# Gangliosides Sensitize Unresponsive Fibroblasts to Escherichia coli Heat-Labile Enterotoxin

JOEL MOSS, SAM GARRISON, PETER H. FISHMAN, and STEPHEN H. RICHARDSON, Laboratory of Cellular Metabolism, National Heart, Lung, and Blood Institute, and Developmental and Metabolic Neurology Branch, National Institute of Neurological and Communicative Disorders and Stroke, National Institutes of Health, Bethesda, Maryland 20205; and Department of Microbiology and Immunology, Bowman Gray School of Medicine, Wake Forest University, Winston-Salem, North Carolina 27103

ABSTRACT Chemically transformed mouse fibroblasts did not raise their cyclic AMP level in response to Escherichia coli heat-labile enterotoxin. These fibroblasts did, however, incorporate exogenous mono-, di-, and trisialogangliosides. After the uptake of monosialoganglioside galactosyl-N-acetylgalactosaminyl-[N-acetvlneuraminyll-galactosylglucosylceramide  $(G_{M1})$ , the cells responded to E. coli heat-labile enterotoxin. The di- and trisialogangliosides were considerably less effective. G<sub>M1</sub>, the putative cholera toxin (choleragen) receptor, has been implicated previously as the receptor for E. coli heat-labile enterotoxin based on the ability of the free ganglioside to inhibit the effects of toxin. This investigation establishes that the ganglioside, when incorporated into fibroblasts, serves a functional role in mediating the responsiveness to the toxin.

#### INTRODUCTION

Certain strains of *Escherichia coli* are believed to be responsible for "traveler's diarrhea" (1–5). In some cases, the symptoms may result in part from production of a heat-labile enterotoxin (LT)<sup>1</sup> (1–5). LT appears to exert its effects through activation of adenylate

cyclase (3, 6–9), and in this way it is similar to the enterotoxin from *Vibrio cholerae*, choleragen (10). In addition, both choleragen and LT require NAD for activation of adenylate cyclase in disrupted cells (9, 11, 12) and possess NAD glycohydrolase and ADP-ribosyltransferase activities (13–15). It has been proposed that both toxins exert their effects through the NAD-dependent ADP-ribosylation of either adenylate cyclase itself or of a protein critical to cyclase activation (9, 11, 13–17).

LT cross-reacts immunologically with antisera directed primarily against the B-subunits of choleragen (9, 18, 19). It is through the B-subunit that choleragen binds to specific cell surface receptors believed to be ganglioside  $G_{M1}^{-1}$  (10). Hence, it is possible that LT and choleragen share similar receptors. In fact,  $G_{M1}$  can block the biological effects of LT (20, 21), but the affinity of LT for  $G_{M1}$  (as well as the specificity for  $G_{M1}$  relative to other gangliosides) was lower than that observed with choleragen (20). In addition, choleragenoid, a protein composed of the B-subunits of choleragen, was effective in inhibiting the biological activity of LT in some studies but not in others (20, 22, 23). Thus, it is unclear whether the toxins actually share the same receptors.

We have shown that a chemically transformed line of mouse fibroblasts (National Collection of Type Cultures [NCTC] 2071), which is deficient in gangliosides and lacks  $G_{\rm MI}$ , responds to choleragen only after the cells have incorporated exogenous  $G_{\rm MI}$  but not other gangliosides (24, 25). In these experiments, we have examined the responsiveness of these cells to LT, before and after incorporation of gangliosides.

## **METHODS**

NCTC 2071 fibroblasts were grown as described previously (24). Gangliosides were purified from brain or erythrocytes

Received for publication 21 November 1978 and in revised form 3 April 1979.

<sup>^1</sup> Abbreviations and nomenclature used in this paper: cAMP; cyclic AMP;  $G_{M1}$ , galactosyl-N-acetylgalactosaminyl-[N-acetylneuraminyl]-galactosylglucosylceramide;  $G_{M2}$ , N-acetylgalactosaminyl - [N - acetylneuraminyl] - galactosylglucosylceramide;  $G_{M3}$ , N-acetylneuraminylgalactosylglucosylceramide;  $G_{D1a}$ , N-acetylneuraminylgalactosyl-N-acetylgalactosaminyl - [N - acetylneuraminyl] - galactosylglucosylceramide;  $G_{D1b}$ , galactosyl-N-acetylgalactosaminyl-[N-acetylneuraminyl] - N - acetylneuraminyl] - galactosylglucosylceramide;  $G_{T1}$ , N-acetylneuraminylgalactosyl-N-acetylgalactosaminyl-[N-acetylneuraminyl] - N - acetylneuraminyl] - galactosylglucosylceramide; LT, heat-labile enterotoxin; NCTC, National Collection of Type Cultures.

Table I Effect of  $G_{M1}$  on the Response of Ganglioside-Deficient Fibroblasts to LT

G <sub>M1</sub> added	cAMP content		
	No LT	Plus LT	
	pmol/mg protein		
None	13	15	
1 nmol	16	27	

Fibroblasts were incubated for 18 h in 10 ml of NCTC 135 medium with or without  $G_{\rm M1}$ . The medium was aspirated, and the cells were washed twice with 2 ml of medium. LT (258  $\mu g$  protein) was then added to 10 ml of NCTC 135 medium where indicated. After 3 h at 37°C, the medium was aspirated, and 2 ml of 5% TCA was added. The cells were harvested and centrifuged. Samples of the supernate were taken for assay of cAMP as described previously (30). Data are means of values from duplicate incubations. The experiment was repeated twice with similar results.

(26) and appeared to be at least 99% pure when separated by thin-layer chromatography and visualized with resorcinol reagent (27). LT was purified by procedures described earlier (28). The low molecular weight LT isolated by this procedure is enterotoxic in infant rabbits and adult ligated ileal loops.<sup>2</sup> All assays comparing the relative effectiveness of gangliosides were performed in the same experiment. Protein was determined by the method of Lowry et al. (29).

### RESULTS AND DISCUSSION

When grown in ganglioside-free chemically defined medium, the transformed fibroblasts did not respond to LT even after 3 h (Table I). Cells exposed to  $G_{M1}$  and then to LT accumulated cyclic AMP (cAMP);  $G_{M1}$  by itself had no effect. There was a delay before cAMP levels began to rise in  $G_{M1}$ -treated cells exposed to LT (Fig. 1); maximal accumulation occurred by 2 h. These results are similar to those reported previously with choleragen (24).

We examined the effects of several gangliosides on the responsiveness of NCTC 2071 fibroblasts to LT (Table II).  $G_{M1}$  was clearly the most effective followed by  $G_{D1a}^{-1}$  and  $G_{T1}$ ; only a slight response was observed with  $G_{M2}$  and  $G_{D1b}$  and none with  $G_{M3}$ .  $G_{D1a}$  and  $G_{M2}$  were shown previously to sensitize NCTC 2071 cells to choleragen but were much less effective than  $G_{M1}$  (25). The cells contain a highly active sialidase that converts exogenous  $G_{D1a}$  to  $G_{M1}$  (25); under conditions where  $G_{D1a}$ -treated cells became responsive to choleragen,  $\cong 25\%$  of the  $G_{D1a}$  taken up by the cells was converted to  $G_{M1}$  (25). The effects observed with  $G_{T1}$  and  $G_{D1b}$  may also be a result of the enzymatic breakdown of these gangliosides to  $G_{M1}$ . When the fibroblasts

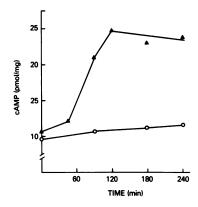


FIGURE 1 Effect of time of incubation with LT on the cAMP content of fibroblasts. Fibroblasts were incubated with ( $\triangle$ ) or without ( $\bigcirc$ )  $G_{M1}$  (1 nmol/10 ml) for 18 h at 37°C. The ganglioside was removed, the cells washed twice with 2 ml of NCTC 135 medium, and 10 ml of NCTC 135 medium was added. LT (172  $\mu$ g) was added where indicated ( $\triangle$ ). At the chosen times, the medium was aspirated, 2 ml of 5% TCA was added, and intracellular cAMP content was determined. Points are the average of duplicate incubations.

were exposed to  $G_{D1b}$ , they became responsive to choleragen (Table III), but again  $G_{M1}$  was more effective. Although the gangliosides used in these studies appear homogeneous, we cannot rule out the possibility that some of the effects observed with other gangliosides might be a result of the presence of trace amounts of  $G_{M1}$  or conversion to  $G_{M1}$  by the cells.<sup>3</sup> It also is conceivable that the higher ganglioside homologues do act directly as receptors for LT.

These studies demonstrate that a line of transformed mouse fibroblasts deficient in gangliosides responds to

TABLE II

Effect of Mono-, Di-, and Trisialogangliosides on the
Response of Transformed Fibroblasts to LT

Ganglioside added	cAMP content	
	pmol/mg protein	
None	6	
$G_{M3}$	6	
$G_{M2}$	10	
$G_{M2}$ $G_{M1}$	21	
$G_{\text{Dia}}$	14	
$G_{D1b}$	9	
$G_{ exttt{D1b}}$ $G_{ exttt{T1}}$	13	

Cells were incubated with gangliosides and LT (258  $\mu$ g), and cAMP was determined as described in Table I. Assays are the average of duplicate incubations. The cAMP content of cells incubated without gangliosides or LT was 6 pmol/mg protein.

<sup>&</sup>lt;sup>2</sup> Personal communication with Dr. D. G. Evans.

<sup>&</sup>lt;sup>3</sup> Both the choleragen and LT preparations used in these studies were assayed for sialidase activity as previously described (25). No activity was detected.

TABLE III

Effect of  $G_{D1b}$  on the Response of the Fibroblasts to Choleragen

Ganglioside added	cAMP content	
	No CT*	Plus CT
	pmol/mg protein	
$G_{\mathtt{M1}} \ G_{\mathtt{D1b}}$	4	15
Gnib	2	9

Cells were incubated with 25 pmol of ganglioside, as described in Table I, and subsequently without or with choleragen,  $10 \mu g/10 \text{ ml}$  of Hanks' solution for 3 h at 37°C. cAMP was determined as described previously. Data are means of values for duplicate incubations.

\* Choleragen.

E. coli LT only after they have been incubated with certain gangliosides. Of these,  $G_{M1}$  is clearly the most effective. These results are similar to those reported previously on the effect of  $G_{M1}$  on choleragen responsiveness and indicate that  $G_{M1}$  can serve as the functional receptor for both toxins in these cells. Thus, although the effects of choleragenoid and gangliosides on the responsiveness of cells to LT were reported to differ from those noted with choleragen (20, 22), it would appear from this study that  $G_{M1}$  can mediate the action of LT on animal cells. Whether  $G_{M1}$  is the native receptor for LT in other cells, especially intestinal cells, has yet to be determined.

### **ACKNOWLEDGMENTS**

We thank Dr. Martha Vaughan for many useful discussions and critical review of the manuscript and Miss Sally J. Stanley for expert technical assistance.

This work was supported in part by grant AI-07772 to Dr. Richardson from the U. S.-Japan Cooperative Medical Science training program, National Institute of Allergy and infectious Diseases, National Institutes of Health.

#### REFERENCES

- Gorbach, S. L., B. H. Kean, D. G. Evans, D. J. Evans, Jr., and D. Bessudo. 1975. Travelers' diarrhea and toxigenic Escherichia coli. N. Engl. J. Med. 292: 933-936.
- Sack, R. B. 1975. Human diarrheal disease caused by enterotoxigenic Escherichia coli. Annu. Rev. Microbiol. 29: 333-353.
- Kantor, H. S., P. Tao, and S. L. Gorbach. 1974. Stimulation of intestinal adenyl cyclase by *Escherichia coli* enterotoxin: comparison of strains from an infant and an adult with diarrhea. *J. Infect. Dis.* 129: 1-9.
- Merson, M. H., G. K. Morris, D. A. Sack, J. G. Wells, J. C. Feeley, R. B. Sack, W. B. Creech, A. Z. Kapikian, and E. J. Gangarosa. 1976. Travelers' diarrhea in Mexico. A prospective study of physicians and family members attending a congress. N. Engl. J. Med. 294: 1299-1305.
- Finkelstein, R. A., M. L. Vasil, J. R. Jones, R. A. Anderson, and T. Barnard. 1976. Clinical cholera caused by an

- enterotoxigenic Escherichia coli. J. Clin. Microbiol. 3: 382-384.
- Evans, D. J., Jr., L. C. Chen, G. T. Curlin, and D. G. Evans. 1972. Stimulation of adenyl cyclase by Escherichia coli enterotoxin. Nat. New Biol. 236: 137-138.
- Hewlett, E. L., R. L. Guerrant, D. J. Evans, Jr., and W. B. Greenough III. 1974. Toxins of Vibrio cholerae and Escherichia coli stimulate adenyl cyclase in rat fat cells. Nature (Lond.). 249: 371-373.
- Hynie, S., H. Rašková, T. Sechser, J. Vaněček, D. Matějovská, V. Matějovská, M. Treu, and L. Polák. 1974.
   Stimulation of intestinal and liver adenyl cyclase by enterotoxin from strains of *Escherichia coli* enteropathogenic for calves. *Toxicon*. 12: 173-179.
- Gill, D. M., D. J. Evans, Jr., and D. G. Evans. 1976. Mechanism of activation of adenylate cyclase in vitro by polymyxin-released, heat-labile enterotoxin of *Escheri*chia coli. J. Infect. Dis. 133(Suppl): S103-S107.
- van Heyningen, S. 1977. Cholera toxin. Biol. Rev. Camb. Philo. Soc. 52: 509-549.
- Gill, D. M. 1975. Involvement of nicotinamide adenine dinucleotide in the action of cholera toxin in vitro. Proc. Natl. Acad. Sci. U. S. A. 72: 2064-2068.
- 12. Gill, D. M. 1976. Multiple roles of erythrocyte supernatant in the activation of adenylate cyclase by *Vibrio cholerae* toxin in vitro. *J. Infect. Dis.* 133(Suppl): S55-S63.
- 13. Moss, J., V. C. Manganiello, and M. Vaughan. 1976. Hydrolysis of nicotinamide adenine dinucleotide by choleragen and its A protomer: possible role in the activation of adenylate cyclase. *Proc. Natl. Acad. Sci. U. S. A.* 73: 4424-4427.
- Moss, J., and M. Vaughan. 1977. Mechanism of action of choleragen. Evidence for ADP-ribosyltransferase activity with arginine as an acceptor. J. Biol. Chem. 252: 2455– 2457
- 15. Moss, J., and S. H. Richardson. 1978. Activation of adenylate cyclase by heat-labile *Escherichia coli* enterotoxin. Evidence for ADP-ribosyltransferase activity similar to that of choleragen. *J. Clin. Invest.* 62: 281-285.
- Cassel, D., and T. Pfeuffer. 1978. Mechanism of cholera toxin action: covalent modification of the guanyl nucleotide-binding protein of the adenylate cyclase system. Proc. Natl. Acad. Sci. U. S. A. 75: 2669-2673.
- Gill, D. M., and R. Meren. 1978. ADP-ribosylation of membrane proteins catalyzed by cholera toxin: basis of the activation of adenylate cyclase. *Proc. Natl. Acad. Sci.* U. S. A. 75: 3050-3054.
- Evans, D. J., Jr., D. G. Evans, and S. L. Gorbach. 1974.
   Polymyxin B-induced release of low-molecular-weight, heat-labile enterotoxin from Escherichia coli. Infect. Immun. 10: 1010-1017.
- Gyles, C. L. 1974. Relationships among heat-labile enterotoxins of Escherichia coli and Vibrio cholerae. J. Infect. Dis. 129: 277-283.
- Holmgren, J. 1973. Comparison of the tissue receptors for Vibrio cholerae and Escherichia coli enterotoxins by means of gangliosides and natural cholera toxoid. Infect. Immun. 8: 851-858.
- Zenser, T. V., and J. F. Metzger. 1974. Comparison of the action of Escherichia coli enterotoxin on the thymocyte adenylate cyclasecyclic adenosine monophosphate system to that of cholera toxin and prostaglandin E<sub>1</sub>. Infect. Immun. 10: 503-509.
- Pierce, N. F. 1973. Differential inhibitory effects of cholera toxoids and ganglioside on the enterotoxins of Vibrio cholerae and Escherichia coli. J. Exp. Med. 137: 1009-1023.
- 23. Nalin, D. R., and J. C. McLaughlin. 1978. Effects of

- choleragenoid and glucose on the response of dog intestine to *Escherichia coli* enterotoxins. *J. Med. Microbiol.* 11: 177-186.
- Moss, J., P. H. Fishman, V. C. Manganiello, M. Vaughan, and R. O. Brady. 1976. Functional incorporation of ganglioside into intact cells: induction of choleragen responsiveness. *Proc. Natl. Acad. Sci. U. S. A.* 73: 1034– 1037.
- Fishman, P. H., J. Moss, and M. Vaughan. 1976. Uptake and metabolism of gangliosides in transformed mouse fibroblasts. Relationship of ganglioside structure to choleragen response. J. Biol. Chem. 251: 4490-4494.
- Pacuszka, T., R. O. Duffard, R. N. Nishimura, R. O. Brady, and P. H. Fishman. 1978. Biosynthesis of bovine thyroid gangliosides. J. Biol. Chem. 253: 5839-5846.
- Fishman, P. H., R. O. Brady, R. M. Bradley, S. A. Aaronson, and G. J. Todaro. 1974. Absence of a specific gan-

- glioside galactosyltransferase in mouse cells transformed by murine sarcoma virus. *Proc. Natl. Acad. Sci. U. S. A.* 71: 298–301.
- 28. Richardson, S. H., W. F. Osborne, D. E. Lockwood, and S. H. Love. 1976. Purification of E. coli heat labile enterotoxin (LT) and its immunological relatedness to isolated subunits of choleragen. In Symposium on Cholera, Proceedings of the 12th Joint Conference U. S.-Japan Cooperative Medical Science Program, Sapporo, Japan. H. Fukumi and Y. Zinnaka, editors. 211-223.
- Lowry, O. H., N. J. Rosebrough, A. L. Farr, and R. J. Randall. 1951. Protein measurement with the Folin phenol reagent. J. Biol. Chem. 193: 265-275.
- Clyman, R. I., A. S. Blacksin, J. A. Sandler, V. C. Manganiello, and M. Vaughan. 1975. The role of calcium in regulation of cyclic nucleotide content in human umbilical artery. J. Biol. Chem. 250: 4718-4721.