# Evidence for *In Vivo* Breakdown of β<sub>1c</sub>–Globulin in Hypocomplementemic Glomerulonephritis \*

CLARK D. WEST,† SARA WINTER, JUDITH FORRISTAL, JANICE M. McCONVILLE, AND NEIL C. DAVIS

(From the Children's Hospital Research Foundation and the Department of Pediatrics, University of Cincinnati College of Medicine, Cincinnati, Ohio)

Summary. Evidence has been obtained for the presence in vivo of alpha<sub>2D</sub>-globulin, a breakdown product of serum  $\beta_{1C}$ -globulin, in patients with acute and persistent hypocomplementemic glomerulonephritis. The protein has been identified by immunoelectrophoretic analysis, and the amounts present have been determined by direct measurement of specific antigenic determinants present on alpha<sub>2D</sub>.  $\beta_{1A}$ -Globulin, another breakdown product of  $\beta_{1C}$ -globulin, may also be present in vivo in severely hypocomplementemic patients, but its levels are much lower than those of alpha<sub>2D</sub>-globulin.

Alpha<sub>2D</sub>-globulin has been identified by immunoelectrophoretic analysis of fresh EDTA plasma from patients with hypocomplementemic nephritis as an arc in the alpha<sub>2</sub> region that shows a reaction of identity with the arc representing alpha<sub>2D</sub>-globulin produced by aged normal serum.  $\beta_{1A}$ -Globulin was not seen in these patterns.

Measurement of specific antigenic determinants has been carried out in both fresh EDTA plasma and aged serum. In the fresh plasma, the concentration of D antigen, found on both  $\beta_{1\mathrm{C}}$ - and alpha<sub>2D</sub>-globulins, has been related to that of B antigen, found only on  $\beta_{1\mathrm{C}}$  and taken as a measure of the concentration of this protein. In the hypocomplementemic patients, the concentration of D antigen, in comparison to that of B, was greater than in the normal subjects. Similarly, in aged serum, the level of alpha<sub>2D</sub> was greater than would be expected from the amount of  $\beta_{1\mathrm{C}}$  that had been broken down in vitro, measured by the concentration of  $\beta_{1\mathrm{A}}$ .

Calculations indicated that the *in vivo* alpha<sub>2D</sub> level in severely hypocomplementemic patients ranged from 7.5 to 18% of that which would be found in a pool of aged normal serum in which  $\beta_{1C}$  is completely broken down. The levels tended to be lower in less severely hypocomplementemic patients, and none could be detected in normal plasma.

Only small quantities of A and D antigens are detectable in the urine of patients with hypocomplementemic nephritis. The rate of excretion is about equal to that of the normal subject.

The study indicates that the low serum levels of  $\beta_{1C}$ -globulin that may be present over long periods in patients with persistent hypocomplementemic glomerulonephritis can be ascribed, in part, to in vivo breakdown of this pro-

<sup>\*</sup>Submitted for publication July 15, 1966; accepted December 20, 1966.

Supported in part by a grant from United Health Foundations, Inc., and by grants AI-03429 and AI-04472 from the National Institute of Allergy and Infectious Diseases, U. S. Public Health Service.

Presented in part at the annual meeting of the Society for Pediatric Research, Atlantic City, N. J., April 1966.

<sup>†</sup> Address requests for reprints to Dr. Clark D. West, The Children's Hospital Research Foundation, Elland Ave. and Bethesda, Cincinnati, Ohio 45229.

tein as a result of reaction with immune complexes. The contribution of  $\beta_{10}$  deposition on immune complexes and of diminished synthesis to the depressed serum levels cannot be assessed by the present study.

## Introduction

A type of persistent glomerulonephritis has recently been described (1-4) in which the serum  $\beta_{1C}$ -globulin concentration remains very low over periods of many months' duration. The persistently low levels of this protein, normally the most abundant of all of the complement components in serum, could be explained in several ways. The protein may be taken up by immune complexes responsible for the disease, and indeed, immunofluorescent techniques have shown deposition of  $\beta_{10}$ -globulin as well as of IgG in the glomerular capillary walls (3). However, over the long periods of serum depletion observed, the amounts accumulating in this manner would become very great, making continued deposition as the complete explanation unlikely.

Alternatively, the low level could be explained by diminished synthesis of  $\beta_{1C}$ -globulin. Studies employing isotopically labeled  $\beta_{1C}$  have shown that the rate of disappearance of the label in hypocomplementemic subjects is identical to that in normal subjects (5). If this technique truly measures  $\beta_{1C}$  disappearance, it must be concluded that the rate of synthesis of the protein is diminished.

A third possible explanation for the persistently low levels is continued in vivo breakdown of the protein. Recent observations by Müller-Eberhard, Dalmasso, and Calcott (6) concerning the uptake and conversion of  $\beta_{1C}$ -globulin in immune hemolysis indicate that this explanation would be plausible, and the present paper reports studies bearing on this point. Müller-Eberhard and associates (6) found that one  $C'_{4,2a}$  site on a red blood cell is capable of "activating" large numbers of  $\beta_{10}$  molecules. The activated molecules may bind to the red cell membrane or can remain in free solution to become "inactive" C'<sub>3</sub>, designated as  $C'_{3i}$ .  $C'_{3i}$  is, by immunoelectrophoretic analysis, known as  $\beta_{1G}$  and has been found in vitro as the result of reaction of purified  $\beta_{1C}$  with EAC'<sub>1,4,2a</sub> cells.  $B_{1G}$  has not been demonstrated in serum.

The present paper gives evidence for the presence, in vivo, of another breakdown product of  $B_{1C}$ , known as alpha<sub>2D</sub> (7), in patients with

hypocomplementemic nephritis. This breakdown product could originate from  $\beta_{1C}$  directly, or  $\beta_{1G}$ could be an intermediate step. Its presence has been demonstrated by immunoelectrophoretic analysis of fresh plasma from patients with hypocomplementemic nephritis and by measurement of the specific antigenic determinant group associated with the protein. This antigenic determinant was described in a recent study (7) in which three major antigenic determinants were found to be associated with  $\beta_{1C}$ -globulin. Each of the two major breakdown products of  $\beta_{1C}$  was found to have a distinctive determinant, and the third determinant was found only on the parent protein. Thus, one determinant, A, was found on the wellknown breakdown product  $\beta_{1A}$ , whereas the other, D, allowed recognition of the second breakdown product, alpha<sub>2D</sub>. Both of these determinants were also found on the parent protein,  $\beta_{1C}$ . The third determinant, B, was found only on  $\beta_{1C}$  and disappeared as serum aged or was reacted with immune precipitates or zymosan. Since reaction with immune precipitates also results in the formation of the two breakdown products,  $\beta_{1A}$  and alpha<sub>2D</sub>, the presence of these products in hypocomplementemic patients would give evidence that  $\beta_{10}$  participates in an immune reaction in vivo and that breakdown could account, at least in part, for the  $\beta_{1C}$  depletion.

#### Methods

Studies were done on serum and plasma specimens from 45 normal adults, 34 children and adults with hypocomplementemic acute glomerulonephritis, 10 children with hypocomplementemic persistent glomerulonephritis, and 5 children with normocomplementemic persistent glomerulonephritis. By renal biopsy, all of the patients with persistent normocomplementemic glomerulonephritis had focal glomerular lesions, but in none was there a history of anaphylactoid purpura. The renal lesions in these patients as well as in those with persistent hypocomplementemic nephritis have been recently described in detail (4).

Immunoelectrophoretic analysis. The micromethod of Scheidegger (8) was employed. Electrophoresis was carried out in 2% Nobel agar with a barbiturate buffer, pH 8.2, 0.030 M with respect to sodium barbital and 0.011 M with respect to diethylbarbituric acid. For the visualiza-

tion of the alpha<sub>2D</sub> arc in fresh plasma, the diameter of the antigen well was increased to 3.5 mm from the usual diameter of 2 mm.

Measurement of A, B, and D determinants in fresh EDTA plasma and in serum. Antigenic determinants on  $\beta_{1C}$ -globulin were measured by a modification of the immunoelectrophoretic precipitin method described previously (9). Specimens were obtained by introducing 3 ml of freshly drawn blood into a centrifuge tube containing 11.2 mg of EDTA (final EDTA concentration, 0.01 mole per L). The contents were mixed, the tubes centrifuged, plasma was removed, and within a 3-hour period the pipetting necessary for quantitation was completed. Other samples of the same blood were allowed to clot and the serum removed and incubated at 37° for 1 to 2 weeks. For preservation during incubation, the tubes contained enough dried merthiolate or NaF to produce concentrations of 1:10,000 and 0.001 M, respectively. Studies showed that neither preservative affected the A, B, or D antigen titration and that  $\beta_{1D}$  breakdown was as complete under these conditions after 1 week of incubation as after 2 weeks. In lieu of incubation at 37°, some specimens of serum were quantitated after storage for periods of 3 months or more at  $-10^{\circ}$  without preservative. The results were the same as those obtained with incubated specimens. Both serum incubated at 37° and that stored in the deepfreeze are referred to in the text as aged serum.

The method of preparation of antiserum containing high titer of antibody to the A, B, and D determinant groups of  $\beta_{10}$ -globulin was described previously (7). All antiserum was produced in goats. For immunoelectrophoretic determination of A antigen, the antiserum was diluted so that the addition of 44.6  $\mu$ g of  $\beta_{1A}$  to 1 ml of the diluted antiserum removed all but the small amount of antibody to A antigen required to produce the very faint end point precipitin arc (9). In this determination, the supernatants were reacted against  $\beta_{1A}$  contained in a dilute euglobulin precipitate of aged human serum (2). For the determination of A antigen in fresh serum, results were expressed in units (2), and in aged serum in milligrams of  $\beta_{1A}$  per 100 ml.

B determinants were measured only in fresh EDTA plasma. The standard antiserum, before use, was absorbed with aged serum to remove antibody to A and D determinants (7) and was standardized with fresh EDTA plasma from a normal individual. The dilution of antiserum was such that approximately 0.03 ml of the fresh plasma would remove virtually all antibody to B antigen from 1 ml of the standard antiserum to produce an end point. Supernatants were run against normal EDTA plasma diluted 1:4. This plasma was stored in samples at  $-70^{\circ}$ , and, once thawed, was used immediately. That not used was discarded.

The antiserum for measurement of D determinants was adjusted in concentration so that values for D and A determinants, expressed in units per milliliter, in pools of aged serum were the same. For example, in a pool from normal subjects, the concentration of both  $\beta_{1A}$  and alpha<sub>2D</sub> would be 33 U per ml (2). A unit of A determinant in aged serum was equivalent to 45.7  $\mu$ g of  $\beta_{1A}$  (2). Be-

cause alpha<sub>2D</sub> has not been isolated in pure form, the milligram equivalence of a unit of this protein has not been determined. Supernatants were reacted against pooled aged serum diluted 1:4. Samples of the diluted pooled serum were stored at  $-10^{\circ}$  and not refrozen for future use.

Measurement of A and D determinants in urine. A modification of the immunoelectrophoretic precipitin method (9) was used for measurement of A and D determinants in urine. Timed urine collections were made so that results could be expressed as excretion rates. From a given urine collection, samples corresponding in volume to the urine formed over periods of 10 to 120 minutes' duration were added to measured amounts of the standard antisera used for A and D determination. The mixtures were incubated for 30 minutes at 37° and for 2 days at 4°, after which they were centrifuged in the cold. The entire supernatant was then placed in dialysis tubing, and the contents were pervaporated in a current of air with one end of the sac immersed in dilute buffer. The concentrated material was then squeezed from the sac, the sac rinsed, and the material made to the volume that would be used in the immunoelectrophoretic precipitin method for the volume of standard antiserum employed. We then used the concentrated supernatants to develop the immunoelectrophoretic pattern in the same manner as supernatants of serum-antiserum mixtures in the immunoelectrophoretic precipitin method. The end point was determined in the same manner.

Recovery experiments indicated that this method has an inherent inaccuracy. To normal urine containing 0.036 U of D determinant per ml, sufficient aged serum was added to raise the level to 0.086 U per ml. The amount found, however, was 0.061 U per ml. In the case of  $\beta_{1A}$ , none was detectable in the original urine, and we added an amount of aged serum to produce a level of 2.5  $\mu$ g per ml. The amount found was 1.9  $\mu$ g per ml. At these low concentrations, the method therefore underestimates both proteins by 20 to 30%. At the urine concentrations observed, this error is of no consequence.

## Results

A systematic search for *in vivo* breakdown products of  $\beta_{1G}$ —globulin was initiated as a result of the chance observation that serum from hypocomplementemic patients, when subjected to immunoelectrophoretic analysis, showed precipitin arcs representing alpha<sub>2D</sub>—globulin with characteristics which suggested that the level of this protein was greater than would be predicted from the concentrations of  $\beta_{1G}$ — and  $\beta_{1A}$ —globulins. If true, this observation would suggest that this breakdown product circulates *in vivo* in these patients. However, such an observation per se means little, since alpha<sub>2D</sub> forms rapidly from  $\beta_{1G}$  as serum stands at room temperature, and esti-

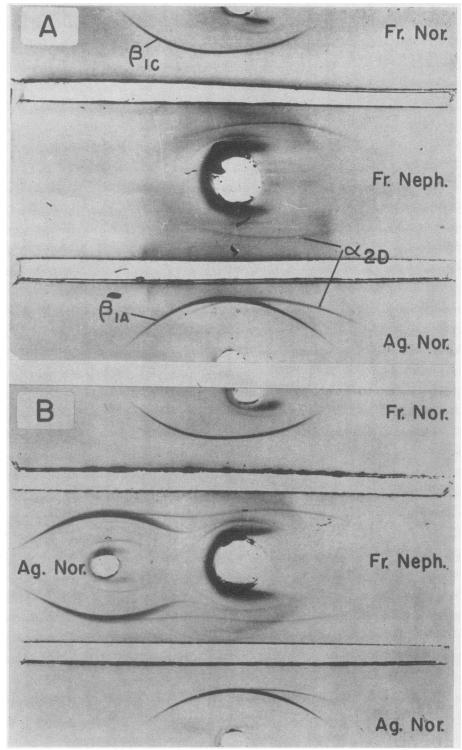


Fig. 1. A) Immunoelectrophoretic patterns of fresh EDTA plasma from normal subject (Fr. Nor.), fresh EDTA plasma from a child with acute glomerulonephritis of recent onset (Fr. Neph.), and aged normal serum (Ag. Nor.). An arc in the alpha<sub>2D</sub> position is produced by the fresh plasma from the nephritic patient but not by the fresh normal plasma.

B) Same as A, with normal aged serum undergoing electrophoresis in the same agar strip as the nephritic plasma. See text for description.

mates of concentration based on the immunoelectrophoretic pattern may be deceiving.

To provide convincing evidence by immunoelectrophoretic analysis of the existence of alpha<sub>2D</sub> in vivo, one must demonstrate the arc representing this protein in fresh plasma that has been drawn with precautions to prevent in vitro  $\beta_{1C}$  breakdown by the addition of EDTA. The demonstration of alpha<sub>2D</sub> in fresh plasma from hypocomplementemic subjects was not possible by the routine immunoelectrophoretic method, apparently because it was present only at low concentrations. However, when the amount of serum subjected to immunoelectrophoresis was increased by enlarging the diameter of the antigen well from the usual 2 mm to 3.5 mm, an arc with the position and configuration of alpha<sub>2D</sub> became readily visible. It is illustrated in Figure 1, A and B.

The antiserum used to produce the patterns in Figure 1 contained antibody to A and D antigens and hence allowed visualization of  $\beta_{1C}$ ,  $\beta_{1A}$ , and alpha<sub>2D</sub> (7). In the bottom panel of Figure 1, A, aged serum from a normal subject has been subjected to immunoelectrophoretic analysis, and a typical strong alpha<sub>2D</sub> arc is seen. No comparable arc is seen in the upper panel in which fresh EDTA plasma from the same normal subject has been analyzed. In the center panel is shown the pattern produced by fresh EDTA plasma obtained from a child shortly after onset of acute glomerulonephritis. Arcs of the same character and mobility as alpha<sub>2D</sub> are present, whereas no  $\beta_{1C}$  or  $B_{1A}$  arc can be identified with certainty.

Figure 1, B, is the same except that aged normal serum has undergone electrophoresis in the same agar strip with the fresh EDTA plasma from the patient with acute glomerulonephritis. junction without spurring of the alpha<sub>2D</sub> arc of the aged normal serum with the arc produced by the patient's plasma identifies the latter as alpha<sub>2D</sub>. This observation constitutes strong evidence that alpha<sub>2D</sub> was present in the circulation in vivo. The arc from the hypocomplementemic serum with which the  $\beta_{1A}$  arc of the aged serum identifies is probably  $\beta_{1C}$ ; in similar immunoelectrophoretic patterns of other patients less hypocomplementemic, a larger arc in this area that reacted with antibody to D antigen and hence interfered with the reaction of identity must of necessity have been  $\beta_{1C}$ -globulin.

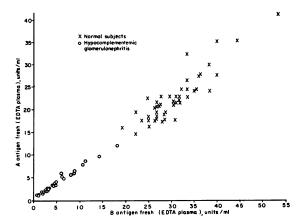


FIG. 2. CONSUMPTION OF ANTIBODY BY FRESH EDTA PLASMA FROM NORMAL AND HYPOCOMPLEMENTEMIC SUBJECTS. Consumption of antibody to the A determinant is plotted against that for B.

With the large antigen well, we observed arcs of the same size and position as the alpha2D arcs in Figure 1, A, on immunoelectrophoretic analysis of fresh EDTA plasma from all hypocomplementemic patients studied. These included six patients with acute glomerulonephritis and five with hypocomplementemic persistent nephritis. Serial immunoelectrophoretic patterns of patients with acute glomerulonephritis, using the large antigen well, showed a rise in  $\beta_{1C}$  levels with convalescence, but alpha2D became increasingly difficult to demonstrate. When the serum  $\beta_{1C}$  levels closely approached normal, no alpha<sub>2D</sub> arc was visible. Under no conditions has alpha<sub>2D</sub> been visualized in freshly drawn EDTA plasma from a normal subject.

To obtain further evidence for the presence of circulating alpha<sub>2D</sub> and to detect the presence of the other  $\beta_{1C}$  breakdown product,  $\beta_{1A}$ , in hypocomplementemic patients, we measured D and A determinants of  $\beta_{1C}$  in fresh plasma and related the results to the concentration of B determinants.

Consumption of antibody to A, B, and D determinants by fresh EDTA plasma. If breakdown products of  $\beta_{1C}$ —globulin do not circulate in significant quantities in normal subjects, consumption of antibody to A, B, and D determinants by normal fresh EDTA plasma would reflect the relative populations of these determinants on the  $\beta_{1C}$  molecule. In presenting the data, we have related the concentrations of both A and D antigens to the concentration of B antigen, since the

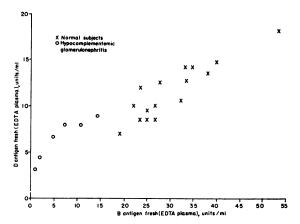


Fig. 3. Consumption of antibody for the D determinant plotted against that for B, as determined in fresh EDTA plasma.

B determinant, found only on intact  $\beta_{1C}$  (7), can be assumed to be a measure of  $\beta_{1C}$ -globulin. In plasma from hypocomplementemic patients, deviation in the relative amounts of B and D antigens from those in normal subjects would constitute further evidence for the presence *in vivo* of alpha<sub>2D</sub>. Deviation in the relation of B to A would give evidence for B<sub>1A</sub>.

In Figure 2, values for antibody consumption by A determinants are plotted vs. those for B, both expressed in units per milliliter of fresh EDTA plasma. The values are from 38 normal adults and from 30 patients with hypocomplementemic glomerulonephritis. Seven of the latter patients had persistent glomerulonephritis, and the remainder had acute glomerulonephritis. Considerable scatter can be seen in the data for the normal subjects. This would not appear to be the result of errors in the method, since determinations made on specimens obtained at different times from four of the subjects gave values that differed little. It is therefore tempting to ascribe this scatter to slight differences among individuals in the structure of their  $\beta_{10}$ -globulin resulting in varying ability to bind antibody to A and B antigens. Despite the scatter it seems reasonable to conclude from this Figure that the relationship of A to B determinants in the hypocomplementemic patients differs little from that in normal subjects.

In contrast to the A determinants, the consumption of antibody to D determinants in relation to that of B is different in the plasma of the hypocomplementemic as compared to the normal sub-

jects. In Figure 3, this relationship is plotted for 16 normal subjects and 6 hypocomplementemic patients (3 with acute and 3 with persistent hypocomplementemic nephritis). The concentration of D antigen in the hypocomplementemic patients is greater than would be expected for the amount of B antigen present. Again, there is some scatter of the points for the normal subjects.

A more sensitive assessment of the relation of A and D to B antigens in hypocomplementemic patients is afforded by plotting the data as ratios. In Figure 4, the ratios of A:B and D:B are plotted against the concentration of B determinant. In the normal subjects, both ratios are relatively constant at all concentrations of  $\beta_{1C}$ . However, in hypocomplementemic patients, both ratios progressively increase as the concentration of B determinant falls. The relatively greater increase in D:B is readily apparent; A:B approximately doubled at low concentrations of B determinant, whereas D:B increased more than tenfold.

A and D determinants of aged serum. To obtain further evidence for the presence of alpha<sub>2D</sub> in vivo, we have measured A and D antigens in serum aged by incubation at 37° for 1 week or longer or stored for long periods at  $-10^{\circ}$ . Since we found  $\beta_{1C}$  to be completely broken down under these conditions, we could relate directly the D antigen concentration of these specimens to the concentration of alpha<sub>2D</sub>, here expressed in units

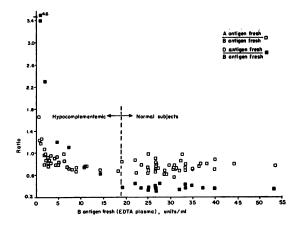


Fig. 4. Demonstration of "excess" A and D antigens in hypocomplementemic plasma by expressing their concentrations in terms of ratios to B antigen. The ratios are plotted on the ordinate, and the B antigen, denoting the concentration of  $\beta_{10}$ , is plotted on the abscissa.

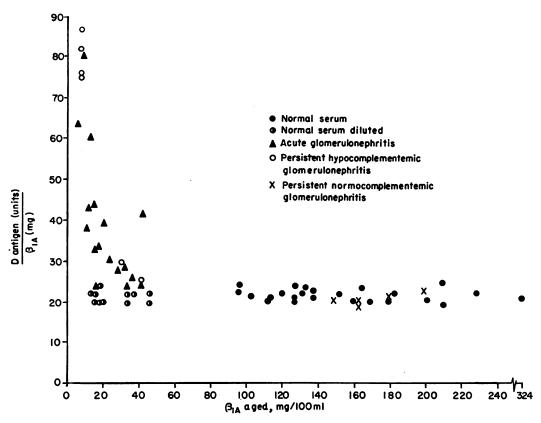


Fig. 5. Alpha<sub>2D</sub> content of aged serum related **TO**  $\beta_{1A}$  content. The ratio of alpha<sub>2D</sub> to  $\beta_{1A}$  is plotted on the ordinate and the  $\beta_{1A}$  concentration on the abscissa.

per 100 ml, and A antigen to the concentration of  $\beta_{1A}$ , here expressed in milligrams per 100 ml. The results are plotted in Figure 5, with the  $\beta_{1A}$  concentration on the abscissa and the ratio of alpha<sub>2D</sub> to  $\beta_{1A}$  on the ordinate.

In the normal subjects, indicated by solid circles, the alpha<sub>2D</sub> per milligram of  $\beta_{1A}$  was constant over a wide range of  $\beta_{1A}$  levels. The values averaged 21.4 U of alpha<sub>2D</sub> per mg of  $\beta_{1A}$ . In the hypocomplementemic subjects with both acute and persistent glomerulonephritis, the ratio was greatly increased. In sera from the most markedly hypocomplementemic patients, alpha<sub>2D</sub> was present in amounts four times as great as would be predicted from the amount of  $\beta_{1A}$  present. Evidence that this was not the result of a systematic error of measurement at low antigen levels is the fact that values for diluted normal serum, designated by the half-shaded circles, are in the same range as those for undiluted normal serum.

The disproportion between A and D concentra-

tions in hypocomplementemic serum would seem most logically ascribed to the presence of alpha<sub>2D</sub> in vivo. Thus, the alpha<sub>2D</sub> measured in aged serum from hypocomplementemic patients would not only be that formed in vitro by the breakdown of  $\beta_{1C}$  but also the alpha<sub>2D</sub> produced by in vivo breakdown of B<sub>1C</sub> and present at the time the blood was drawn. As an alternative explanation, it might be proposed that the disproportion is the result of a change in the character of  $\beta_{1C}$ —globulin with hypocomplementemia such that it breaks down in vitro to generate large amounts of alpha<sub>2D</sub>. The results of immunoelectrophoretic analysis (Figure 1) make this explanation unlikely.

In Figure 5 are also shown the values for five patients with normocomplementemic persistent nephritis, indicated by crosses. The values are in the same range as for normal subjects. Blood specimens for these determinations were obtained when many manifestations of nephritis were present, 10 to 70 days after the presumed onset. In

Subject	Age	Diag- nosis†	Dura- tion of disease	Serum levels (aged)			Excretion			
				$\beta_{1A}$	Alpha <sub>2D</sub>	Alpha <sub>2D</sub> / β <sub>1A</sub>	Actual urine flow	Protein	$oldsymbol{eta_{1A}}$	${ m Alpha_{2D}}$
	years		days	mg/100 ml	U/ml	U/mg	ml/min	mg/hour	μg/hour	U/hour
P.T.	8	AGN	7 21	5.2 9.1	3.3 7.3	63.5 80.2	$0.81 \\ 0.68$	19.4 12.2	<34 <23	0.75 0.5
Z.T.	9	AGN	8	41	17	41.4	0.32	10.9	< 23	< 0.5
R.W.	8	PHGN	660	7.5	5.7	76.0	0.28	0	< 34	0.5
ormal adult				90-220‡	20-50‡	21.4§	0.35	0	<23	0.75

TABLE I Excretion of A and D determinants\* of  $\beta_{1C}$ -globulin in the urine of normal and hypocomplementemic subjects

\* Expressed as  $\beta_{1A}$  and alpha<sub>2D</sub>-globulins, respectively.

§ See Figure 5.

all five patients, biopsies gave evidence of focal inflammatory reactions in the glomeruli (4). The normal ratios of alpha<sub>2D</sub> to  $\beta_{1A}$  in these patients indicate that if an immune reaction involving complement is occurring, it has no discernible effect on the serum content of either  $\beta_{1A}$  or alpha<sub>2D</sub>.

In vivo concentration of alpha<sub>2D</sub> in plasma. The plasma concentration of alpha<sub>2D</sub> in vivo can be calculated from the measurements of B and D determinants in fresh plasma and of A and D determinants in aged serum. If we assume that alpha<sub>2D</sub> is not present in vivo in normal subjects, the data allow an estimate of the amount of D determinant contributed by  $\beta_{1C}$  in hypocomplementemic plasma or serum. Subtracting this value from the total alpha<sub>2D</sub> concentration gives the order of magnitude of the alpha<sub>2D</sub> concentration in vivo.

From Figure 5 it can be seen that in aged serum the  $\beta_{1A}$  concentration in milligrams per 100 ml can be multiplied by 21.3 to estimate the units of alpha<sub>2D</sub> per 100 ml derived from  $\beta_{1C}$  breakdown during aging. In hypocomplementemic serum, the amount in excess of this estimate would be preformed alpha<sub>2D</sub>. Although there is considerable scatter of the data, there is a definite trend for the preformed alpha<sub>2D</sub> present in vivo to be in higher concentration in specimens with the lowest  $\beta_{1A}$  concentrations. For example, at  $\beta_{1A}$  levels between 5 and 10 mg per 100 ml, preformed alpha<sub>2D</sub> concentration averaged 4.2 U per ml (six patients), whereas between 30 and 45 mg per 100 ml the average was 2.1 U per ml (six patients). As suggested by the results of immunoelectrophoretic analysis then, the concentration of this breakdown product in hypocomplementenic patients tends to vary inversely with the level of  $\beta_{1C}$ .

A similar inverse relationship probably also obtains in fresh plasma. However, it could not be demonstrated by calculations with the data for D and B antigens given in Figures 3 and 4, probably because of the relatively small number of specimens available.

The data become more understandable if the concentration of alpha<sub>2D</sub> in hypocomplementemic serum can be related to some readily available standard. Since this protein is not detectable in fresh plasma from normal subjects, the most appropriate comparison would be with aged normal serum. The calculation given above indicates that in the most hypocomplementemic patients, the alpha<sub>2D</sub> level *in vivo* would range from 7.5 to 18% of the level in a pool of aged serum from normal subjects.

Urinary excretion of  $\beta_{1A}$  and alpha<sub>2D</sub>. Accumulation of alpha<sub>2D</sub> in the circulation of hypocomplementemic patients without evidence of in vivo  $\beta_{1A}$  accumulation might be the result of different rates of urinary excretion of the two products. Measurement was therefore made of the excretion rates of A and D antigens in the urine of hypocomplementemic patients. The results, expressed in terms of  $\beta_{1A}$  and alpha<sub>2D</sub>, are given in Table I for two patients with acute glomerulonephritis, one with persistent hypocomplementemic nephritis, and a normal subject. The three patients with nephritis had amounts of alpha<sub>2D</sub> in their aged serum considerably in excess of the normal value of 21.4 U per mg of  $\beta_{1A}$  (Figure 5). None had

<sup>†</sup> AGN = acute glomerulonephritis; PHGN = persistent hypocomplementemic glomerulonephritis. ‡ Ranges for  $\beta_{1A}$  in normal subjects determined previously (4), for alpha<sub>2D</sub>, in the present study.

marked proteinuria, even though in some the  $\beta_{1A}$  levels were extremely low.

In all instances, excretion of  $\beta_{1A}$  was too low to allow accurate measurement. Values were less than 34  $\mu$ g per hour. In fact, there is reason to believe that there is virtually no excretion of this determinant, insofar as addition of increasing amounts of urine to the standard antiserum resulted in no detectable depletion of antibody, as evidenced by the lack of change in the intensity and character of the precipitin arc produced by the supernatants. The observation suggests that the urine was completely devoid of A determinants.

Alpha<sub>2D</sub> was present in measurable amounts in the urine of the normal subject and of two of the patients with nephritis. The normal adult had a rate of excretion equal to or greater than that of the patients with nephritis. In the first specimen from P.T., the rate of excretion of alpha<sub>2D</sub> of 0.75 U per hour gives a renal clearance of 0.23 ml of plasma per hour, assuming his alpha<sub>2D</sub> concentration *in vivo* was the same as that of his aged serum.

Although these excretion values may be erroneously low by a factor of 25 to 30% due to methodological error, it is nevertheless apparent that the excretion rates of  $\beta_{1C}$ —globulin and its breakdown products are almost negligible and would not account for the reduced serum levels.

## Discussion

Two lines of evidence, immunoelectrophoretic analysis and titration of specific antigenic determinants, indicate that a small but significant amount of alpha<sub>2D</sub> is present in vivo in hypocomplementemic patients, presumably as a breakdown product of  $\beta_{1C}$ -globulin. There was no experimental evidence of its presence in normal subjects. In contrast, the data indicate that very little  $\beta_{1A}$  is present in hypocomplementemic patients. The only evidence for the presence of  $\beta_{1A}$ was the slightly increased ratio of A to B antigen in fresh plasma (Figure 4). This result is contrary to previous reports by Morse, Müller-Eberhard, and Kunkel (10) and by Lachmann (11). These workers demonstrated the presence of circulating  $\beta_{1A}$ -globulin by immunoelectrophoretic analysis of fresh serum or fresh EDTA plasma from patients with lupus nephritis. Their observations indicate a concentration of  $\beta_{1A}$  far greater than observed in the present study. The reason for the different results is not known. Perhaps in lupus nephritis the reaction of  $\beta_{1C}$ —globulin or the disposition of its breakdown products is quantitatively different from that in other types of hypocomplementemic nephritis.

The readily demonstrable levels of alpha<sub>2D</sub> in the face of a very low concentration of  $\beta_{1A}$ , as found in the present study, cannot be explained at present. Studies by others (3) indicate that  $\beta_{1C}$  is not excreted in the urine, and the more detailed observations of the present paper indicate that neither  $\beta_{1A}$  nor alpha<sub>2D</sub> is excreted in significant amounts. The discrepancy in the *in vivo* levels of the two proteins will have to be explained by a difference in their rates of clearance by metabolic processes.

Definite evidence for in vivo presence of  $\beta_{1G}$ -globulin ( $C'_{3i}$ ) (6, 7) was not found. The small amount of A determinant, attributed above to  $\beta_{1A}$ , could conceivably be contributed by A determinants on  $\beta_{1G}$ . In this case, a moiety of the "excess" of D determinants should also be assigned to  $\beta_{1G}$ . It should, however, be noted that  $\beta_{1G}$  has only been produced from purified  $\beta_{1G}$  in the absence of whole serum; in serum, the condition that produces this transformation presumably has as its end products  $\beta_{1A}$  and alpha<sub>2D</sub>.

Although alpha<sub>2D</sub> was the only breakdown product unequivocally found in the hypocomplementemic patients, its presence constitutes further evidence for the current concept that the reducd complement level in nephritis is the result of participation of complement in an *in vivo* immune reaction. One of the mechanisms by which the third component of this complex,  $\beta_{1C}$ , can remain reduced over long periods must therefore be *in vivo* breakdown. The roles of  $\beta_{1C}$  deposition and of diminished synthesis in keeping concentrations low cannot be defined by the present study.

The present observations coincide with the investigations of Müller-Eberhard and associates (6), who used radiolabeled  $\beta_{1C}$  in detailed studies

 $<sup>^{1}</sup>$  In vivo breakdown of  $\beta_{1C}$  and persistence of breakdown products in the circulation could introduce error in the isotopic measurement of  $\beta_{1C}$  disappearance. If the portion of the molecule that yielded alpha<sub>2D</sub> were isotopically labeled, the breakdown product would contribute counts and the half-life calculated for  $\beta_{1C}$  would be erroneously long.

of the uptake and conversion of this complement component during immune hemolysis. Attachment of  $C'_3$  ( $\beta_{1C}$ ) to the red cell membrane was found to be the result of the reaction of  $\beta_{1C}$  with C'<sub>1,4,2a</sub> or C'<sub>4,2a</sub> sites on the red cell surface. Because one C'<sub>4,2a</sub> group could effect the binding of several hundred  $\beta_{1C}$  molecules, it was impossible to assume that the attachment was to the  $C'_{4,2a}$ group. The point applicable to the present study was that, in addition to bound  $C'_3$ , the reaction also caused conversion of large quantities of C'3 to inactive  $C'_3$ , designated  $C'_{3i}$ , or  $\beta_{1G}$ . In fact, C'<sub>4,2a</sub> groups in free solution were also capable of effecting conversion to C'3i, and here none of the product was bound to cell membranes, even though red cells were suspended in the reaction The results required those authors to postulate that C'4,2a groups are capable of converting, by enzymatic action, large numbers of C'3 molecules to an "activated" form. This form is unstable and can either bind to cell membranes or convert to C'3i. The half-life of activated C'3 appears to be very short.

If we assume that  $C'_{4,2a}$  groups are present in the glomerular capillary walls or on circulating complexes, the above observation can be applied to the events in hypocomplementemic nephritis. C'<sub>3</sub>, present in circulating plasma, is activated and, at least initially in the disease, is bound to surrounding glomerular structures or to complex. As the reaction proceeds, available receptors for attachment of C'3 become saturated and the majority of the molecules convert to C'3i. C'3i is presumably in turn broken down by serum factors to  $\beta_{1A}$  and alpha<sub>2D</sub>. The alpha<sub>2D</sub> produced from C'31 breakdown appears to circulate for a time, whereas  $\beta_{1A}$  must be quickly removed from the circulation. It should be noted that the enzymatic action of C'2a groups would allow relatively few C'<sub>4,2a</sub> sites to be effective in maintaining a low circulating level of  $\beta_{1C}$ . It is possible that the rate of inactivation and breakdown of  $\beta_{1C}$  and hence its serum level is a function of the number of these sites available to circulating plasma.

For a time it was considered that the failure to find significant amounts of  $\beta_{1A}$  in the circulation as compared to the relatively large amounts of alpha<sub>2D</sub> might be explained by attachment of  $\beta_{1A}$  to red cell membranes. If activated  $C'_3$  binds with avidity to membranes, even, as shown by Müller-

Eberhard and associates (6), in the absence of antibody, it would seem plausible that the erythrocytes of hypocomplementemic patients, abundant in the glomerular capillary walls in sites where  $C'_3$  is activated, might have attached to them large quantities of  $C'_3$ . However, our initial attempts to demonstrate attached  $\beta_{1A}$  by hemagglutination techniques using antibody directed at A determinants have not been successful. Agglutination of cells from hypocomplementemic patients by such antiserum has been no greater than that of cells from normal subjects.

## References

- West, C. D., A. J. McAdams, J. D. Northway, J. M. McConville, N. C. Davis, and N. H. Hollands. Serum levels of beta-1C globulin in the nephritides: differentiation of two types of chronic nephritis (abstract). J. Pediat. 1964, 65, 1022.
- West, C. D., J. D. Northway, and N. C. Davis. Serum levels of beta<sub>10</sub> globulin, a complement component, in the nephritides, lipoid nephrosis, and other conditions. J. clin. Invest. 1964, 43, 1507.
- Gotoff, S. P., F. X. Fellers, G. F. Vawter, C. A. Janeway, and F. S. Rosen. The beta<sub>10</sub> globulin in childhood nephrotic syndrome. Laboratory diagnosis of progressive glomerulonephritis. New Engl. J. Med. 1965, 273, 524.
- West, C. D., A. J. McAdams, J. M. McConville, N. C. Davis, and N. H. Holland. Hypocomplementemic and normocomplementemic persistent (chronic) glomerulonephritis; clinical and pathologic characteristics. J. Pediat. 1965, 67, 1089.
- Alper, C. A., A. S. Levin, and F. S. Rosen. Beta-1C-globulin: metabolism in glomerulonephritis. Science 1966, 153, 180.
- Müller-Eberhard, H. J., A. P. Dalmasso, and M. A. Calcott. The reaction mechanism of β<sub>10</sub>-globulin (C'3) in immune hemolysis. J. exp. Med. 1966, 123, 33.
- West, C. D., N. C. Davis, J. Forristal, J. Herbst, and R. Spitzer. Antigenic determinants of human β<sub>10</sub>- and β<sub>10</sub>-globulins. J. Immunol. 1966, 96, 650.
- 8. Scheidegger, J. J. Une micro-méthode de l'immunoélectrophorèse. Int. Arch. Allergy 1955, 7, 103.
- West, C. D., V. Hinrichs, and N. H. Hinkle. Quantitative determination of the serum globulins beta<sub>2A</sub> and beta<sub>2M</sub> by immunoelectrophoretic analysis. J. Lab. clin. Med. 1961, 58, 137.
- Morse, J. H., H. J. Müller-Eberhard, and H. G. Kunkel. Antinuclear factors and serum complement in systemic lupus erythematosus. Bull. N. Y. Acad. Med. 1962, 38, 641.
- Lachmann, P. J. Complement in Clinical Aspects of Immunology, P. G. H. Gell and R. R. A. Coombs, Eds. Philadelphia, F. A. Davis, 1962, p. 260.