Endogenous Carbon Monoxide Production in Patients with Hemolytic Anemia *

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It is desirable in many studies to be able to rapidly measure rates of erythrocyte and hemoglobin destruction. Labeling techniques have been utilized for this purpose but have the disadvantage of requiring several days to weeks to obtain the desired data plus the hazards of radioactive isotopes. Estimations of hemoglobin turnover from measurements of fecal bile pigment or serum bilirubin concentrations contain large sources of error (1). Studies in which bile pigment is collected via bile duct fistulas have been of value in animal experiments (2) but are not usually possible in man, and subjects who have bile fistulas cannot be called normal. At the present time it appears that many in vivo investigations of red cell survival in patients with hemolytic disease are limited by the lack of a precise rapid method for detecting changes in the rate of red cell destruction.

It has been shown previously (3–9) that CO is an in vivo catabolic by-product of hemoglobin and that it probably originates from the heme α-methylene bridge carbon atom. Data obtained in experiments where CO production was measured after intravenous injections of suspensions of damaged erythrocytes or solutions of hemoglobin have indicated that the molar yield of CO produced to heme catabolized is approximately unity in normal man (6) and anesthetized dogs (9). Apparently, therefore, the measurement of CO production is useful in studying hemoglobin and red cell turnover and can be utilized in studies of patients with increased erythrocyte destruction. The quantitation of erythrocyte and hemoglobin catabolism from CO production measurements has the advantages that it is rapidly accomplished, does not require isotopes, and can be repeated.

Early workers (4, 10) attempted to estimate red cell survival with measurements of the venous blood carboxyhemoglobin per cent saturation ([COHb]) in patients with hemolytic anemia; this approach seems inadequate in view of the uncertainties of the relationship of [COHb] to the actual rate of CO production in these patients (11). The development of a method of estimating the rate of CO production (Vco) based on the measurement of the rate of increase in CO body stores under conditions where CO excretion from the body is prevented by having the subject rebreathe in a closed system has apparently circumvented this difficulty (5).

In the present report we describe a modification of the rebreathing system that has allowed us to measure Vco in seriously ill and debilitated patients. With this system Vco has been determined in seven patients with hemolytic anemia. The results show that the measurement can be used to quantitate erythrocyte and hemoglobin destruction in these patients and that there is a fraction of the Vco that apparently cannot be explained on the basis of catabolism of circulating hemoglobin.

* Submitted for publication May 10, 1965; accepted December 9, 1965.


Supported in part by a grant from the Life Insurance Medical Research Fund and grant 3 M 01 FR-40-05 from the Clinical Research Center, National Institutes of Health, Bethesda, Md.

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460

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**Methods**

**CO production measurements.** The rate of CO production was determined by the rebreathing method described in detail in a previous publication (5), except that the apparatus used was modified so that the subject breathed in a plastic hood rather than through a mouthpiece. The studies were performed with the $O_2$ tension in the hood and rebreathing system adjusted to approximately 150 mm Hg. The first venous blood sample was drawn at least 15 minutes after the start of rebreathing; additional samples were taken every 30 minutes for a total of 2 hours. After this the body CO dilution was determined by adding 10 ml of CO to the rebreathing system and measuring the resultant increase in [COHb]. Blood [COHb] was measured with an infrared CO method (12), the rate of increase in [COHb] was determined graphically, and the Vco was calculated from this value and the CO blood volume or CO dilution in the body (5). The error in the Vco measurement is ±3.1 amoles per hour (SD) (5). This method assumes rapid mixing and equilibration of endogenously produced CO and CO added at the end of each experiment in the blood and extravascular CO stores, which include CO bound in muscle, liver, bone marrow, and possibly other organs (5). The available data regarding equilibration and mixing of the body CO stores have been reviewed previously (5). These data and unpublished data obtained in our laboratory (13) indicate that for the purposes of the Vco measurement the body CO stores can be assumed to be a single rapidly equilibrating “pool” without the introduction of significant error. The hemolytic patients studied in the present investigation presumably had more bone marrow than normal man, and it seems likely that the portion of the body CO stores present in this tissue would be greater. The marrow is, however, a vascular structure, and it would be expected that marrow CO, like other extravascular CO stores, would equilibrate rapidly with intravascular CO and that this larger marrow CO pool would not cause significant error in the Vco measurement.

**The rebreathing system.** We have modified the rebreathing system used in our previous investigations so that the subject does not have to breathe through a mouthpiece and wear a noseclip but rather breathes in a plastic hood that fits over the head and is sealed about the neck with a rubber diaphragm (Figure 1). A pump constantly circulates the gas in the hood through a CO$_2$ absorber and cooler. The hood is made of 4 cm plastic and has the shape of a cylinder closed at one end. The outside diameter of the cylinder is 30 cm, and it is 23 cm in length. Five gas taps lead into the hood. Two of these are 3 cm in diameter and are used in pumping gas to and from the CO$_2$ absorber and cooler. A tap 1 cm in diameter leads to the oxygen meter. The fourth tap, 4 cm in diameter, is connects to a rubber bag that is used as a gas reservoir. The fifth tap is connected to an oxygen demand valve adjusted so that oxygen is added to the system at the end of each inspiration. The patient's neck is greased with silicone grease before fitting of the diaphragm to improve the seal between the neck and hood. We have diaphragms for varying neck sizes, and it has been possible to achieve a tight seal in all patients without neck constriction. The air pump used in the system has a maximal flow through our system of 60 L per minute. The canister contains approximately 800 ml of CO$_2$ absorbant and is placed in an insulated container filled with ice. The air flow through the CO$_2$ absorber is sufficient to keep the CO$_2$ in the system below 0.5%.

The apparatus as described has a total gas volume of approximately 7 L (with head in hood and CO$_2$ canister filled with Baralyme). The volume of the apparatus plus

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**Figure 1. Schematic representation of the rebreathing system.** The hood, diaphragm, and pumping system are shown. RB, rubber bag; CO$_2$ ABS, carbon dioxide absorber; P, pump; O$_2$ M, oxygen meter; O$_2$ DV, oxygen demand valve.
During most of the studies we checked the system for leaks by injecting small quantities of neon into the gas in the rebreathing system and monitoring the neon concentration in this gas with a gas chromatograph. In all of the studies the neon concentration decreased slightly during the first 30 minutes and then remained constant during the remainder of the study. The initial decrease in neon concentration presumably resulted from wash-in of neon into the lungs and solubility in body tissues rather than leakage out of the system. Although the system proved to be free of leaks, the Vco measurement is not highly sensitive to leaks. A leak of 1 L out of the system in 2 hours would cause an error of less than 0.5 μmole per hour in measured Vco in a normal subject; leaks into the system would result in proportionately smaller error, since the CO concentration in room air is usually less than in the equilibrated air in the rebreathing system.

Simulated CO production experiments. We performed two experiments designed to measure the precision of the rebreathing method and determine if the precision of ± 3.1 μmoles per hour (SD) (0.07 ml per hour) could be achieved when Vco was elevated. A normal amount of CO was periodically added to the body stores by injecting CO into the rebreathing gas. The rate of CO injection was compared with the measured Vco calculated from the rate of increase in [COHb]. Data from an experiment performed on a normal male subject are shown in Figure 2. The patient initiated breathing in the closed system. A venous blood sample was drawn, and 1.82 ml (standard temperature and pressure, dry) of CO was added to the rebreathing system. This sequence was repeated every 30 minutes so that 3.64 ml of CO was added per hour to the body stores. Measured Vco was 4.00 ml per hour. The difference between the simulated Vco and measured Vco was 0.36 ml per hour and is entirely explainable on the basis of normal endogenous CO production. The Vco in normal young male subjects averages 0.42 (SD ± 0.07) ml per hour (5). The second experiment was performed on an anesthetized dog breathing in a closed system.

**FIG. 2. SIMULATED CO PRODUCTION IN A NORMAL MALE SUBJECT.** Endogenous CO production was "simulated" by adding CO to the rebreathing system every 30 minutes for 2 hours. The "rate of CO production" was determined in the usual manner by measuring the rate of increase in blood carboxyhemoglobin per cent saturation and calculating the rate of increase in the body CO stores. In this experiment 3.64 ml CO per hour was added. The measured rate of CO increase in the body was 4.00 ml per hour. The difference is entirely explainable by endogenous CO production.

Lung volume is approximately 12 L. In designing this system we kept the volume as small as possible to minimize CO loss from the body into gas, which occurs as [COHb] increases during the procedure. The CO loss that could occur in our system from this process is less than 0.02 μmole per hour in a normal subject.

<table>
<thead>
<tr>
<th>Patient</th>
<th>Age</th>
<th>Sex</th>
<th>Height</th>
<th>Weight</th>
<th>Diagnosis</th>
<th>Total body hemoglobin</th>
<th>$^{14} \text{C}t$</th>
<th>$K^*$</th>
</tr>
</thead>
<tbody>
<tr>
<td>BS</td>
<td>16</td>
<td>F</td>
<td>59</td>
<td>88</td>
<td>Autoimmune</td>
<td>196</td>
<td>4.7</td>
<td>13.41</td>
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<tr>
<td>HM</td>
<td>22</td>
<td>M</td>
<td>67</td>
<td>170</td>
<td>S-S</td>
<td>518</td>
<td>6.8</td>
<td>8.99</td>
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<tr>
<td>GC</td>
<td>22</td>
<td>F</td>
<td>61</td>
<td>109</td>
<td>S-S</td>
<td>235</td>
<td>7.0</td>
<td>8.61</td>
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<tr>
<td>MC</td>
<td>62</td>
<td>F</td>
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<td>144</td>
<td>Autoimmune</td>
<td>349</td>
<td>18</td>
<td>2.55</td>
</tr>
<tr>
<td>FS</td>
<td>25</td>
<td>M</td>
<td>72</td>
<td>137</td>
<td>Hodgkin's disease</td>
<td>467</td>
<td>22</td>
<td>1.84</td>
</tr>
<tr>
<td>SF</td>
<td>26</td>
<td>M</td>
<td>72</td>
<td>141</td>
<td>Hodgkin's disease</td>
<td>306</td>
<td>15</td>
<td>3.31</td>
</tr>
<tr>
<td>CJ</td>
<td>21</td>
<td>M</td>
<td>70</td>
<td>159</td>
<td>S-S</td>
<td>621</td>
<td>16</td>
<td>3.13</td>
</tr>
<tr>
<td>IH</td>
<td>23</td>
<td>M</td>
<td>68</td>
<td>131</td>
<td>S-S</td>
<td>430</td>
<td>7</td>
<td>8.61</td>
</tr>
</tbody>
</table>

* $K^*$, the measured decay in blood $^{14} \text{C}$ minus correction for elution of 1.29% per day.
+ S-S, sickle cell anemia.
+ In these patients we measured serum folic acid (17) and urine formiminoglutamic and urocanic acids after a histidine load (18). Normal values were found in GH, FS, and CJ. IH had slightly decreased serum folic acid level and slightly increased urine formiminoglutamic and urocanic acid levels.
ENDOGENOUS CARBON MONOXIDE

Table II
CO production and hemoglobin destruction data*

<table>
<thead>
<tr>
<th>Patient</th>
<th>Date</th>
<th>Rate increase [COHb]</th>
<th>[COHb]</th>
<th>Vco</th>
<th>Vheme</th>
<th>Vco/Vheme</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>%/hr</td>
<td>% sat</td>
<td>μmoles/hr</td>
<td>μmoles/hr</td>
<td>μmoles/hr/μmole/hr</td>
</tr>
<tr>
<td>BS</td>
<td>10-23-61</td>
<td>0.56</td>
<td>2.62</td>
<td>72</td>
<td>65</td>
<td>1.10</td>
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<tr>
<td>HM</td>
<td>4-19-62</td>
<td>0.36</td>
<td>2.46</td>
<td>115</td>
<td>114</td>
<td>0.99</td>
</tr>
<tr>
<td></td>
<td>4-20-62</td>
<td>0.36</td>
<td>2.33</td>
<td>109</td>
<td></td>
<td></td>
</tr>
<tr>
<td>GH</td>
<td>7-9-62</td>
<td>0.42</td>
<td>2.24</td>
<td>62</td>
<td>49</td>
<td>1.32</td>
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<td></td>
<td>7-11-62</td>
<td>0.51</td>
<td>1.71</td>
<td>68</td>
<td></td>
<td></td>
</tr>
<tr>
<td>MC</td>
<td>1-9-62</td>
<td>0.15</td>
<td>1.34</td>
<td>31</td>
<td>22</td>
<td>1.41</td>
</tr>
<tr>
<td>FS₁</td>
<td>5-14-62</td>
<td>0.11</td>
<td>0.81</td>
<td>32</td>
<td>21</td>
<td>1.62</td>
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<td>5-16-62</td>
<td>0.13</td>
<td>0.77</td>
<td>35</td>
<td></td>
<td></td>
</tr>
<tr>
<td>FS₂</td>
<td>10-20-62</td>
<td>0.26</td>
<td>2.47</td>
<td>48</td>
<td>25</td>
<td>1.92</td>
</tr>
<tr>
<td>GJ</td>
<td>9-10-62</td>
<td>0.16</td>
<td>1.46</td>
<td>62</td>
<td>47</td>
<td>1.33</td>
</tr>
<tr>
<td></td>
<td>9-12-62</td>
<td>0.18</td>
<td>1.85</td>
<td>64</td>
<td></td>
<td></td>
</tr>
<tr>
<td>IB</td>
<td>12-11-62</td>
<td>0.53</td>
<td>2.10</td>
<td>138</td>
<td>91</td>
<td>1.54</td>
</tr>
<tr>
<td></td>
<td>12-13-62</td>
<td>0.58</td>
<td>2.12</td>
<td>143</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Average</td>
<td>Normal</td>
<td>0.05</td>
<td>0.88</td>
<td>18.7</td>
<td>14.7</td>
<td>1.40 ± (SE) 0.10</td>
</tr>
</tbody>
</table>

* [COHb], venous blood carboxyhemoglobin; Vco, rate of CO production; Vheme, rate of blood heme catabolism.

Vco was again 3.64 ml per hour, and measured Vco was 3.70 ml per hour. The average endogenous Vco found in anesthetized dogs is 0.18 (SD ± 0.08) ml per hour (9). These experimental results suggest that the precision of the measurement is similar with normal and markedly elevated Vco.

Measurement of erythrocyte and hemoglobin destruction rates. Erythrocyte survival studies were performed with radiochromate by the method of Read (14) except that acid-citrate-dextrose anticoagulant was used. The method was then nearly identical to that of Cline and Berlin (15). These data were expressed as a half-time, t₁, the time at which blood radioactivity reached 50% of the initial value. From this we calculated the time constant, K, using the following equation which assumes that erythrocyte destruction was random: N = N₀ e⁻Kt. N₀ is the initial red cell radioactivity at zero time, N the radioactivity at time t₁, and K the time constant in per cent per day. K was corrected for elution, assuming this process is constant and exponential and equal to 1.29% per day (15) giving a corrected time constant K'. The rate of blood heme destruction was calculated by multiplying K' and the total body hemoglobin calculated from the CO dilution measurements (16). The results are expressed as micromoles heme destroyed per hour.

The patients used in this study had documented hemolytic diseases. Vital statistics and diagnoses are listed in Table I. All of the patients were thought to be clinically in a stable or steady state. Five of the seven patients were brought into the hospital specifically for this study. Drug regimens were not changed during the study period. None of the patients had been transfused for 3 months before the study. One of the patients (FS) was studied completely at two different stages of his disease (May and November of 1962). Three of the patients (BS, FS, and CJ), were smokers but were not allowed to smoke for 2 days before the Vco measurements; the remainder of the patients denied smoking.

Results

Total body hemoglobin, ⁵¹Cr t₁, and calculated K' values are listed for each patient in Table I. The t₁ of ⁵¹Cr varied from 4.7 to 22 days; K' ranged from 1.84 to 13.41% per day. Calculated values of Vheme are listed in Table II. These were found to range from 21 to 114 μmoles per hour, or 0.36 to 1.94 g hemoglobin per hour. The rates of blood hemoglobin catabolism were, therefore, approximately 1 ½ to 8 times the normal value of 14.7 μmoles per hour.

Table II shows the data on CO production in these patients. The venous blood [COHb] of the first sample drawn in each study was greater than normal in seven of the eight studies, ranging from 0.77 to 2.62%. The average [COHb] in patients without hemolytic disease is 0.88 ± 0.11% (SD) (12). Vco was elevated compared to normal in all of the patients. The range of values was 0.70
to 3.18 ml per hour or 31 to 143 μmoles per hour. Normal Vco is 18.7 ± 0.8 μmoles per hour (SE) (5). Vco was estimated in five patients on two occasions 1 to 2 days apart. Vco varied only ± 4.5 μmoles per hour (SD) in the duplicate studies, and [COHb] varied ± 0.35% (SD). Figure 3 shows the increase in blood [COHb] in four patients studied on two occasions and illustrates the reproducibility and precision of the method.

Vco is compared to Vheme in Figure 4, where both are plotted in micromoles per hour. The data obtained from the patients with hemolytic anemia are plotted in closed circles, and the average Vco and calculated Vheme in ten normal supine male subjects at rest (5) are depicted by the filled square. The slope of the regression line is 1.04. The correlation coefficient is + 0.94.

The ratio of Vco to Vheme is listed in Table II for each study. This ratio varied from 0.99 to 1.92, averaging 1.40 ± 0.10 (SE).

The relationship between venous blood [COHb] and Vheme is plotted in Figure 5. The correlation coefficient between these variables is + 0.59.

**Discussion**

The rebreathing system developed for this study has made the measurement of Vco clinically feasible. Only two of the first twenty patients studied were not able to complete the procedure: one was a 70-year-old man with arteriosclerotic heart disease who could not lie flat as required with the present equipment.

In this study elevated Vco and blood [COHb] were found in seven patients with hemolytic anemia. The finding of elevated [COHb] in these patients confirms the report of Engstedt (10), and the elevated Vco demonstrates that the elevated [COHb] in patients with hemolytic anemia is actually a result of increased CO production. Rather poor correlation between the venous blood [COHb] level and Vheme was found (r = 0.59) in this study. The explanation for this finding appears to be that the blood [COHb] does not closely reflect the Vco. It has been shown in a previous publication (11) that [COHb] is influenced by the physiological processes that determine excretion of CO via the lungs, and by the quantity of CO in
inspired air, as well as by Vco. In that study Vco and venous blood [COHb] were compared in 13 experiments, including the eight in the present investigation, and a correlation of + 0.62 was found, a value similar to the correlation found here between Vheme and [COHb]. Since the relationship between Vheme and [COHb] cannot be closer than Vco to [COHb], the venous blood [COHb] measurements appear to be a relatively imprecise index of red cell survival.

In the present study those patients with the shortest red cell survival times had the highest Vco values, suggesting the Vco might be used to quantitate red cell destruction rates. Since the error in the Vco measurement is very small (less than 5% of total Vco in these patients), the precision with which erythrocyte destruction could be quantitated would depend on a) whether CO was produced as a result of processes other than catabolism of hemoglobin in significant amounts and b) whether the molar yield of heme to CO varied when the rate of hemoglobin catabolism was markedly increased.

Evidence is available that CO arises from processes other than circulating hemoglobin catabolism. In normal man approximately 20% of the Vco cannot be explained on the basis of destruction of hemoglobin from circulating erythrocytes (5). It is of interest that this “extra” CO is of the same magnitude as the “extra” stercobilin, which does not arise from circulating hemoglobin (19). It has recently been shown (20) that 14CO appears in the expired air of humans shortly after the administration of glycine-2-14C and that the CO specific activity parallels that in bile pigment. These findings suggest a common molecular origin or origins for extra CO and extra bile pigment. Extra bile pigment, sometimes called “shunt” bile pigment, is thought to originate as a result of ineffective erythropoiesis (catabolism of immature erythrocytes in the bone marrow, or short-lived cells in the circulation) or as a by-product of porphyrin metabolism in the liver, or both (19, 21–23). It would be expected in patients with hemolytic anemia that the quantity of CO originating from catabolism of erythrocytes might be very large compared to that originating from these
other sources, since the turnover of erythrocytes is so rapid.

To determine the per cent of total CO production arising from destruction of circulating erythrocytes in our patients, we estimated the rate of erythrocyte hemoglobin catabolism from $^{51}$Cr-labeled red blood cell measurements and compared this value, expressed as $\text{Vheme}$, with $\text{Vco}$. The estimation of $\text{Vheme}$ from $^{51}$Cr curves has many inherent errors. It requires an assumption regarding the mechanism of hemolysis and has uncertainties regarding a) elution of chromate from the labeled erythrocytes and b) steady state. The calculation of $\text{Vheme}$ is in addition affected by errors in measuring total red blood cell mass; however, this error cancels out in the comparison of $\text{Vheme}$ and $\text{Vco}$, since the red blood cell mass measurement is used in the calculation of $\text{Vco}$ as well.

In all the patients the curve of disappearance of $^{51}$Cr from the blood was exponential, and it therefore appeared that the hemolytic mechanism was random. Six of the seven patients had either S-S disease or autoimmune hemolytic anemia, and in these conditions others have reported that red cell destruction is random (24, 25). We, therefore, assumed that destruction was random in all the patients. The error from some degree of non-random destruction in these patients would be small; for example, if one-half of the red cells were destroyed in a non-random fashion, assuming a random mechanism would lead to an overestimation of $\text{Vheme}$ of only 12%.

The recent study of Cline and Berlin (15) has made it possible to estimate errors in $^{51}$Cr red blood cell survival measurements arising from uncertainty regarding the rate of elution of the label from the injected cells. These investigators measured decay curves of erythrocytes labeled simultaneously with $^{32}$diisopropyl fluorophosphate, $^{14}$C, and $^{51}$Cr in 38 patients who had a variety of hematological diseases. It was assumed that the difference in decay rates between $^{51}$Cr and the other labels was due to chromium elution. The rate of elution was found to be constant in most of the patients, was not related to the rate of hemolysis or the disease, and averaged 1.29% per day. We have calculated the standard deviation from the data presented by Cline and Berlin in this paper and found it to be $\pm 0.42$% per day. This uncertainty in chromate elution rate is relatively small compared to our average rate decrease in $^{51}$Cr radioactivity of 7.6% per day; the average error in $\text{Vheme}$ resulting from variation in elution rates should be considerably less than 10%. Even if any of our patients were at the extremes of the rates of chromium elution reported by Cline and Berlin, error in calculated $\text{Vheme}$ would be less than $\pm 15$% of total $\text{Vheme}$.

It is not possible to assess the error due to possible variations in the rate of erythrocyte destruction from day to day. The constant hemoglobin values and the close agreement of duplicate $\text{Vco}$ measurements suggest the patients were in a steady state.

From these considerations the error in $\text{Vheme}$ is estimated to be approximately $\pm 6$ to 8 $\mu$moles per hour. Most of these errors should be random and thus cause scatter of points in Figure 4. This estimate of the error in $\text{Vheme}$ appears to be in the correct range, since the scatter about the regression line in Figure 4 averages approximately 8 $\mu$moles per hour. The error in measurement of $\text{Vco}$ is only approximately $\pm 3$ $\mu$moles per hour, so it would appear that most of the scatter found in this Figure is a result of errors in the calculation of $\text{Vheme}$.

If the average value of 1.29% per day used in the correction for elution is in error, or if finite cell destruction was significant in our patients, a consistent error would result that would alter the regression line in Figure 4. Both of these possibilities appear to be very unlikely as discussed above.

If 1 mole of CO is produced from the destruction of 1 mole of heme, the data in Table II and Figure 4 show that an average of 74% of the CO produced arises from the destruction of circulating heme, whereas the remainder arises from other sources. This extra CO appears not to be an artefact. It cannot be explained on the basis of error in the calculation of $\text{Vheme}$. The extra CO is most likely the result of the same processes that yield extra CO in normal man. The absolute amount of extra CO is considerably greater than in normal man, suggesting that this fraction may arise primarily as a result of ineffective erythropoiesis. This concept is supported by the study of Gray and Scott (21), who stimulated erythropoiesis in one man by hemorrhage and found an
increase in shunt bile pigment. Furthermore, in patients with sickle cell anemia (26) the plasma radioiron turnover ratio is increased followed by reduced red cell radioiron incorporation, as is seen in patients with the ineffective erythropoiesis syndrome. It seems unlikely that the extra CO found in our patients could be explained on the basis of folic acid deficiency. The serum folic acid levels and quantities of histidine metabolites in the urine were found to be normal in three of four patients where these measurements were performed as indicated in Table I.

Estimation of mean erythrocyte life-span from CO production. The mean erythrocyte life-span (MLS) can be considered to be equal to the total body hemoglobin (T_heme) divided by the quantity of hemoglobin destroyed per day if a steady state exists where the rate of hemoglobin destruction equals the rate of production. Since CO is probably produced in a molar ratio to blood heme catabolized in these patients, the following relationship should exist: MLB = T_heme (micromoles) / V_co (micromoles per day). In Figure 6 is shown the relationship of MLS calculated from V_co, to MLS calculated from the 51Cr red cell survival studies, where MLS = 1/k' × 100. The points plotted in this Figure are all close to the least squares regression line. The slope of this line is 0.6 instead of 1.0 because of the extra CO production already discussed.

Because of the extra CO fraction, MLS should be calculated from V_co as above, and then the "actual" life-span can be read from the abscissa in Figure 6. MLS can be computed from the V_co in this manner in our series with an error of only ±3 days compared to the chromium method.

If most of the extra CO originates as a result of ineffective erythropoiesis, calculation of erythrocyte life-span from V_co without correction might indicate the average life-span of all erythrocytes, not just those labeled in the circulation. The mean erythrocyte life-span calculated from V_co without correction for extra CO may also prove to be a most useful determination when compared with results of measurements obtained by labeling circulating erythrocytes with radioactive tracers.

Summary

We have investigated the relationship of endogenous carbon monoxide production to red blood cell and hemoglobin catabolism in seven patients with hemolytic anemia. The rate of carbon monoxide production was markedly elevated in all of the patients. The rate of carbon monoxide production was found to correlate closely with the rate of blood heme destruction calculated from radiochromate red blood cell labeling studies (r = +0.94). Greater amounts of carbon monoxide were produced in six of eight studies than could be explained on the basis of blood heme catabolism. The average molar ratio was 1.41 ± 0.28 (SD). We suggest that the fraction of CO apparently not explainable on the basis of circulating hemoglobin catabolism, "extra CO," is a by-product of the metabolic processes that cause "shunt" bile pigment. Mean erythrocyte life-spans can be calculated from the rate of carbon monoxide production corrected for extra CO with an error of ±3 days (SD).

Acknowledgments

We gratefully acknowledge the expert technical assistance of Robert Ersek, Dolores Thyrum, Mary Friedmann, and Irving Nagleberg and the advice and cooperation of Dr. Robert E. Forster.

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