Heterogeneity of DNA Fragments Associated with the Sickle-Globin Gene

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ABSTRACT We have examined the genetic polymorphism previously reported to be associated with the sickle-cell (β) gene. The polymorphism involves an alteration of the DNA sequence 3' to the β-globin gene as detected with the restriction endonuclease, Hpa I. In normal individuals, the β-globin gene is contained within a DNA fragment of 7.6 kilobases (kb), whereas 87% of individuals with sickle-cell anemia have been reported to have the β°-gene associated with a 13.0-kb Hpa I fragment. We have studied this polymorphism in 31 New York Black individuals homozygous for sickle-cell anemia to ascertain its genetic and biochemical significance and to evaluate its potential use in the prenatal diagnosis of sickle-cell disease. Our results show only a 58% association of the β°-gene and the 13.0-kb Hpa I fragment, as well as the presence of additional variants involving the Hpa I site. In addition, the 13.0-kb fragment is also associated with the β°- and β°°-genes. Thus, the Hpa I polymorphism probably represents a change in DNA not specifically associated with the β°-gene, and appears to antedate the β°- and β°°-mutations.

INTRODUCTION

Until recently, genetic polymorphisms have been studied by analyzing and defining precise structural differences in normal and variant gene products by serological and biochemical methods. Newer techniques, including the use of restriction endonucleases, now permit an analysis of such polymorphisms at the gene level by detecting changes in DNA sequence organization (1). Restriction endonuclease mapping with the Southern “blotting” technique has provided detailed information on the organization of the human δ- and β-globin structural genes (2–5). This procedure involves the cleavage of cellular DNA with restriction enzymes, separation of the DNA fragments by size with agarose gel electrophoresis, transfer of the DNA to nitrocellulose filters, and subsequent hybridization to specific radioactive globin complementary DNA (cDNA)4 probes.

With restriction endonuclease mapping, Kan and Dozy (1) recently reported a polymorphism in the DNA sequence =5 kilobases (kb) from the 3' end of the β-globin gene in individuals with the sickle-cell (β) gene. In normal individuals, Hpa I generates one β-globin fragment, 7.6 kb in size. In association with 87% of β°-genes, there is a change in the sickle DNA sequence at a site which is cleaved by Hpa I that results in a 13.0-kb fragment.

To determine the potential usefulness of this finding in the prenatal diagnosis of sickle-cell anemia and its significance in association with the sickle gene, we have examined this region of the DNA in 70 individuals of African and Caucasian origin. We find that the frequency with which the 13.0-kb fragment is associated with the β°-gene is considerably less than previously reported. Approximately 42% of the time, the β°-gene is present with a normal 7.6-kb Hpa I fragment, which reduces its use as a unique genetic marker for the β°-gene in prenatal diagnosis. In addition, two other Hpa I fragments, 7.0 kb and 5.6 kb, are detected with the β°-gene, which suggests significant heterogeneity of the DNA sequence at the Hpa I site. The 13.0-kb fragment is also found in Black individuals with the β°- and β°°-globin genes, which suggests that it is most likely a result of a change in DNA structure which preceded the β° and β°°-mutations.

METHODS

High molecular weight DNA was isolated from peripheral blood leukocytes and cultured lymphocyte cell lines (6). 20 µg of cellular DNA was digested in a 150-µl reaction mixture

4Abbreviations used in this paper: cDNA, complementary DNA; kb, kilobase(s).
for 3 h at 37°C with 5 U of Hpa I (New England Biolabs, Beverly, Mass.) in a buffer that contained 10 mM Tris-Cl (pH 7.4), 10 mM MgCl₂, 20 mM KCl, 1 mM dithiothreitol, and 100 μg/ml bovine serum albumin. The DNA was reisolated, and fragments were separated on 0.9% vertical agarose gels in a buffer (pH 7.9) consisting of 40 mM Tris-acetate, 20 mM Na acetate, 20 mM EDTA, and 0.5 μg/ml ethidium bromide. Gels were electrophoresed at 25–35 V for 18 h at room temperature with the buffer recirculating. The gels were soaked in 1 M KOH for 0.5 h, neutralized at 1 M Tris-Cl (pH 7.0) for 1 h, and equilibrated with 6 × SSC (1 × SSC = 0.15 M NaCl plus 15 mM trisodium citrate, pH 6.8) for 0.5 h. Separated DNA fragments were then transferred from the agarose gels to nitrocellulose filters (12 × 3 cm) over 24 h. After transfer, the filters were rinsed in 2 × SSC and dried under vacuum at 80°C for 4 h. Pure β-globin cDNA was obtained with plasmid JW102 (kindly supplied by Dr. John Wilson), a PMB9 plasmid containing a double-stranded β-cDNA insert (7). Double-stranded β-cDNA was isolated from PMB9 plasmid DNA after restriction with Hha I and purification of the 1.6-kb fragment by neutral sucrose gradient centrifugation. The cDNA probe was nick-translated, as described, to a 2–4 × 10⁶ cpm/μg sp act (8). The hybridization, washing of filters, and autoradiography were performed as described (4, 5). A Cot value of 0.001 was attained after hybridization at 68°C for 24 h (4, 5).

RESULTS

64 Black individuals were studied including 31 with sickle-cell disease (SS, ββ'), 10 with sickle-cell trait (AS, ββ'), 13 with normal hemoglobin (AA, ββ'), four with SC (ββ'), one with ββ'-Harlem, one with CC (ββ'), one with AC (ββ'), and three ββ'-thalassemia heterozygotes. Six Caucasian DNA were also analyzed. The genotypes of the SS homozygotes were established by measuring the biosynthesis of α- and β-globin in peripheral blood reticulocytes (9). The β/α ratio was ≈1.0 in all individuals with SS studied. 10 of the 31 SS homozygotes showed only the 13.0-kb variant indicating that the Hpa I sites 3' to both β'-genes had been altered (Table I, Fig. 1). 14 individuals had both 13.0-kb and 7.6-kb bands, and five had only the 7.6-kb fragments (Table I). Our data indicate that the frequency of the 13.0-kb variant in association with the sickle-cell gene is ≈58% (Table I). In addition, 13.0/7.0 and 13.0/5.6 sickle-cell variants were detected (Fig. 2).

Several families were studied to confirm the genetic basis of this polymorphism and evaluate its usefulness as an antenatal diagnostic tool. The Hpa I globin gene fragments are inherited in a simple Mendelian fashion in six families studied. We found one family in which both AS parents were 7.6/13.0, and in which the Hpa I polymorphism could be used for a definitive prenatal diagnosis. In another family, both parents with AS had only 7.6-kb fragments, thus preventing a distinction between the β'- and β'-genes. In four of the six families studied, one AS parent was 13.0/7.6, and the other was 7.6/7.6 (Fig. 3).

We also examined DNA from normal individuals, AS heterozygotes, and patients with other hemoglobin variants. In these experiments, we detected the 13.0-kb variant with the β'-gene in three of six opportunities for analysis (Table I). The β'-gene codes for a single amino acid change at position 6 (glu → lys) of the normal βA-globin chain. One individual with only hemoglobin C (CC) had a 13.0- and 7.6-kb fragment, whereas two individuals doubly heterozygous for sickle and C hemoglobin (SC) had only the 13.0-kb variant (Fig. 4). One subject heterozygous for the β'- and β'-genes (AC) had a 13.0- and 7.6-kb fragment (Table I, Fig. 4). We have also found the 13.0-kb Hpa I fragment in association with the β'-gene in the Black population studied (Table I).

**Table I**

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β-Thal., β-thalassemia.
* One SS individual had 13.0- and 5.6-kb fragments, and one had 13.0- and 7.0-fragments.
† One individual was an SC-Harlem variant.

**FIGURE 1** Restriction map of Hpa I sites within and surrounding the δ- and β-globin genes. These sites, originally described by Kan and Dozy (1), were confirmed by digests of normal, sickle cell, and Lepore DNA with Hpa I and Eco RI, alone and in combination. Hpa I generates one fragment, 7.6 kb in size, that contains the entire β-globin gene. In some sickle-cell individuals, variants other than 7.6 kb are found. Hpa I cleaves the intervening sequence of the β-globin gene to yield two fragments: 1.8 kb containing the 5' end of the δ-gene and most of the δ-intervening sequence and 1.3 kb comprising the 3' end of the δ-gene.

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We have detected other polymorphisms in patients with sickle-cell anemia. One individual had only 13.0- and 7.0-kb *Hpa* I fragments. Kan and Dozy (1) reported the presence of a 7.0-kb variant with the β*-gene. Our results indicate that this variant is also found with the β*-gene, although the 7.0-kb fragments seen in these two cases may not be identical. Another SS patient showed two *Hpa* I β-globin fragments (13.0 and 5.6 kb) (Fig. 2). The 5.6-kb band hybridized with consistently greater intensity than other β-globin fragments in several separate experiments. This is not a result of better transfer because of its small size since other comparably sized fragments are not nearly as intense. In this case, the 5.6-kb fragment may either be associated with the β*-gene, or the patient may be homozygous

**FIGURE 2** *Hpa* I restriction fragments containing β- and δ-globin sequences from homozygotes with hemoglobin A (AA) or sickle hemoglobin (SS). Sizes on the left are in kilobases of DNA. 20 μg of each DNA were electrophoresed in a 0.9% neutral agarose gel, transferred to nitrocellulose filters, and hybridized to 32P-labeled β-globin cDNA from plasmid JW102. Globin fragment sizes were estimated by comparison with a Hind III digest of phage λ-DNA run in a parallel channel. Column 1 shows DNA from an AA individual with only a 7.6-kb β-band. Columns 2, 3, and 4 show DNA from SS individuals. Column 2, only the 13.0-kb β-fragment; Column 3, 13.0-kb and 5.6-kb β-fragments; and Column 4, only the 7.6-kb band. The 1.3-kb δ-fragment only weakly hybridizes to the probe and may be detected in long exposures (>7 d) of film. The weak hybridization may reflect poor homology between the 3’ end of the δ-gene and the β-cDNA probe.

**FIGURE 3** *Hpa* I restriction pattern of β- and δ-globin genes in a family. The procedure is described in Methods and Fig. 2. From left to right: AS son (7.6/7.6); SS daughter (13.0/7.6); AS father (13.0/7.6); and AS mother (7.6/7.6).
sickle-cell patients. Although we confirm that the Hpa I site changed is 3' to the β-structural gene as previously reported (1), the exact nature of this polymorphism has not been determined. For example, the 13.0-kb fragment could result from a single nucleotide change in the recognition sequence of Hpa I or the insertion of a DNA fragment. The frequency of the 13.0-kb variant with the β* gene in our study was 58%.

**DISCUSSION**

Our results indicate the considerable heterogeneity in the restriction fragments detected with Hpa I in

**FIGURE 4** Hpa I restriction pattern of β- and δ-globin genes in individuals with the β* gene. Left to right: an SC patient (13.0/13.0); AA homozygote (7.6/7.6); and a CC patient (13.0/7.6).

for the 13.0-kb variant (i.e., 13.0/13.0), and also have an extra 5.6-kb fragment unrelated to a functional β* gene. One parent of this patient had 7.6- and 13.0-kb fragments; the other has not been available for study. This patient's DNA also yields a unique restriction pattern with β-cDNA when cleaved with several other enzymes. Large-sized fragments (one 60 kb) which contained β-like gene material were seen with Eco RI, which suggests a partial gene duplication not involving the intragenic Eco RI site (Fig. 5).

**FIGURE 5** Eco RI restriction pattern of β- and δ-globin genes in one AA and three SS patients. DNA from the patient with the intensely hybridizing 5.6-kb fragment in Fig. 2 is SS2. The AA patient and patients SS1 and SS3 have the normal Eco RI fragments previously reported (1, 4, 5). The SS2 DNA has at least two larger DNA fragments which contain β-like genes not seen in the other samples.
substantially less than the 87% frequency previously reported (1). However, it will require analysis of a much larger population study to determine the true incidence of the 7.6- and 13.0-kb fragments with the β°-gene. We detected no correlation of the 13.0-kb variant with the clinical severity of sickle-cell disease. Surprisingly, we also found the 13.0-kb variant commonly associated with the β°-gene. In addition, three of the βA-genes were linked to the 13.0-kb fragments. The finding that the 13.0-kb variant occurs with the β°- and βA-genes suggests that the 13.0-kb fragment is not specifically linked to the β°-gene and probably occurred in a subpopulation of β°-Blacks who subsequently developed the β°- and βA-genes. Kurnit (10) recently suggested that the βA-genes associated with 7.6- and 13.0-kb Hpa I fragments arose independently. Our data are consistent with this interpretation, and would further suggest that the βA-genes also arose independently in separate 7.6- and 13.0-kb Hpa I-containing populations. The New York Black population studied is of African origin, and migrated to New York from the southern United States and the West Indies. In this regard, we have recently found a high incidence of the 13.0-kb fragment in Blacks of West African origin (Nigeria and Ghana). Large-scale studies of defined Black populations should clarify the true frequency and origins of the fragments in different populations. The specific significance of the linkage of the β°-gene with the 13.0-kb fragment, if any, remains to be elucidated.

Our results also indicate that the application of this polymorphism in the prenatal diagnosis of sickle-cell disease is less useful than previously suggested (11). Clearly, DNA analysis with amniotic fluid cells is more desirable than fetoscopy because of the reduced risk to mother and fetus and a relatively easier technical procedure. However, the Hpa I polymorphism will be most useful in cases where both parents possess the 13.0/7.6 genotype. Only 30% (3 of 10) of our sickle-trait subjects carried the 13.0/7.6-kb genotype. The Hpa I polymorphism will be less useful in families in which one βAβ°-parent has 13.0/7.6 genotype and the other βAβ°-parent has 7.6/7.6. If amniocentesis DNA reveals a 7.6/7.6 genotype, the βAβ°-genotype is most unlikely. If, however, amniocentesis reveals a 13.0/7.6 genotype, sickle-cell anemia and sickle-cell trait in the fetus cannot be distinguished. This situation poses special problems in genetic counselling.

In the past, polymorphisms have proven exceedingly valuable in many types of genetic analyses. The data presented here and previously (1) indicate that they may become particularly useful at the gene level in the diagnosis and prevention of hereditary defects. It will become increasingly important, however, to define if a specific relationship exists between the polymorphism and the defective gene or whether the association is a result of unrelated genes occurring in the same population. In general, large population studies will be required for an accurate assessment of this kind.

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REFERENCES