Effects of Replacement Doses of Sodium-L-Thyroxine on the Peripheral Metabolism of Thyroxine and Triiodothyronine in Man

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ABSTRACT Studies of the effect of L-thyroxine administration (0.3 mg daily for 7-9 wk) on the peripheral metabolism of ¹³¹I-labeled triiodothyronine (T₃) and ¹²⁵Ilabeled thyroxine (T₄) and on the concentration and binding of T4 and T8 in serum were carried out in 11 euthyroid female subjects. Administration of L-thyroxine led to consistent increases in serum T₃ concentration (137 vs. 197 ng/100 ml), T₈ distribution space (39.3) vs. 51.7 liters), T₈ clearance rate (22.9 vs. 30.6 liters/ day) and absolute T₈ disposal rate (30 vs. 58 μ g/day), but no change in apparent fractional turnover rate (60.3 vs. 60.6%/day). The proportion and absolute concentration of free T₈ also increased during L-thyroxine administration. Increases in serum total T4 concentration (7.3 vs. 12.8 µg/100 ml) and in both the proportion and absolute concentration of free thyroxine also occurred. In five of the subjects, the kinetics of peripheral T4 turnover were simultaneously determined and a consistent increase in fractional turnover rate (9.7 vs. 14.2%/day), clearance rate (0.84 vs. 1.37 liters/day), and absolute disposal rate (64.2 vs. 185.0 µg/day) occurred during L-thyroxine administration. Despite these increases in the serum concentration and daily disposal rate of both T4 and T3, the patients were not clinically thyrotoxic. However, basal metabolic rate (BMR) values were marginally elevated and, as in frank thyrotoxicosis,

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T₄-binding capacities of thyroxine-binding globulin (TBG) and thyroxine-binding prealbumin (TBPA) reduced, suggesting that subclinical thyrotoxicosis was present. Thus, the often recommended replacement dose of 0.3 mg L-thyroxine daily may be greater than that required to achieve the euthyroid state.

The studies have also provided additional evidence of the peripheral conversion of T₄ to T₈ in man and have permitted the calculation that approximately one-third of exogenously administered T₄ underwent deiodination to form T₈. To the extent that a similar fractional conversion occurs in the normal state, it can be calculated that a major fraction of the T₈ in serum derives from the peripheral deiodination of T₄ and that only a lesser fraction derives from direct secretion by the thyroid gland.

INTRODUCTION

In 1970, studies from our laboratories provided the first conclusive evidence that thyroxine (T₄)¹ is converted

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¹ Abbreviations used in this paper: ETT4, extrathyroidal T4 pool; %FT3, per cent free T5; %FT4, per cent free T4; SSKI, saturated solution of potassium iodide; TBG, thyroxine-binding globulin; TBGcap, measurements of T4-binding capacities by TBG; TBPA, thyroxine-binding prealbumin; TBPAcap, measurements of T4-binding capacities by TBPA; triiodothyronine; T3DS, T3 distribution space; T3(RIA), T3 measured by radioimmunoassay; T4, thyroxine; T4(D), T4 measured by displacement; T4DS, T4 distribution space; TSH, thyroid-stimulating hormone.

to triiodothyronine (T₃) in the peripheral tissues of man (1). This conclusion has since been repeatedly confirmed (2-5), and conversion of T₄ to T₅ has been demonstrated, in addition, in the intact rat (6) and in isolated human cells in vitro (7-9). Our conclusion was based in part on the finding that athyreotic patients receiving exogenous L-thyroxine therapy had in their serum concentrations of T₃ far greater than those which would have been expected on the basis of contamination of the administered T. by T. (1). An unexplained feature of this study was the finding that although these patients did not appear thyrotoxic by clinical criteria, serum T₄ concentrations were well into the range found in spontaneously thyrotoxic patients, and serum Ts concentrations also appeared distinctly elevated by the methods of analysis then available.

In an effort to examine this apparent discrepancy, to examine the conjointly-measured kinetics of peripheral T₄ and T₅ metabolism in patients receiving L-thyroxine, and to assign quantitative dimensions to the extent of peripheral conversion of T₄ to T₅, studies were conducted in which the effects of L-thyroxine on the peripheral metabolism of T₄ and T₅ in euthyroid individuals were assessed. A portion of the studies has been the subject of a preliminary communication (10).

METHODS

Studies were conducted in 11 normal female volunteers ranging from 22 to 54 yr in age. Studies of the peripheral metabolism of T_s were carried out in all 11 before and after a 7–9 wk period of L-thyroxine administration (0.3 mg/day orally [P.O.]). In five, studies of the peripheral metabolism of T_s were conducted concurrently with those of T_s .

Studies of the peripheral metabolism of T₂ were made using 1811 I-labeled T_{3.2} After its receipt from a commercial source, labeled T₈ was mixed with sterile human serum albumin (final concentration 4 g/100 ml) and was dialyzed against 2 liters of sterile physiological saline solution at 4°C for 2 h. As judged by chromatography in two solvent systems in which iodide and T4 or T3 were well separated (butanol-ethanol ammonia, 5:1:2; butanol-dioxane ammonia, 4:1:5), this procedure decreased the contaminating [121] iodide from approximately 1.6 to approximately 0.6%. Approximately 1% of the 181 I migrated as T4, and this was unchanged by dialysis. Dialyzed solutions were passed through a Millipore filter before their administration to patients. Patients received a single intravenous (i.v.) injection of 100 μ Ci (4.6-8.5 μ g) of isiI-labeled T₈. Blood samples were drawn at 24-h and at 12-h intervals thereafter for the next 2 days. Sera were separated and were treated with Iobeads to remove labeled inorganic iodide.8

Studies of the peripheral metabolism of T_4 were conducted with ¹²⁵I-labeled hormone that was not predialyzed but which was diluted with sterile human serum albumin (final concentration 4 g/100 ml) and passed through a Millipore filter before administration to the patients. Chromatographic analysis revealed approximately 2% of the radioactivity to be in the form of iodide and none in the form of T_3 . Each of the five patients in whom such studies were carried out received a single i.v. injection of 40 μ Ci (approximately 4 μ g) of ¹²⁶I-labeled hormone administered immediately after the injection of ¹²⁶I-labeled T_3 . In these patients, in addition to the samples of serum obtained for studies of T_3 metabolism, specimens were obtained daily for 6 more days.

2 ml samples of serum were counted for ¹⁸¹I and ¹⁸⁵I in a scintillation spectrometer, correction being made when indicated for the ¹⁸¹I counts detected in the ¹⁸⁵I spectrum. Radioactivity in serum samples was expressed in relation to that present in appropriately diluted samples of the administered dose. Calculations of the kinetics of T₈ metabolism were based on data obtained in samples drawn from 24 to 72 h after administration of [¹⁸¹I]T₈ and, in the case of T₄, samples drawn from 2 to 9 days after [¹⁸⁵I]T₄ administration. Fractional turnover rates were calculated by the method of least squares, and volumes of T₄ and T₈ distribution were determined by extrapolation of the calculated regression curves to zero time.

In order to prevent the thyroidal uptake of inorganic ¹³¹I or ¹³⁵I generated by the peripheral deiodination of T₄ or T₈ during the control period, five drops of saturated solution of potassium iodide (SSKI) were given orally each day. SSKI was also administered during the second turnover study, although thyroidal iodine uptake had been shown to be suppressed by the exogenous hormone administered.

Samples of serum from each turnover study were pooled and analyzed in duplicate for T4 concentration (T4[D]) by a modification of the method of Murphy and Pattee (12) and for T₈ concentration (T₈[RIA] by a modification of the radioimmunoassay of Gharib, Mayberry, and Ryan (13), using antibody kindly provided by these authors. By the method employed, normal values in 26 subjects averaged 150±25 ng/100 ml (mean ±SD). Measurements of T₄-binding by TBG (TBG_{eap}) and TBPA (TBPA_{eap}) and of the per cent of free T₄ (%FT₄) and of free T_s (%FT_s) were carried out in sera obtained on two separate days during each turnover study. Absolute concentrations of FT4 and FT8 were calculated as the product of the per cent and total concentration of T4 and T₃ in serum, respectively. For each function studied, analyses of all four sera from each subject were carried out on the same day. TBGcap and TBPAcap were measured by reverse flow and conventional filter paper electrophoresis, respectively, using glycine acetate buffer, pH 8.6, account being taken of the endogenous serum T4 concentration (14). %FT4 and %FT8 were measured in triplicate by a modification of the equilibrium dialysis technique of Ingbar, Braverman, Dawber, and Lee (15).

Statistical analyses were conducted according to methods described by Snedecor (16).

^{2 121}I-labeled T₈ and ¹²⁵I-labeled T₄ were obtained from Abbott Laboratories, Chicago, III.

^a Control studies have revealed that treatment of serum with Iobeads for 5 min removes more than 99% of inorganic ¹²¹I and less than 1% of labeled iodothyronines.

⁴Abbreviations used to denote the serum concentrations of T_4 and T_8 , the proportions and absolute concentrations of T_4 and T_8 and the T_4 -binding capacities of TBG and TBPA are those recently adopted by the American Thyroid Association (11).

TABLE I Effect of L-thyroxine Administration on the Concentration and Binding of T4 and T3 in the Serum of Normal Subjects*

Subject	Age	Treatment	Serum T ₈	PFT ₈	AFT:	Serum T4	PFT4	AFT4	TBG	ТВРА
			ng/100 ml	%	ng/100 ml	ug/100 ml	%	ng/100 ml	ugT4/ 100 ml	ugT ₄ / 100 ml
1	47	Control	132	0.16	0.21	7.0	0.018	1.3	26.0	240
		LT ₄ ‡	208	0.21	0.44	11.5	0.025	2.8	23.0	196
2	30	Control	100	0.16	0.16	6.5	0.018	1.2	23.5	205
		LT_4	160	0.22	0.36	12.0	0.025	3.0	16.5	173
3	49	Control	144	0.19	0.27	5.5	0.018	1.0	17.0	208
		LT ₄	184	0.26	0.48	11.0	0.026	2.8	15.5	187
4	33	Control	168	0.16	0.28	7.5	0.018	1.4	22.0	169
		LT_4	212	0.21	0.44	13.5	0.022	2.9	19.0	155
5	25	Control	120	0.17	0.21	7.0	0.019	1.3	23.5	207
		LT ₄	200	0.24	0.49	13.0	0.027	3.6	20.0	176
6	38	Control	132	0.19	0.25	9.0	0.018	1.6	21.0	269
-		LT4	200	0.23	0.46	12.5	0.022	2.7	21.0	226
7	22	Control	140	0.17	0.24	6.5	0.018	1.1	23.0	228
		LT ₄	156	0.22	0.34	12.0	0.022	2.7	19.0	204
8	54	Control	156	0.18	0.28	8.0	0.017	1.4	19.0	241
_	-	LT ₄	208	0.25	0.52	15.0	0.031	4.6	16.5	197
9	26	Control	152	0.19	0.29	8.5	0.018	1.5	21.5	231
•		LT ₄	192	0.21	0.41	13.5	0.023	3.2	21.5	187
10	37	Control	116	0.19	0.23	7.5	0.019	1.4	21.0	236
	• •	LT ₄	152	0.24	0.37	13.0	0.024	3.1	21.0	190
11	29	Control	152	0.17	0.27	7.5	0.016	1.2	23.5	229
		LT.	232	0.23	0.53	14.0	0.021	2.9	19.0	182
Mean ±	-SD	Control	137 ± 20	0.18 ± 0.01	0.24 ± 0.03	7.3 ± 1.0	0.018 ± 0.001	1.3 ± 0.2	21.9 ± 2.5	223 ± 26
		LT.	191 ± 25	0.23 ± 0.02	0.44 ± 0.06	12.8 ± 1.1	0.024 ± 0.003	3.1 ± 0.6	19.3 ± 2.4	188 ± 18
P value	P value (paired t test)		< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.001	< 0.005	0.001

^{*} PFT₃, proportion of free T₃; AFT₃, absolute concentration of free T₃; PFT₄, proportion of free T₄; AFT₄, absolute concentration of free T₄.

RESULTS

Concentration and binding of T. in serum (Table I). Before administration of L-thyroxine, T₄(D) in the 11 patients studied averaged 7.3±1.0 µg/100 ml (mean± SD); %FT4, $0.018\pm0.001\%$; and FT4, 1.29 ± 0.19 ng/100ml. All values were within the normal range. Also normal were values for TBG_{cap} (21.9 \pm 2.4 μ g/100 ml) and TBPAcap (223±26 µg/100 ml). During L-thyroxine administration, mean T₄(D) increased to 12.8 \pm 1.0 μ g/100 ml and %FT4 to an average of 0.024±0.003%. Values for FT4 were strikingly increased, averaging 3.1±0.5 ng/ 100 ml, an increase of 141% from control values. TBG_{cap} (19.3±2.4 μg T₄/100 ml) and $TBPA_{\text{cap}}$ (188± 18 µg T₄/100 ml) were decreased slightly, but significantly, from control values, (P < 0.005 and P < 0.001, respectively).

Peripheral metabolism of T₄ (Table II). In the five subjects in whom the appropriate studies were performed, indices of the peripheral metabolism of T4 during control periods were within the normal range. During the administration of L-thyroxine, T4 distribution space (T₄DS) was not significantly changed (8.7±0.8 liters vs. 9.7±1.0 liters), but the fractional rate of turnover (K) increased from a mean of $9.7\pm0.8\%/\text{day}$ to a mean of 14.2±1.8%/day. As a result, the rate of total T₄ clearance (C) increased from a mean of 0.84±0.09 to a mean of 1.37±0.10 liters/day. The content of the extrathyroidal T4 pool (ETT4) increased during the administration of L-thyroxine by an average of 100%. Total daily disposal of T₄ increased markedly from a mean of 64.2± 9.6 to a mean of $185\pm23 \,\mu g \, T_4/day$.

Concentration and binding of T_s in serum (Table I). During administration of L-thyroxine, values of T₈(RIA) increased from a control mean of 137±20 to a mean of 191±25 ng/100 ml. %FTs, which averaged 0.18±0.01 in the control period, increased to a mean of 0.23±0.02%. Values for FT3 were about doubled, increasing from a mean of 0.24 ± 0.03 to a mean of 0.44 ± 0.06 ng/100 ml.

Peripheral metabolism of T: (Table III). Before L-thyroxine administration, indices of the peripheral

[‡] Each subject received 0.3 mg L-thyroxine P.O. daily for 7-9 wk.

TABLE II

Effect of L-thyroxine Administration on the Peripheral Metabolism of T₄*

Subject	Age	Treatment	Serum T4	T_4DS	K	ETT4	С	D
			ug/100 ml	liters	%/day	ug	liters/day	ug/day
7	22	Control	6.5	8.6	10.2	559	0.88	57
		LT ₄ ‡	12.0	9.3	13.3	1,116	1.23	147
8	54	Control	8.0	7.8	9.0	624	0.70	56
		LT_4	15.0	10.2	13.8	1,530	1.40	210
9	26	Control	8.5	9.7	9.7	824	0.94	80
		LT ₄	13.5	9.1	15.2	1,228	1.38	186
10	37	Control	7.5	8.1	10.8	607	0.87	65
		LT ₄	13.0	8.9	16.9	1,157	1.50	195
11	29	Control	7.5	9.5	8.9	712	0.84	63
		LT ₄	14.0	11.3	12.0	1,582	1.35	189
Mean ±SD		Control	7.6 ± 0.7	8.7 ± 0.8	9.7 ± 0.8	665 ± 104	0.84 ± 0.09	64.2 ± 9.6
		LT_4	13.5 ± 1.2	9.7 ± 1.0	14.2 ± 1.8	$1,322 \pm 217$	1.37 ± 0.10	185.0 ± 23.0
P value (paired t test)			< 0.001	NS	< 0.005	< 0.005	< 0.005	< 0.001

^{*} T₄DS, T₄ distribution space; K, fractional rate of T₄ turnover; ETT₄, extrathyroidal T₄ pool; C, rate of total T₄ clearance; D, total daily disposal of T₄.

Table III

Effect of L-thyroxine Administration on the Peripheral Metabolism of T_3^*

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Subject	Age	Treatment	Serum T ₃	T ₃ DS	К	ETT:	С	D
			ng/100 ml	liters	%/day	ug	liters/day	ug/day
1	47	Control	132	34.0	64.0	45	21.7	29
		LT ₄ ‡	208	39.8	70.7	83	28.3	59
2	30	Control	100	35.5	61.6	36	21.8	22
		LT_4	160	42.6	67.2	68	28.6	46
3	49	Control	144	32.1	65.8	46	21.1	30
		LT_4	184	60.7	58.1	111	35.2	64
4	33	Control	168	30.0	70.6	50	21.2	35
		LT_4	212	38.7	67.4	82	26.1	55
5	25	Control	120	27.2	72.4	33	19.7	24
		LT ₄	200	47.9	67.8	96	32.5	66
6	38	Control	132	41.4	76.8	55	31.8	42
		LT ₄	200	50.1	76.8	100	38.5	77
7	22	Control	140	50.4	46.7	71	23.5	33
		LT_4	156	59.2	47.3	92	28.0	43
8	54	Control	156	39.0	43.4	61	16.9	26
		LT_4	208	51.7	49.4	107	25.5	53
9	26	Control	152	58.8	44.4	89	26.1	39
		LT ₄	192	59.9	47.2	115	28.3	54
10	37	Control	116	49.4	51.3	57	25.3	29
		LT₄	152	69.3	48.2	105	33.4	51
11	29	Control	152	34.7	66.2	53	23.0	35
		LT₄	232	48.9	66.9	113	32.7	76
Mean ±SD		Control	137 ± 20	39.3 ± 9.7	60.3 ± 11.8	54 ± 16	22.9 ± 3.8	30±6
		LT ₄	191 ± 25	51.7 ± 9.6	60.6 ± 10.9	97 ± 15	30.6 ± 4.0	58 ± 11
P value	e (pairec	l t test)	< 0.001	< 0.001	NS	< 0.001	< 0.001	< 0.001

^{*} T₃DS, T₃ distribution space; K, fractional rate of T₃ turnover; ETT₃, extrathyroidal T₃ pool; C, rate of total T₃ clearance; D, total daily disposal of T₄.

[‡] Each subject received 0.3 mg L-thyroxine P.O. daily for 7-9 wk.

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metabolism of T_8 were similar to those previously reported for euthyroid adults (17–20). The T_8 distribution space (T_8DS) averaged 39.3±9.7 liters; the fractional turnover rate, 60.3±11.8%/day; and the total T_8 clearance rate, 22.9±3.8 liter/day. The extrathyroidal pool of T_8 averaged 54±15 μ g and daily T_8 disposal, 30±6 μ g/day. With the exception of the fractional rate of T_8 turnover (60.6±10.9%/day), which appeared to be unchanged, all other functions cited above increased significantly (P < 0.001) during L-thyroxine administration. Serum T_8 concentration increased by an average of 40%, T_8DS by an average of 32%, total T_8 clearance by 34%, and total daily disposal of T_8 by 93%.

Conversion of T₄ to T₅. On the likely assumption that thyroidal secretion of T₈ during administration of L-thyroxine was negligible, values for T₄ and T₅ disposal during this period could be employed to calculate the proportion of T₄ converted to T₅. Individual values for daily T₅ disposal were multiplied by a factor of 1.193 to ascertain the molar equivalent quantity of T₄. The resulting values were divided by the corresponding values for daily T₄ disposal to obtain the proportion of T₄ converted to T₅. These ranged from 30.2 to 48.0%, averaging 35.7±7.2.

Proportionate conversion of T4 to T3 during control periods could not be estimated, since it was impossible to distinguish T₈ arising from deiodination of T₄ from that secreted directly by the thyroid gland. The latter function could be estimated if it were assumed that values for the proportionate conversion of T₄ to T₃ obtained in each patient during L-thyroxine administration also pertained during control periods. To obtain such estimates, control values for daily T4 disposal were multiplied by the value for percentage conversion of T₄ to T₈ during L-thyroxine administration. The resulting product, representing the micrograms of T₄ estimated to undergo monodeiodination daily, was divided by 1.193 to obtain the molar equivalent quantity of T₃. The resulting value for the micrograms of To generated from To daily was then divided by the total T₃ disposal rate, providing an estimate of the fraction of total daily T₈ disposal derived from conversion of T₄. Values of this estimate ranged between 51 and 72%, averaging $59\pm8\%$.

DISCUSSION

In previous studies, we have demonstrated that athyreotic patients receiving traditional replacement doses of L-thyroxine (0.3 mg daily) have in their serum substantial concentrations of T₈, indicating appreciable conversion of T₄ to T₈ in the peripheral tissues of man (1). This conclusion, which has received strong additional support (2-5) and has been extended to the rat (6), has raised inter-related questions as to the origin of the

T_s found in serum, the nature of the thyroidal secretory product, and the ultimately active form of the thyroid hormone. Central to each of these questions is the actual extent of peripheral conversion of T₄ to T₅; however, such could not be determined from our previous studies in athyreotic patients, since the rates of turnover and clearance of T₄ and T₅ were not ascertained.

The present studies were undertaken to explore this and other questions concerning thyroid hormone physiology in patients receiving replacement doses of L-thyroxine. Studies were conducted in euthyroid subjects so that absolute rates of T₄ and T₈ turnover and disposal during L-thyroxine administration could be compared with those obtained in the basal state. Moreover, in view of the marked suppression of thyroid function produced by L-thyroxine administration, as evidenced by decreases in 24 h thyroid ¹⁸¹I uptakes from a mean of 28 to a mean of 3%, T₈ present in serum during this period can be considered to have arisen largely or entirely from extrathyroidal sources, permitting estimates of the rate of T₄ to T₈ conversion.

The normal or increased concentrations of Ta found in serum of the present patients during L-thyroxine administration have confirmed the findings obtained in the anatomically athyreotic patients reported earlier.5 In addition, in the five patients in whom concurrent studies of T₄ and T₈ disposal were carried out, the extent to which T₄ was converted to T₈ during L-thyroxine administration could be estimated, since thyroidal secretion of T₈ could be presumed to be negligible. Calculated values for the per cent of T4 converted to T3 ranged from 30.2 to 48.0, averaging 35.7±7.2. These results agree closely with those obtained by Pittman, Chambers, and Read (2) based on studies with ["C]T₄. They also agree rather well with those obtained by Surks, Schadlow, and Oppenheimer (4) in similar studies of four athyreotic patients treated with L-thyroxine. They reported that approximately half of the T4 deiodinated was converted to Ts. Since about 20% of Ts is metabolized by nondeiodinative pathways, a value of approximately 40% overall T4 to T8 conversion is obtained. Similar estimates have been obtained by Cullen, Burger, Woeber. and Ingbar in untreated euthyroid subjects by a differ-

⁵ When, in 1969 and 1970, the present studies were actually performed, analyses for serum T₂ were conducted by displacement analysis (21), a method now known to yield values substantially and possibly erroneously higher than those obtained by radioimmunoassay. At that time, it was beginning to be evident that this was the case. Therefore, samples of serum were kept continuously frozen until clarification of methodological problems was achieved and a radioimmunoassay for T₂ was established in the authors' laboratories. The values herein presented are the results of the latter analyses, and differ substantially in a quantitative sense from those presented in our preliminary report (10).

ent technique in which conversion is calculated from the concurrently measured clearance rates of $[^{125}I]T_4$ and $[^{131}I]T_3$ and equilibrium values for the ratio of $[^{125}I]T_4$ / $[^{125}I]T_4$ in serum.⁶

The finding that about one-third or one-half of the peripheral T4 is metabolized to T3, together with the twoor threefold greater metabolic potency of Ta than Ta, has led to the suggestion that virtually all of the metabolic activity of T4 resides in the T3 generated from it and that T₄ is merely a prohormone for T₈. Several considerations prompt caution in this interpretation, however. All estimates of T4 to T3 conversion to date have been what might be termed "plasma conversion rates," since they are based on concentrations and disappearance rates for T₈ in plasma. They do not reflect any T₈ generated from T4 that is locally degraded or excreted without entering the plasma. The extent to which true conversion rates exceed plasma conversion rates is unknown. Moreover, it is unknown whether T₈ generated and metabolized locally has any metabolic action and, if it does, how its intrinsic potency compares with that of T3 that enters the cell from the extracellular fluid. In view of these uncertainties, conclusions concerning the intrinsic potency of T4 based on the fractional conversion of T4 to T_s and the overall potency of the two in the whole animal would seem premature.

Plasma conversion rates do bear directly, however, on the proportion of total daily disposal of T_s that is derived from the monodeiodination of T₄, when disposal rates are based upon plasma disappearance rates and plasma concentrations of the two hormones. Thus, the plasma T₄ to T₈ conversion rate times the daily disposal of T₄, when corrected for the lower molecular weight of T₈, will indicate the quantity of T₈ in plasma derived from T₄ daily. The present estimates of conversion rates derived from data obtained while patients were receiving L-thyroxine, when applied to the present conjoint measurements of T₄ and T₈ disposal in the basal state, would indicate that approximately 60% of daily T₈ disposal (and of plasma T₈ concentration) derives from the monodeiodination of T₄.

At the time these studies were carried out, the effect of labeled iodoproteins arising from $T_{\rm s}$ deiodination on the apparent disappearance from the serum of the labeled $T_{\rm s}$ was just becoming evident (22). Hence, separation of labeled iodoproteins from the labeled $T_{\rm s}$ remaining in serum was not carried out, although removal of labeled iodide was. The question arises, therefore, as to the extent to which the measured kinetics of $T_{\rm s}$ metabolism were influenced by failure to exclude iodoprotein. It is most unlikely that this influenced the results of control studies materially, since values for

T₂DS and fractional turnover rate did not differ substantially from those obtained when labeled iodoprotein is excluded (19). This is doubtless the result of the fact that in normal individuals iodoprotein contributes but little to the plasma radioactivity during the 24 to 72 h period in which measurements are made (17). The effect of iodoprotein on measurements made during L-thyroxine administration is less certain, however. To the extent that the clearance of T₈ is increased, lower values for plasma labeled T₈ concentration at any point in time would be expected. In addition, to the extent that increased clearance of T₈ resulted from increased deiodination, a more rapid generation of iodoprotein would be expected. Together, these effects would yield a slower apparent rate of decline of T₃ concentration in plasma than is actually the case. This effect was probably operative in the present studies, and may have resulted in the lack of change in values for T₃ fractional turnover rate during L-thyroxine administration.

It is substantially less clear, however, what the effect of iodoprotein on calculated values for T₂DS would be. If, as previous studies would suggest (17), iodoprotein is but a small percentage of total organically-bound radioiodine at 24 h after labeled T₃ administration and is a progressively larger proportion at later times, then the zero time intercept of the least squares regression curve based on total organic radioiodine would be erroneously low and values for TaDS erroneously high. This spurious increase in T₃DS would tend to reduce, but not eliminate, the error in calculated values for T₃ clearance caused by underestimation of the fractional turnover rate. The magnitude of such an error is difficult to estimate, however, since there have apparently been no studies in which sera freed of radioiodide were studied with and without removal of iodoprotein to ascertain the magnitude of the effect of including iodoprotein on calculated values of T₈DS and fractional turnover rate.7 It is apparent, however, that, to the extent that T3 clearance was underestimated, then the percentage peripheral conversion of T4 to T3 would have been proportionately underestimated and calculated values for the thyroidal secretion of T₈ too high. The close agreement between values for T4 to T3 conversion found in the present studies and those found in the studies of Pittman, Surks, and Cullen and their respective co-

⁶ Cullen, M. J., A. Burger, K. A. Woeber, and S. H. Ingbar. Unpublished observations.

⁷ Nicoloff, Low, Dussault, and Fisher (19) have published curves depicting plasma disappearance of total radio-activity and iodothyronine radioactivity after administration of labeled T₈. From these curves it is quite clear that values for both T₈DS and k would be greatly reduced if total plasma radioactivity, rather than iodothyronine activity, were considered. However, since inorganic iodide comprises approximately 40% of total plasma radioactivity after a dose of labeled T₃ (17), much of the apparent decrease in T₈DS can be ascribed to inclusion of iodide, rather than iodoprotein.

workers (2, 4) in which iodoprotein was eliminated from serum before analysis, would suggest, however, that the error introduced by our failure to eliminate iodoproteins was small.

Until recently, it has been assumed that adequate replacement therapy with L-thyroxine would require doses sufficient to establish values for serum T₄ concentration above the normal range, since the metabolic contribution normally provided by T₅ was presumed to be lacking (23–27). With the demonstration of peripheral T₄ to T₅ conversion, however, the rationale for this approach no longer seemed valid, provided that the rate of production of T₅ was physiologically significant. Although our earlier studies of athyreotic patients receiving L-thyroxine therapy revealed normal or increased concentrations of serum T₅, production rates could not be inferred, since peripheral T₅ turnover was not studied under these circumstances.

The present studies have shown, however, that euthyroid patients receiving 0.3 mg L-thyroxine daily undergo at least a doubling in the daily disposal rate for T₈ and a trebling in that for T4, as compared to the production and disposal rates sustained by their endogenous thyroid function. It would appear, therefore, that the conventional replacement dose of 0.3 mg of L-thyroxine daily does provide both T₄ and T₈ in excess of normal physiological requirements. One might then expect patients receiving this dose to manifest signs of thyroid hormone excess. Frank thyrotoxicosis would be unlikely in view of the fact that quantities of T4 and T3 provided by 0.3 mg L-thyroxine daily, although greater than normal, were far less than the usual production rates of T₄ and T₃ in hyperthyroid patients, as judged from reports in the literature (17, 19, 28-33). The present patients manifested no clinical signs of thyrotoxicosis. On the other hand, several findings provide suggestive though not conclusive evidence that they were mildly hypermetabolic. First, in six of the eight patients, values of the BMR (+8-+15%) during suppressive therapy were above the normal range for our laboratory (-15+5%); however, in none were pretreatment values available.

Second, TBG_{cap} and TBPA_{cap} decreased during L-thyroxine therapy, changes reminiscent of those found in patients with spontaneous thyrotoxicosis (34, 35). The few available data of other workers also supports the conclusion that 0.3 mg of L-thyroxine daily may often be an excessive replacement dose. Most authors emphasize the difficulty of selecting between 0.2 mg daily and 0.3 mg daily on the basis of subjective findings alone (26, 27, 36–38); however, when objective criteria such as BMR have been assessed, doses of 0.2 mg daily frequently seem sufficient (36, 39). The most telling evidence, however, is the recent finding of Cotton, Gorham,

and Mayberry (40) that in patients with hypothyroidism doses of 0.2 mg daily usually suffice to restore serum thyroid-stimulating hormone (TSH) concentration to normal.

The apparent physiological (and clinical) tolerance of euthyroid young adults to slight or moderate excesses of thyroid hormone is well-known, particularly from instances where the hormone is administered in an effort to treat obesity. In addition, patients with autonomous thyroid nodules and complete suppression of paranodular thyroid function often are not detectably thyrotoxic, although it is most unlikely that the nodule is secreting quantities of hormone precisely equivalent to physiological needs. Such apparent resistance to mild excesses of thyroid hormone may merely bespeak the insensitivity of clinical and biochemical indices of thyroid hormone excess currently available. It might also reflect enhanced activity of pathways for hormonal metabolism that are purely degradative or excretory, i.e., that forestall hormonal action. Finally, to the extent that hormonal excess is judged to exist on the basis of increased serum concentration or disposal rate of T4, the validity of such judgments will depend upon the extent to which T4 proves to have a metabolic action independent of its conversion T₃. All of these considerations, and perhaps others, await clarification.

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