag and Ab pI 8–10), (8) electrophoretically heterogeneous Ag and focused cationic Ab (Ab pI 8–10), (9) chemically modified anionic Ag and focused cationic Ab (Ab pI 8–10), (10) chemically modified cationic Ag and focused anionic Ab (Ab pI 2.5–5.5). Noncomplexed proteins given intravenously in equivalent amounts to control mice were (11) normal BGG and normal RGG or (12) chemically modified cationic Ag and

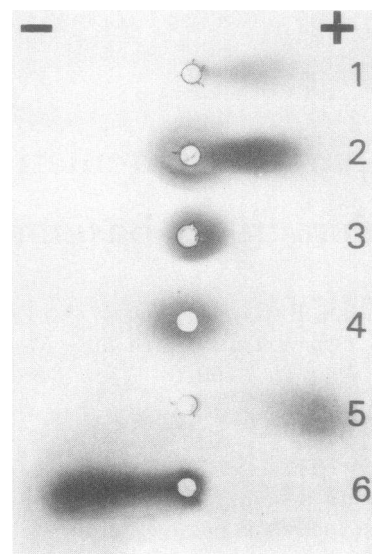


FIGURE 1 Agarose gel electrophoresis in 0.06 M Na barbital, pH 8.6, after Coomassie Blue staining to show the relative mobilities of the antibodies (wells 1–3) and antigens (wells 4–6) used to prepare immune complexes. The rabbit antibodies are from $(\text{NH}_4)_2\text{SO}_4$ precipitates of antiserum against BGG. (1) Anionic fraction prepared by isoelectric focusing; (2) unfocused heterogeneous fraction; (3) cationic fraction prepared by isoelectric focusing; (4) unmodified heterogeneous BGG; (5) chemically modified anionic BGG; and (6) chemically modified cationic BGG. Focused unmodified BGG is less highly charged than chemically modified BGG by the above technique. Also, by isoelectric focusing in 6 M urea, chemically modified anionic, and cationic BGG are about 0.4 pH U more acidic and 1.4 pH U more basic, respectively than the focused anionic and cationic BGG.

normal RGG. Isoelectric focusing could not be used to compare soluble IC directly because the chemically modified BGG were insoluble in the ampholyte solutions.

In the 10 different interacting groups used (see above), the percentage of specific Ab to BGG in each RGG fraction ranged between 6 and 26%. The Ab/Ag ratio at equivalence was 7.1 for group 1, which employed unaltered and unfocused Ab and Ag, and was lower in the other groups that ranged between 2.1 and 5.7. These lower ratios reflect the selection brought about by isoelectric focusing and the loss of antigenic determinants by modification.

Ultracentrifugation of IC. Sizing of IC was performed by centrifugation through a 10–40% linear sucrose density gradient in PBS, pH 7.2, with a 60% sucrose cushion. Soluble IC in 0.2–0.3 ml containing 0.5 mg specific Ab and fivefold excess of Ag (total protein 15–30 mg) were run at 30,000 rpm for 16 h in a Spinco model L5-65 ultracentrifuge in an SW 50.1 rotor. Human IgM and BGG were used as 19S and 7S markers, respectively. Fractions were collected dropwise from the bottom, diluted, and protein content was determined by OD 280 nm and double immunodiffusion. All preparations of IC yielded multiple peaks sedimenting between 9–25S. Selected profiles are shown in Fig. 2.

Injection protocol. Male BALB/c mice (Charles River Breeding Laboratories, Inc., Wilmington, Mass.) 6–8 wk old, weighing 25 g were used. In preliminary studies, a number of different injection protocols were tried with soluble IC com-

TABLE I
Glomerular Localization of RGG and BGG 1 h after Intravenous Injection
of Immune Complexes or Noncomplexed BGG and RGG

Group	Substances injected*	Number of mice	Immunoreactants in glomeruli†		Ultrastructural sites of deposits		
			BGG	RGG	Subepithelial	Subendothelial	Mesangial
Heterogeneous IC							
1	Elect Het Ag/Elect Het Ab	15	2+Mes	2+Mes	—	—	2+
Anionic IC							
2	Chem Mod An Ag/Foc An Ab	6	1+Mes	1+Mes	—	—	1+
3	Foc An Ag/Foc An Ab	2	1+Mes	1+Mes	—	—	1+
Cationic IC‡							
4	Chem Mod Cat Ag/Foc Cat Ab	9	3+GBM	3+GBM	3+	3+	2+
5	Chem Mod Cat Ag/Foc Neut Ab	2	3+GBM	2–3+GBM	1+	2+	2+
6	Chem Mod Cat Ag/Elect Het Ab	2	3+GBM	2–3+GBM	1+	2+	2+
7	Foc Cat Ag/Foc Cat Ab	8	2+Mes	2+Mes	—	—	1+
8	Elect Het Ag/Foc Cat Ab	3	2+Mes	2+Mes	—	—	1+
Mixed IC							
9	Chem Mod An Ag/Foc Cat Ab	2	1+Mes	1+Mes	—	—	1+
10	Chem Mod Cat Ag/Foc An Ab	7	3+GBM	1+Mes	1+	1+	2+
Noncomplexed proteins							
11	Nor BGG/Nor RGG	6	1+Mes	1+Mes	—	—	—
12	Chem Mod Cat Ag/Nor RGG	3	3+GBM	1+Mes	1+	3+	2+

* *Abbreviations used in this table:* (1) Elect Het Ag/Elect Het Ab, electrophoretically heterogeneous antigen (unfocused, unmodified BGG) and the native unfocused electrophoretically heterogeneous antibody; (2) Chem Mod An Ag/Foc An Ab, chemically modified anionic antigen and focused anionic Ab; (3) Foc An Ag/Foc An Ab, focused anionic antigen and antibody; (4) Chem Mod Cat Ag/Foc Cat Ab, chemically modified cationic antigen and focused cationic antibody; (5) Chem Mod Cat Ag/Foc Neut Ab, chemically modified cationic antigen and focused neutral antibody; (6) Chem Mod Cat Ag/Elect Het Ab, chemically modified cationic antigen and unfocused electrophoretically heterogeneous antibody; (7) Foc Cat Ag/Foc Cat Ab, focused cationic antigen and antibody; (8) Elect Het Ag/Foc Cat Ab, electrophoretically heterogeneous antigen and focused cationic antibody; (9) Chem Mod An Ag/Foc Cat Ab, chemically modified anionic antigen and focused cationic antibody; (10) Chem Mod Cat Ag/Foc An Ab, chemically modified cationic antigen and focused anionic antibody; (11) Nor BGG/Nor RGG, normal BGG and normal RGG; (12) Chem Mod Cat Ag/Nor RGG, chemically modified cationic antigen and normal RGG.

† Determined by immunofluorescence; all animals in a given group had the same result. Mes, mesangium; GBM, glomerular basement membrane.

‡ The complexes under this subheading (groups 4–8) all have a net positive charge. Since Ag is in excess and is therefore the major determinant of net charge and because the chemically modified cationic Ag is the most cationic preparation used in this study (more cationic than the focused cationic Ag), complexes 4–6 are more cationic than complexes 7, 8. Group 4 complexes are the most cationic of all.

posed of native heterogeneous Ag and Ab. A comparison by immunofluorescence was made of the amount and distribution of deposits in glomeruli. Variables included the Ag to Ab ratio (between 5- and 20-fold excess Ag), the amount of Ab per injection (between 0.25 and 6 mg), the number of injections (1–3) over a 24-h period, and the time of sacrifice (0 min to 6 h after the last injection). The maximum deposition was generally seen 1 h after injection. Accumulation of IC was always restricted to the mesangium and the amount was not appreciably greater after three than one injection. Also, there were no consistent differences in animals given IC prepared at different equivalence ratios. The regimen eventually chosen was one injection of IC containing 0.5 mg of specific rabbit Ab and a fivefold excess of BGG Ag given over 1–2 min by tail vein in a volume of 0.3–0.5 ml. Mice were sacrificed 1 h later.

Tissue processing. Portions of kidney were fixed in 10%

buffered formalin or Zenker's Fu 48 solution (Technicon Instruments Corp., Tarrytown, N. Y.). Paraffin embedded blocks cut at 2–3 μ m were stained with hematoxylin and eosin or periodic acid-Schiff for light microscopic examination. For immunofluorescence studies, multiple blocks were snap frozen in 2-methylbutane suspended in liquid nitrogen, and stored at –70°C. Cryostat sections (4 μ m) were fixed in acetone or in 10% buffered formalin for 10 s, washed in PBS, incubated in a moist chamber with antiserum, and finally washed in PBS and mounted in glycerol-PBS. Duplicate sections were reacted with the fluorescein-conjugated Ig fraction of rabbit anti-BGG (Cappel N. L. Laboratories, Inc., Cochranville, Pa.), goat anti-rabbit Ig (Calbiochem-Behring Corp., American Hoechst Corp., San Diego, Calif.), and goat anti-mouse C3. (By double immunodiffusion, the fluorescent antisera were found to precipitate specifically and well with all the BGG and rabbit gammaglobulin (RGG) preparations used.) Kidneys

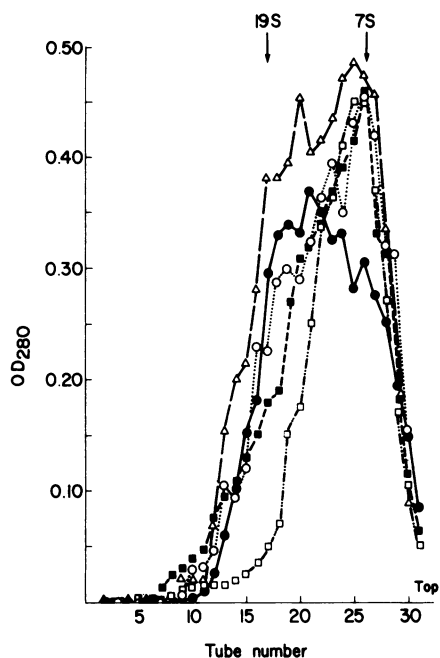


FIGURE 2 Sucrose density gradient ultracentrifugation of five of the immune complexes. To facilitate comparison, the results have been normalized to roughly comparable areas under the immune complex peaks and to 7S IgG appearing at tube 26. The arrows indicate the positions of the 7S and 19S markers. The large amount of 7S sedimenting material is due to some uncomplexed excess BGG antigen and normal (non-anti-BGG) RGG. Symbols for the various IC: ■, group 1; □, group 2; Δ, group 4; ●, group 9; ○, group 10.

from normal control mice incubated with these reagents showed only trace mesangial staining in a few glomeruli or no fluorescence. Some sections were examined by indirect immunofluorescence with sheep anti-mouse C3 (generously donated by Dr. Victor Nussenzweig) followed by fluorescein-conjugated rabbit anti-goat Ig (Calbiochem-Behring Corp.), the latter having been shown to cross-react strongly with sheep Ig. The coded slides were examined in a Leitz Dialux fluorescence microscope (E. Leitz Inc., Rockleigh, N. J.). The intensity of staining in the mesangium and glomerular basement membrane was semiquantitatively graded as -, no or trace staining; 1+, definite but small amounts of staining in some glomeruli; 2+, moderately bright staining in most glomeruli; or 3+, maximal staining. Extraglomerular staining of tubular basement membranes was also noted. For electron microscopy, 0.5 mm³ blocks were fixed by immersion in 2.5% glutaraldehyde in 0.2 M cacodylate buffer, pH 7.4, or in Karnovsky's fixative (29), postfixed in 1% osmium, stained en bloc in uranyl acetate, dehydrated, and embedded in Epon 812 (Shell Chemical Co., Houston, Tex). Thin sections of 2-4 glomeruli from representative groups of mice were double-stained with uranyl acetate and lead citrate and examined and photographed in a Zeiss EM 10A microscope (Carl Zeiss Inc., New York).

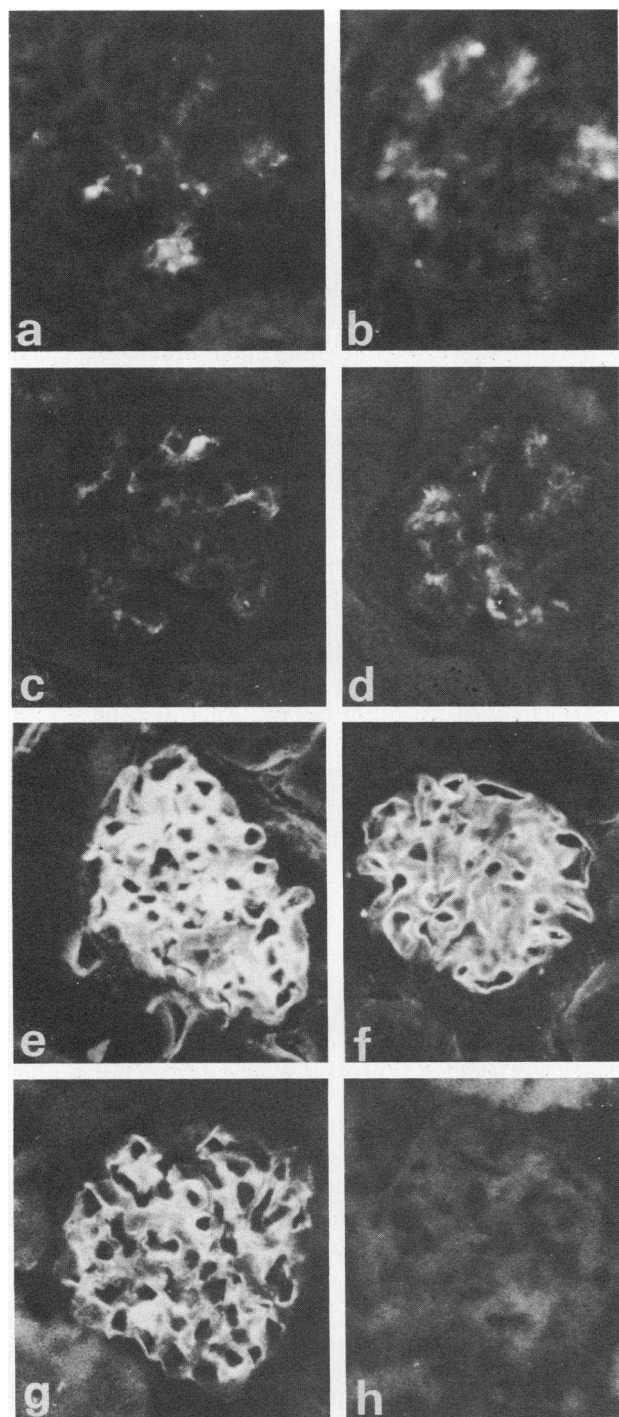
RESULTS

The results are summarized in Table I. By light microscopy, glomeruli appeared normal in all animals.

Controls injected with a mixture of normal BGG and normal RGG in amounts equivalent to the injected IC showed minimal staining for RGG or BGG, only in the mesangium, on immunofluorescence examination (group 11). Those given a mixture of noncomplexed chemically modified cationic Ag and normal RGG (group 12) had BGG but not RGG diffusely bound to glomerular (and tubular) basement membranes indicating that free cationic BGG uncomplexed to Ab could be bound purely on the basis of positive charge.

Striking differences were observed by immunofluorescence in the amount and distribution of IC in the glomeruli of animals given electrophoretically heterogeneous or anionic IC compared to those given cationic IC. Granular deposits of both BGG and RGG were present in the mesangium but not in the peripheral glomerular basement membranes from mice given heterogeneous (group 1) or anionic (groups 2 and 3) IC (Fig. 3a-d). In contrast, in animals given the most cationic IC (groups 4-6), both BGG and RGG were bound in a diffuse, uniform, smooth pattern in all glomeruli along the peripheral basement membranes (Fig. 3e and f). The presence of both Ag and Ab in the same distribution in the glomerular basement membranes indicates that these were present as IC. This was true only when the IC were prepared with the chemically modified cationic Ag and focused cationic or neutral fractions of Ab, or with the unfocused heterogeneous Ab (groups 4-6) but not when IC were prepared with the focused anionic fraction of Ab (Fig. 3g and h; Table I, group 10). In the last case, IC deposition did not occur since only BGG was fixed to the glomerular basement membranes in a manner similar to that seen in controls (group 12) when noncomplexed cationic Ag was injected. The finding of BGG alone in group 10 is explained by the fact that IC were prepared in Ag excess with some free cationic Ag available for binding to the glomerular basement membrane. IC prepared with the focused cationic fractions of Ag and Ab (group 7), with the heterogeneous unfocused Ag and the focused cationic fraction of Ab (group 8), or with the chemically modified anionic Ag and the focused cationic fraction of Ab (group 9) did not localize in the glomerular basement membrane since neither BGG nor RGG was detectable there by immunofluorescence. Mouse third component of complement was not associated with the IC, which can perhaps be attributed to the fact that the IC used were prepared in Ag excess; such complexes are known to fix complement poorly.

It should be emphasized that the IC were prepared in Ag excess. In the limiting case for a bivalent IgG Ab there would be two molecules of Ag per molecule of Ab in the IC. The charge of the Ag is, therefore, the major determinant of the net charge of the IC. The failure of the IC of groups 7-9 to localize in the glomerular base-



BGG

RGG

FIGURE 3 Immunofluorescence with fluorescein-labeled anti-BGG (left) and anti-RGG (right). Glomeruli from mice receiving IC prepared from heterogeneous Ag and Ab, group 1, (a and b) or anionic Ag and Ab, group 2, (c and d) demon-

ment membrane can thus be explained by their being, in aggregate, less cationic than those of groups 4–6, groups 7–9 not having been made with the most cationic Ag preparation, the chemically modified one.

BGG but not RGG localized in the tubular basement membranes only in mice given chemically modified cationic Ag either as ICs or as noncomplexed protein indicating that IC deposition did not occur at this site and that BGG was deposited there on the basis of positive charge alone. The staining was diffuse and generally smooth, but a fine granular pattern was discernible in tangential views (Fig. 4).

Ultrastructurally, there were no deposits in the glomerular basement membranes of mice given electrophoretically heterogeneous (group 1) or anionic (group 2) IC (Fig. 5a and b). In those given IC formed with the chemically modified cationic Ag (groups 4–6 and 10), deposits were present in subendothelial and subepithelial sites. In the latter location, deposits never abutted on the overlying cell membrane. The deposits were most numerous and largest in mice given IC formed with the focused cationic fraction of Ab (Fig. 5c; group 4) as compared to IC formed with neutral (Fig. 5d; group 5) or heterogeneous fractions of Ab, suggesting that the more highly positively charged IC accumulate in greater amounts in the glomerular basement membrane. This differential accumulation was most pronounced (3+ vs. 1+) in the subepithelial location. The noncomplexed cationic Ag was also seen as electron-dense deposits in similar locations, i.e., in subendothelial and subepithelial sites (Fig. 5e; group 12). These deposits ultrastructurally resembled those formed by IC, which could only be distinguished by the immunofluorescence findings demonstrating RGG. In mice given IC prepared with chemically modified cationic Ag and the focused anionic fraction of Ab (group 10), glomerular basement membrane deposits, though present, were smaller and less regular than in those given IC prepared with cationic or neutral Ab (groups 4 and 5). As determined by the absence of RGG on immunofluorescence, group 10 basement membrane deposits were not IC but instead were excess cationic Ag unbound to Ab. The deposits were less numerous than in control mice given un-

strate respectively 2+ and 1+ mesangial deposits of BGG and RGG. Glomeruli from mice injected with cationic IC prepared from chemically modified cationic Ag and the focused cationic Ab, group 4, (e and f) show diffuse 3+ binding of BGG and RGG in peripheral glomerular basement membrane. The deposits appear linear, but at the ultrastructural level are discrete and granular (Fig. 5c). In mice given IC prepared with chemically modified cationic Ag and focused anionic Ab, group 10, (g and h), there is no RGG in glomeruli (h) indicating that IC deposition has not occurred. The localization of BGG (3+) alone (g) is due to deposition of excess uncomplexed cationic Ag. $\times 400$.

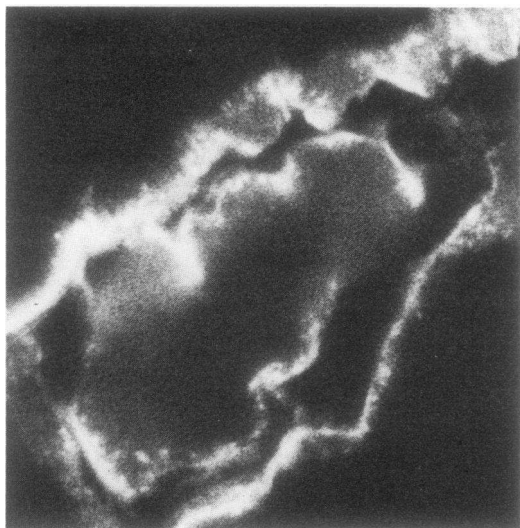


FIGURE 4 Kidney from a mouse receiving chemically modified cationic BGG and normal RGG. Finely granular deposits of BGG are present in tubular basement membranes. There was no staining for RGG (not shown). $\times 2,960$.

complexed cationic Ag (group 12) presumably because there was less uncomplexed Ag available for binding to anionic sites in the glomerular basement membrane. Electron dense deposits in the tubular basement membranes (Fig. 5f; group 12) were due to binding of cationic Ag only as determined by the absence of RGG on immunofluorescence.

DISCUSSION

Our results provide the first evidence that passively administered IC can lead to diffuse and prominent deposition in the glomerular basement membrane on the basis of IC charge. This was demonstrated by the fact that cationic, but not anionic or native heterogeneous IC injected intravenously into mice, were heavily deposited in subendothelial and subepithelial layers.² Differences in the size of the ICs are unlikely to have played a significant role since all preparations of IC sedimented in the same broad range of 9–25S and there were no distinct peaks whose presence or absence correlated with the ability to deposit in glomeruli. Previous attempts in the passive serum sickness model have generally failed to demonstrate convincingly that intravenously injected IC can deposit prominently in subepithelial sites (14–20). Factors such as amount (14, 18, 19), size (20) and affinity (18), or modification of IC by reduction and alkylation (17) have generally had no effect on subepithelial glomeru-

² In this paper, subepithelial layer or space is synonymous with the lamina rara externa of the glomerular basement membrane.

lar basement membrane localization. In mice given low avidity complexes, subepithelial deposits have been described (21) but in these experiments much larger and more numerous doses of IC were used and deposition appeared less extensive than in the present work.

Several lines of evidence suggest that intrinsic properties of the glomerulus could favor the trapping of IC in the glomerular basement membrane. Anionic sites in the subendothelial and subepithelial layers demonstrated by cationic probes (22, 23) and characterized enzymatically and biochemically as sulfated glycosaminoglycans containing heparan sulfate (30, 31) provide a structural basis for the binding of positively charged molecules. Electrostatic interactions are known to occur under physiologic conditions between negatively charged sulfated polysaccharides and positively charged proteins (32). The anionic sites in the glomerular basement membrane have been shown to bind lysozyme (22), cationic ferritin (23, 25), protamine (33), and protamine-heparin complexes formed by perfusion with protamine followed by heparin (34). These considerations suggested that charge interactions between the anionic sites and IC could be a basis for deposition.

To test this hypothesis, we used the model of passive serum sickness. Rabbits were used as a source of Ab because their response after hyperimmunization to foreign protein is largely IgG, and rabbit IgG appears not to contain subclasses. Thus, a differential distribution of subclasses in the various focused fractions should not complicate interpretations. BGG was chosen as the Ag for two reasons. It is readily available and has only two subclasses (35); IgG1 and IgG2 were both present in all fractions used.

Two methods of varying net charge of IC were employed. Isoelectric focusing of Igs, which are normally electrophoretically quite heterogeneous, affords a means of fractionation that does not involve or require modification of the native structure. However, it was not possible with focused Ag and focused Ab to prepare IC that were sufficiently cationic to deposit in the glomerular basement membrane. Consequently, chemical modifications of BGG were used to provide more highly cationic (and also anionic) Ag. The methods used each provide a net alteration of two charges for each side chain modified. Corresponding treatment of the anti-BGG Ab greatly inhibited its ability to precipitate with BGG Ag. Hence, the relationship between charge and localization was best studied with IC made with unmodified focused fractions of Ab (pI, 2.5–5.0; 6.5–7.0; > 8.0) and the chemically modified cationic or anionic Ag. Pronounced differences according to IC charge were observed: the most cationic IC clearly localized in the peripheral glomerular basement membrane while anionic and heterogeneous IC did not.

The ultrastructural distribution of the cationic IC was

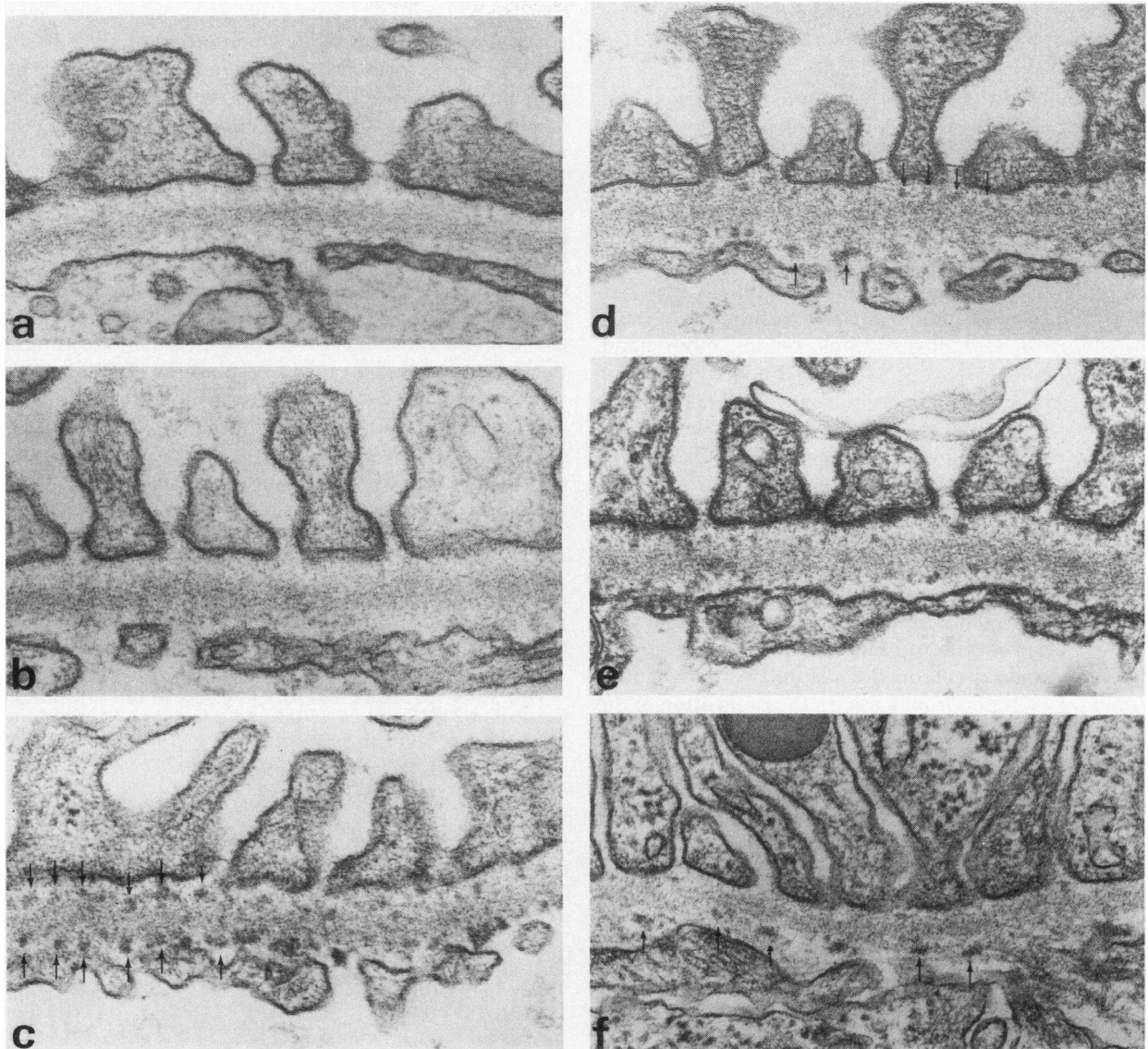


FIGURE 5 Electron micrographs of peripheral glomerular basement membrane of mice given native heterogeneous IC, group 1, (a) and anionic IC prepared from the chemically modified anionic BGG and the focused anionic Ab to BGG, group 2, (b) show no deposits in subendothelial or subepithelial sites. (c) Glomerular basement membrane from a mouse given cationic IC made from chemically modified cationic BGG and focused cationic Ab (group 4) demonstrates deposits in subepithelial and subendothelial sites (arrows). Similar, though smaller deposits (arrows) are present in glomerular basement membrane from an animal injected with IC prepared from chemically modified cationic BGG and focused neutral Ab, group 5, (d); deposits by immunofluorescence were shown to contain both BGG and RGG. Control mouse (e) injected with chemically modified cationic BGG and normal RGG (group 12) also shows deposits, which contained BGG but not RGG by immunofluorescence. Wall of proximal tubule (f) shows deposits (arrows) in tubular basement membrane in a mouse given chemically modified cationic BGG and normal RGG (group 12). $\times 61,000$.

similar to that observed after the administration of cationic BGG alone, or cationic ferritin (23), or in vitro perfusion of kidneys with lysozyme and ruthenium red (22, 23), namely in the layers of the glomerular basement membrane where anionic sites occur. All of

these cationic substances are known to bind to anionic sites in the inner and outer layers of the glomerular basement membrane. These ultrastructural similarities strongly suggest that cationic IC bind to anionic sites in the same manner. In the present study, the most

cationic IC (group 4) localized even more extensively in the subepithelial zones than did free cationic BGG (group 12), 3+ vs. 1+.

Although the significance of charge of the circulating immune reactants in the present experiments seems clear, the sequence by which the deposition occurs is not certain. In the past, the general failure to find deposits in subepithelial spaces after the intravenous injection of IC, though such deposits are readily found after passive administration of Ab to renal tubular Ag in the model of passive Heymann nephritis, has suggested that subepithelial deposits result from *in situ* formation (9). Our observations, on the other hand, support the idea that direct deposition of circulating IC may be a mechanism in some systems. In our experiments Ag and Ab were both deposited in the same sites of the glomerular basement membrane soon after injection of the most cationic IC. Furthermore, when mice were sacrificed immediately after injection of cationic IC, both BGG and RGG were present. Thus, at no time was cationic Ag detectable in the absence of Ab after injection of cationic IC.

However, since the free chemically modified cationic Ag by itself was able to bind to glomerular basement membrane, variations on the planted Ag mechanism cannot be excluded from a role in the deposition of the cationic IC. ICs containing occasional free Ab combining sites could bind to excess free cationic Ag previously deposited alone. If this were so, IC formed from the chemically modified cationic Ag and the anionic focused Ab might have lodged equally well as those formed from the cationic or neutral fractions of focused Ab. This was not the case. Alternatively, free Ab dissociated from circulating IC could cross the glomerular basement membrane and bind to the planted cationic Ag. In this event, free cationic Ab would be more likely to cross the basement membrane than free anionic Ab. This possibility is also considered less likely because the Ab are in Ag excess and are bivalent, which favors multipoint binding. Thus, it is improbable for a whole Ab molecule to diffuse completely away from the IC. It must be emphasized that the various mechanisms just discussed for leading to the deposition of the cationic IC are by no means mutually exclusive.

Under other circumstances, IC containing cationic Ag could form *in situ* in the glomerular basement membrane. Any sufficiently cationic substance, whether of endogenous or exogenous origin, that reaches the circulation could potentially bind through electrostatic interaction, independent of any immune mechanism. If the process were short lived, it would not necessarily be deleterious. However, if the cationic deposit were antigenic, an Ab response could ensue and the resulting IC formed *in situ* could bring about an immunologically mediated inflammatory response.

The binding of cationic BGG alone demonstrates

that Ig as well as other cationic molecules may accumulate in glomeruli on a nonimmunological basis. Such a mechanism may conceivably account for the accumulation of monoclonal Igs in glomeruli and tubular basement membranes in some patients with monoclonal gammopathies in whom evidence for an immune mechanism is lacking (36, 37).

Our studies emphasize that anionic sites in the glomerular basement membranes can interact electrostatically with IC or free Ag under physiologic conditions. Other nonimmunologic mechanisms for binding potential Ag are illustrated by the interaction of the lectin concanavalin A with specific glycoprotein in the glomerular basement membrane (11) and by the interaction of DNA, a polyanion, with collagen of isolated glomerular basement membrane (38). Thus, it is quite likely that a multiplicity of factors can contribute to IC localization in the glomerulus. These remain to be demonstrated in human forms of glomerulonephritis.

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