

## ALTERATIONS IN SULFOBROMOPHTHALEIN SODIUM-REMOVAL MECHANISMS FROM BLOOD DURING NORMAL PREGNANCY \*

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During the past few years, we have been impressed by the observation that our patients with pre-eclampsia and eclampsia usually demonstrate sulfobromophthalein sodium (BSP) retention in blood in the 45-minute BSP test. On the basis of these findings, we undertook a study of hepatic BSP-removal mechanisms during the course of normal pregnancy. Increased BSP retention in 45 minutes was not detected in normal pregnancy when compared to results obtained in a group of nonpregnant women of comparable age. When hepatic BSP-removal mechanisms were appraised quantitatively by a prolonged-infusion method (1, 2), however, significant changes in both hepatic uptake and excretion of BSP were observed. The following study presents in detail such data obtained from women in various stages of normal pregnancy and in the early postpartum period, and compares these findings with data obtained in a separate group of control nonpregnant women.

### METHODS

The method used in the present studies was devised by Wheeler and his associates (1, 2) and is based on the observations that removal of BSP from plasma depends on the simultaneous operation of at least two separate hepatic mechanisms: 1) uptake of BSP into a hepatic

"storage compartment," and 2) active secretion into bile (1-8). The quantity of BSP taken up into the "storage compartment" appears to be directly proportional to plasma concentration (1). Thus, uptake into storage during a period of changing plasma concentration can be represented in symbols as  $S \times \Delta P / \Delta t$ , where the "relative storage capacity"  $S$ , a proportional factor, is defined as the number of milligrams of BSP stored in the liver per milligram per 100 ml of plasma concentration, and  $\Delta P / \Delta t$  signifies the rate of change of plasma concentration in milligrams per 100 ml per minute. Secretion of BSP into bile is a rate-limited process characterized by a maximal excretory rate or  $T_m$  (1, 2, 8), defined in terms of milligrams per minute. The maximal rate of BSP secretion into bile is achieved when plasma BSP concentration is maintained in excess of 2 to 3 mg per 100 ml during prolonged infusions of BSP (1, 8). At these plasma concentrations, the general equation for the hepatic removal rate of BSP (1),  $R_H$ , in milligrams per minute, is as follows:  $R_H = S \times (\Delta P / \Delta t) + T_m$ .

Values for  $R_H$  and  $\Delta P / \Delta t$  obtained during two or more different rates of BSP infusion are substituted in the formula, providing a set of simultaneous equations that can be solved for both  $S$  and  $T_m$ . In practice, values for  $R_H$  and  $\Delta P / \Delta t$  were obtained during three separate infusion rates of BSP, each given for an hour. As indicated by Wheeler, when the corresponding values for  $R_H$  and  $\Delta P / \Delta t$  are plotted on the ordinate and abscissa, respectively, the relationship between  $R_H$  and  $\Delta P / \Delta t$  appears to be approximately linear. Thus, the equation of the line that best fits the three points is calculated by the method of least squares (9). Since the general equation for hepatic removal rate of BSP is in the form of a straight line,  $y = m \times x + c$ ,  $S$  is equal to the slope of the least-squares line, and  $T_m$  is equal to the value of the  $y$  intercept.

### Procedures

Single, prolonged-infusion studies were conducted before surgery on 10 control nonpregnant women in the reproductive age range admitted to the gynecology service, usually for therapy for stress incontinence or for simple hysterectomy; on 15 women in various stages of normal pregnancy; and on 11 women during the first week postpartum. The studies were usually carried out in the morning, approximately 1 hour after a fat-free breakfast. The procedure used was essentially the same as that described by Wheeler, Meltzer, and Bradley (2), with the exception that the BSP administered was di-

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TABLE I  
Hepatic sulfobromophthalein sodium (BSP)-removal mechanisms in control nonpregnant women

Patient	Age, years	BSA, m <sup>2</sup>	Plasma volume ml	Hemato- crit %	Total blood volume ml	Infu- sion period	Infu- sion rate mg/min	Average P* mg/100 ml	Distribution in plasma			Average ΔP mg/100 ml/ min	BSP reten- tion 45 min %	St mg/mg/100 ml	T <sub>m</sub> † mg/min	
									Conju- gated %	Free %	Average ΔP mg/100 ml/ min					
O.S.	24	1.45	1,835	41.1	3,115	I II III	14.70 5.01 8.75	7.84 3.77 5.12								
E.S.	35	1.52	2,150	40.0	3,585	I II III	12.95 4.12 7.10	8.28 6.44 6.74	5.7 25.9 26.3	94.3 74.1 73.7	+126 -039 +004	2.3	38.0		8.7	
J.N.	35	1.57	2,315	35.2	3,575	I II III	21.91 7.51 12.96	11.83 11.53 15.64	8.6 28.8 30.1	91.4 71.2 69.9	+207 -005 +091	2.5	45.2		7.4	
A.H.	28	1.58	3,050	37.9	4,910	I II III	17.20 5.80 10.40	5.02 3.09 3.94			+087 -032 +032		57.4		8.0	
R.W.	38	1.69	2,590	35.8	4,035	I II III	16.43 5.85 9.74	9.01 6.34 7.95			+125 -064 +026	3.7	32.4		9.2	
M.R.	39	1.72	2,405	35.8	3,745	I II III	14.40 5.30 8.35	9.48 6.46 6.39	7.2 32.9 25.0	92.8 67.1 75.0	+127 -054 -002		26.3		8.2	
B.B.	32	1.80	2,550	42.8	4,460	I II III	23.28 7.84 14.61	12.77 12.09 17.48	6.5 24.9 22.8	93.5 75.1 77.2	+225 +015 +099	4.5	48.0		7.0	
G.M.	45	1.80	2,770	40.4	4,650	I II III	22.90 7.92 14.65	9.23 7.88 10.14	5.3 21.8 18.9	94.7 78.2 81.1	+133 -034 +039	3.5	61.8		10.8	
J.A.	34	1.81	2,565	41.0	4,345	I II III	18.48 6.60 11.44	8.17 7.52 10.01			+140 -038 +032	6.1	41.3		9.2	
D.C.	29	1.86	2,815	39.0	4,615	I II III	18.16 6.65 11.08	7.85 5.02 5.34	5.3 24.6 23.6	94.7 75.4 76.4	+102 -032 +006		55.5		9.8	
Mean						I II III	18.04 6.32 10.91	8.95 7.01 8.88	6.4 26.5 24.5	93.6 73.5 75.5	+140 -032 +033	3.6	43.4		8.5	
													±SD 12.3		±SD 1.3	

\* P = plasma concentration of BSP.

† S = relative storage capacity, defined as number of milligrams BSP stored in liver per milligram per 100 ml plasma concentration.

‡ T<sub>m</sub> = maximal excretory rate of BSP into bile.

luted in isotonic saline. Plasma volume was measured in each subject during the first hour of the study with  $I^{131}$ -labeled human serum albumin. A plasma sample was obtained 10 minutes after injection of approximately  $5 \mu\text{C}$  of radioactive tracer for estimation of plasma volume. A 45-minute BSP test (10) was carried out in most control and pregnant subjects on the day before, and in postpartum women, just before the prolonged-infusion study.

In 14 additional studies, constant infusions of BSP were administered for 24 to 156 minutes to women in labor. At the time of delivery, samples of maternal and fetal cord blood were obtained simultaneously for estimation of plasma concentration of BSP. Placentas were obtained from 10 of these women and analyzed for BSP content.

The quantity of BSP excreted into urine during 65 to 97 minutes of BSP infusion was determined in four other studies conducted in women at term.

#### Analytical methods

**Plasma.** One ml of each plasma sample was mixed with 9 ml of 0.1 N KOH. The optical density was determined in a Beckman DU spectrophotometer set at  $580 \text{ m}\mu$ . Plasma obtained before infusion of BSP was treated similarly and served as a blank. Standard BSP curves were prepared for each patient by addition of known amounts of BSP to plasma obtained before BSP infu-

sion. Since blank samples of fetal cord blood could not be obtained, BSP concentration in fetal cord plasma was estimated by the method of Gaebler (11).

**Infusion mixture and urine.** Samples of infusion mixture and urine samples were diluted with distilled water and alkalinized, and the optical density was determined at  $580 \text{ m}\mu$ . For analyses of urine, samples obtained just before infusion of BSP were diluted to the same extent as the sample containing BSP and served as blanks.

**Chromatography of BSP compounds in plasma.** Equal volumes of plasma from each of the four plasma samples obtained during each hour of BSP infusion were pooled. BSP compounds were extracted from the pooled plasma and prepared for paper chromatography on Whatman 3MM paper as outlined by Carbone, Grodsky, and Hjelt (12). The chromatograms were developed in a descending system employing glacial acetic acid: water: *n*-propyl alcohol (1:5:10 by vol), for 16 to 18 hours. BSP bands were identified by exposing the dried papers to ammonia vapors. BSP was eluted from each band and the concentration determined by methods described in detail in previous publications (13, 14). One or two bands corresponding to BSP conjugates and a band containing free BSP were identified on the chromatograms. The proportion of BSP appearing as conjugated BSP was determined by dividing the sum of the optical den-

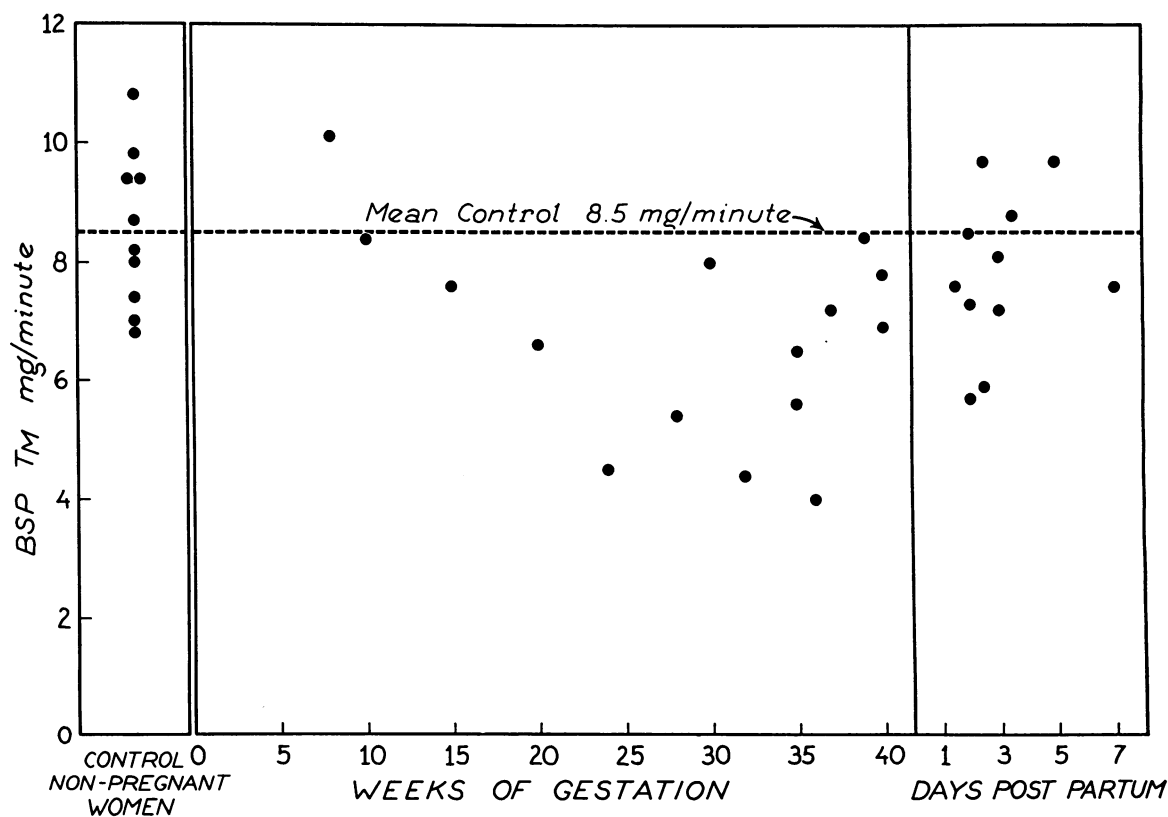


FIG. 1. ALTERATIONS IN MAXIMAL EXCRETORY RATE OF BSP INTO BILE (BSP  $T_m$ ) DURING NORMAL PREGNANCY AND THE FIRST WEEK POSTPARTUM.

TABLE II  
*Hepatic BSP-removal mechanisms during normal pregnancy*

Patient Age, years BSA, m <sup>2</sup> *	Weeks of gesta- tion	Plasma volume ml	Hemato- crit %	Total blood volume ml	Infu- sion period	Infusion rate mg/min	Average P mg/100 ml	BSP				BSP reten- tion 45 min %	S mg/mg/ 100 ml	T <sub>m</sub> mg/min
								Distribution in plasma			Average ΔP mg/100 ml/ min			
								Confu- gated %	Free %	%				
L.S. 33 1.87	8	2,645	40.2	4,425	I	24.38	16.73	9.8	90.2	+0.376	36.5	10.1		
					II	8.37	22.57	28.4	71.6	-0.002				
					III	14.29	28.18	31.3	68.7	+0.057				
B.H. 30 1.65	10	3,050	36.1	4,775	I	16.00	5.21	3.6	96.4	+0.102	46.3	8.4	1.4	
					II	5.72	3.74	19.7	80.3	-0.027				
					III	9.53	3.97	12.8	87.2	+0.004				
A.L. 26 1.46	15	2,855	30.0	4,080	I	16.13	5.17	9.0	91.0	+0.067	102.5	7.6	1.4	
					II	5.63	3.27	35.3	64.7	-0.007				
					III	9.75	3.70	29.2	70.8	+0.005				
L.W. 37 1.84	20	3,600	35.4	5,575	I	19.06	8.08	12.6	87.4	+0.156	46.1	6.6	7.3	
					II	6.86	9.44	41.4	58.6	+0.010				
					III	12.56	12.22	36.7	63.3	+0.062				
C.T. 23 1.80	24	4,665	30.0	6,665	I	20.36	4.59	4.2	95.8	+0.070	144.1	4.5	1.4	
					II	7.05	3.85	23.5	76.5	+0.009				
					III	11.75	6.16	23.5	76.5	+0.040				
L.W. 17 1.81	28	3,625	31.3	5,275	I	23.78	6.72	6.6	93.4	+0.125	98.7	5.1	3.3	
					II	8.25	6.98	29.2	70.8	+0.016				
					III	13.70	11.56	27.9	72.1	+0.086				
L.J. 29 1.77	30	3,305	29.3	4,719	I	18.86	5.26	5.7	94.3	+0.074	110.8	8.0	2.8	
					II	6.56	4.29	15.7	84.3	-0.011				
					III	11.83	6.04	9.6	90.4	+0.026				
D.S. 37 1.75	32	5,520	31.2	8,125	I	20.00	5.83			+0.088	107.0	4.4		
					II	6.96	6.83			+0.022				
					III	11.44	9.95			+0.045				
J.M. 30 1.62	35	3,970	36.1	6,215	I	19.04	5.32			+0.072	105.9	6.5		
					II	6.21	3.95			-0.001				
					III	11.18	4.99			+0.013				
R.M. 17 1.68	35	2,940	41.0	4,985	I	14.5	4.42			+0.062	91.5	5.6	6.1	
					II	5.25	3.48			-0.006				
					III	8.58	5.07			+0.018				

\* Body surface area based on prepregnant weight.

TABLE II—Continued

BSP													
Patient Age, years BSA, m <sup>2</sup> *	Weeks of gesta- tion	Plasma volume ml	Hemato- crit %	Total blood volume ml	Infu- sion period	Infusion rate mg/min	Average p	Distribution in plasma			BSP reten- tion 45 min %	S mg/mg/ 100 ml	T <sub>m</sub> mg/min
								Conju- gated %	Free %	Average ΔP mg/100 ml/ min			
P.S. 26 1.45	36	3,060	36.0	4,780	I II III	13.07 4.56 7.75	5.25 4.51 6.28			+ .069 - .002 + .027		92.0	4.0
B.N. 22 1.60	37	3,595	36.0	5,615	I II III	17.60 5.90 9.84	7.60 7.52 10.01	9.1 22.7 24.9	90.9 77.3 75.1	+ .111 - .018 + .035	10.8	56.3	7.2
M.K. 16 1.73	39	4,110	32.1	6,640	I II III	22.79 7.75 13.36	6.80 6.46 8.07	3.8 17.3 19.0	96.2 82.7 81.0	+ .106 + .010 + .019	4.2	92.3	8.5
L.S. 24 1.69	40	3,750	32.2	5,530	I II III	16.2 5.83 9.78	4.22 4.60 5.49			+ .081 - .012 + .031	5.5	71.6	6.9
K.R. 23 1.83	40	3,970	33.0	5,925	I II III	19.67 6.42 11.14	4.80 4.54 6.30			+ .084 - .015 + .036	5.1	91.8	7.8

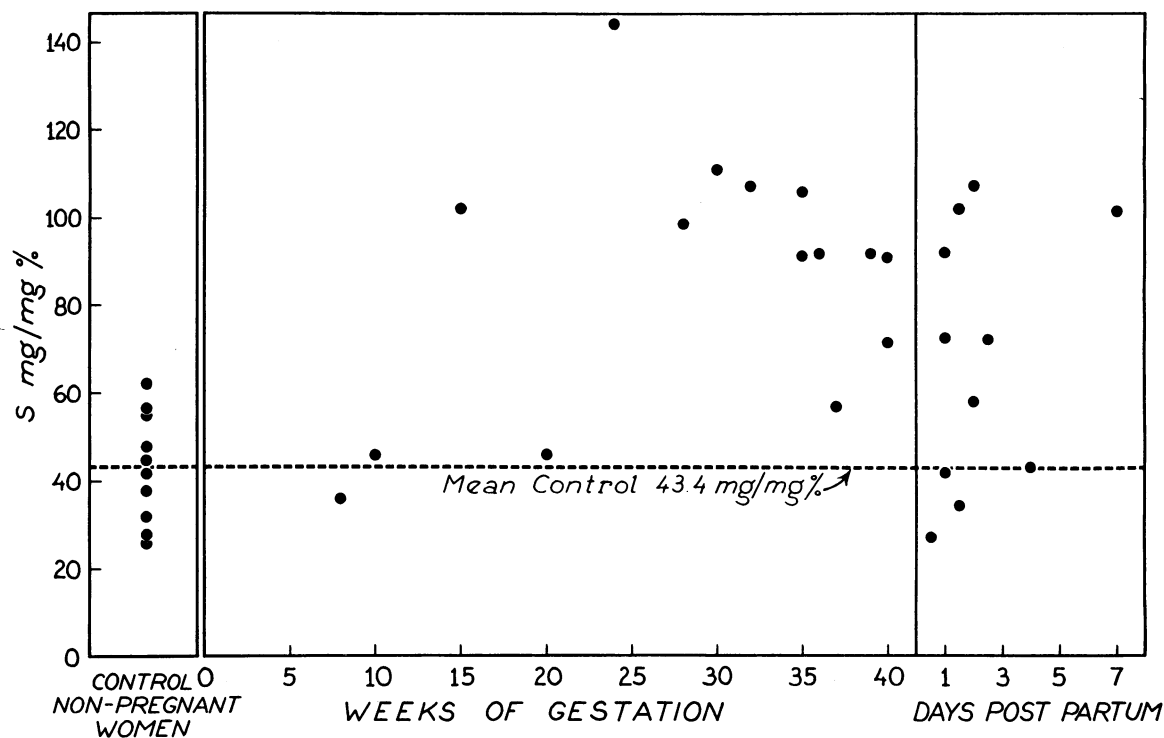


FIG. 2. ALTERATIONS IN HEPATIC RELATIVE STORAGE CAPACITY FOR BSP (S) DURING NORMAL PREGNANCY AND THE FIRST WEEK POSTPARTUM.

sities of the conjugate bands by the sum of the optical densities of the conjugate and free BSP bands. For these studies, it was assumed that the molecular extinction coefficient for free and conjugated BSP was identical.

**Extraction of BSP from placenta.** The amnion and chorion were stripped off, and the umbilical cord was cut at its juncture with the placenta. The placenta was weighed, cut into small pieces roughly 1-inch square, and homogenized in a Waring Blendor with a volume of distilled water equal to the weight of the placenta. Two-ml samples of homogenate, equivalent to 1 g of placenta, were pipetted in triplicate into glass-stoppered, conical centrifuge tubes. To extract BSP, 4 ml of 80% acetone was added to each tube. After thorough shaking, the tubes were centrifuged at 4,500 rpm for 10 minutes. The supernatant fluid was removed and the precipitate extracted twice with 3 ml of 80% acetone. The supernatant fluids were pooled and the volume recorded. Duplicate 3-ml samples of the combined acetone extract were mixed with 1 ml of 0.1 N KOH, and the optical density was determined by using 0.1 N KOH as a blank in a Beckman DU spectrophotometer set first at 580  $m\mu$  and then at 620  $m\mu$ . The alkalinized acetone extracts exhibited varying amounts of turbidity. The optical density at 580  $m\mu$  of similarly turbid alkalinized acetone extracts of placentas free of BSP averaged 1.10 times the optical density at 620  $m\mu$ . Thus, the BSP content of each placenta was calculated as follows:  $[\text{OD } 580 - 1.10 \text{ OD } 620$

$(\text{unknown})]/[\text{OD } 580 - 1.10 \text{ OD } 620 (\text{standard})] \times \text{concentration of standard solution in milligrams per 100 ml} \times \text{volume of acetone extract in fractions of 100 ml} \times \text{weight of the placenta in grams}$ . Since recovery of known amounts of BSP homogenized with placentas averaged 62%, the value above was multiplied by 100/62.

## RESULTS

**1. Maximal excretory rate of BSP into bile, BSP  $T_m$ .** In 10 control nonpregnant women, the average BSP  $T_m$  was 8.5 mg per minute  $\pm$  1.3 SD. The individual results are listed in Table I.  $T_m$  appeared to rise with increasing surface area, but the correlation coefficient of 0.5 was not statistically significant,  $p > 0.05$ . BSP  $T_m$  decreased during the latter part of normal pregnancy (Table II, Figure 1). The average  $T_m$  in the last half of pregnancy of 6.2 mg per minute  $\pm$  1.7 SD represented a fall of 27% below control, a statistically significant decrease,  $t = 3.38$ ,  $p < 0.01$ . A rapid return of  $T_m$  to control values was observed during the first week postpartum (Table III, Figure 1).

**2. Hepatic relative storage capacity, S.** The average value for S in 10 control nonpregnant

TABLE III  
Hepatic BSP-removal mechanisms during first week postpartum

Patient Age, years BSA, m <sup>2</sup> *	Days post- partum	Plasma volume ml	Hemato- crit %	Total blood volume ml	Infu- sion period	Infu- sion rate mg/min	Average P mg/100 ml	Distribution in plasma			Average ΔP mg/100 ml/ min	BSP reten- tion 45 min %	S mg/mg/ 100 ml	T <sub>m</sub> mg/min
								Conju- gated %	Free %	Average P mg/100 ml				
C.A. 37 1.60	1½	2,435	38.2	3,940	I II III	19.20 3.96 11.06	7.70 6.56 8.97	11.0 48.0 29.9	89.0 52.0 70.1	+ .228 - .033 + .029	4.7	27.3	7.6	
M.T. 20 1.57	1	2,720	31.3	3,960	I II III	16.38 5.86 9.17	8.01 6.89 9.97	13.9 36.9 28.8	86.1 63.1 71.2	+ .085 - .003 + .035	3.6	91.9	5.7	
E.J. 31 1.63	1	2,910	29.0	4,100	I II III	17.63 5.86 10.12	5.98 6.59 8.67	12.9 45.8 46.0	87.1 54.2 54.0	+ .147 + .004 + .013	5.3	41.5	7.3	
E.J. 24 1.70	1	2,855	37.5	4,570	I II III	15.64 5.44 9.18	5.12 3.18 3.32	6.0 27.5 21.9	94.0 72.5 78.1	+ .074 - .028 + .002		72.5	8.5	
M.D. 19 1.55	1½	3,070	38.5	4,990	I II III	20.38 4.58 8.02	14.04 12.82 13.42	9.7 39.8 41.7	90.3 60.2 58.3	+ .220 - .024 + .038	14.5	34.9	5.9	
D.J. 18 1.75	1½	2,885	37.8	4,640	I II III	20.68 7.02 12.28	7.60 6.64 6.77	8.8 26.7 36.9	91.2 73.3 63.1	+ .096 - .021 + .008		101.9	9.7	
E.M. 23 1.83	2	3,695	38.0	5,960	I II III	17.37 5.79 10.05	4.65 3.03 3.95	6.9 26.6 26.4	93.1 73.4 73.6	+ .063 - .016 + .014	4.1	106.7	8.1	
A.W. 17 1.70	2	2,670	27.3	3,675	I II III	17.55 0.38 10.37	8.53 7.35 8.92	9.4 32.0 27.9	90.6 68.0 72.1	+ .124 - .004 + .030	4.7	58.3	7.2	
D.W. 23 1.83	2½	3,620	38.1	5,850	I II III	19.61 7.51 11.27	5.92 4.90 6.25	7.8 31.2 29.2	92.2 68.8 70.8	+ .099 - .013 + .026		72.0	8.8	
J.H. 21 1.61	4	2,895	31.0	4,195	I II III	18.16 6.85 10.94	7.77 6.84 7.69	8.4 31.0 30.2	91.6 69.0 69.8	+ .120 - .030 + .012	3.0	43.1	9.5	
E.B. 25 1.72	7	2,315	43.0	4,130	I II III	17.63 5.54 1.10	6.10 4.35 5.46	3.9 20.1 20.4	96.1 79.9 79.6	+ .083 - .011 + .014		101.4	7.6	

\* Body surface area based on prepregnant weight.

women was  $43.4 \text{ mg per mg per } 100 \text{ ml} \pm 12.3 \text{ SD}$  (Table I). No correlation between S and surface area was apparent. S did not change during the early part of pregnancy (Table II, Figure 2). Later, however, S rose markedly and averaged  $96.5 \text{ mg per mg per } 100 \text{ ml} \pm 22.6 \text{ SD}$  during the last half of pregnancy. The increase of 122% above control in the latter half of pregnancy was highly significant,  $t = 6.56$ ,  $p < 0.001$ . After delivery, S tended to fall toward control values, although it still remained elevated as long as one week postpartum in some patients (Table III, Figure 2).

3. *BSP concentration in maternal and fetal cord plasma.* Constant infusions of BSP were administered to 14 women at term for a mean period of 73 minutes. During this time, an average of 1,177 mg of dye was removed from the circulation. The concentration of BSP in maternal plasma at the time of delivery averaged  $9.2 \text{ mg per } 100 \text{ ml}$ . In fetal cord plasma, sampled at the same time, no BSP was detected in 10 specimens. BSP was found in the other 4. In these instances, the concentration of dye, in milligrams per 100 milliliters, in fetal cord and maternal plasma, respectively, was 0.08 fetal to 14.6 maternal; 0.16 to 5.8; 0.63 to 17.0; and 3.6 to 7.6.

4. *BSP uptake by placenta.* BSP was infused for an average of 70 minutes in 10 women at term. Of the 1,085 mg of BSP removed from blood during this period, an average of only 7.2 mg could be accounted for in the placentas. Since placentas contain approximately 50 ml of maternal plasma (15), the quantity of BSP to be expected in this volume of plasma (maternal plasma concentration of BSP in milligrams per 100 ml at time of delivery  $\times 0.5$ ) averaged 5.1 mg. Actual tissue content of BSP was approximately 2.1 mg of BSP per placenta, therefore.

5. *Urinary excretion of BSP.* An average of 1,526 mg of BSP was removed from blood during a mean infusion period of 83 minutes in four women at term. Only 8.1 mg of BSP was excreted into urine during this period, on the average.

#### DISCUSSION

Almost all of the results of the 45-minute BSP tests carried out during the course of normal preg-

nancy (Table II) fell within the range of values obtained in a group of control nonpregnant women. It seemed unlikely, therefore, that any alterations in hepatic BSP-removal mechanisms would be noted during the course of normal pregnancy. However, two significant changes were observed when the prolonged-infusion method of Wheeler and his associates (1, 2) was utilized. Thus, S increased markedly, whereas BSP  $T_m$  decreased during the last half of pregnancy. It is apparent that the Wheeler method provides an extremely useful tool for examining hepatic BSP-removal mechanisms. It not only permits an appraisal of both hepatic uptake and biliary excretion of BSP in quantitative terms, but obviously it is capable of detecting significant changes even when the sensitive clinical BSP test yields normal results.

The most impressive finding in the present studies was the marked rise in S during the last half of pregnancy. There are several possible explanations for this change. First, it was necessary to demonstrate that significant BSP removal from blood in sites other than the liver did not account spuriously for the higher values of S observed in the last half of pregnancy. Since S was as high at 24 weeks of gestation as at 40, it seemed unlikely that BSP uptake by the placenta or the fetus, which would enlarge considerably during this period of gestation, contributed in any significant way to BSP removal from plasma. This was corroborated by our studies which demonstrated that virtually no BSP is taken up by or crosses the placenta. Others have examined transplacental movement of BSP after administration of a single iv injection of 5 mg per kg. Freiheit detected no dye in cord blood of two infants delivered within half an hour of BSP administration (16). Smith, Moya, and Shnider (17) observed BSP in umbilical vein blood as early as 20 seconds after BSP injection, and low levels of BSP were found in nine of 15 subjects. No BSP was detected in umbilical vein blood of six infants. In our constant infusion studies in which cord blood was sampled, no measurable amount of BSP was found in 10 of 14 subjects. If a large quantity of BSP had passed into the fetal circulation, it seems likely that relatively high levels of BSP would have been detected,



since BSP removal from blood is delayed in the newborn infant (18–22). In the present studies, urinary loss of BSP was small in pregnancy. On the basis of earlier observations (1, 23–25), it seems unlikely that other extrahepatic sites accounted for a marked increase in BSP removal from blood, although this possibility is not excluded. Significant uptake of BSP by tissues other than the liver apparently does not explain the rise in *S*.

Before attributing increased *S* to an intrinsic change in the liver, it is necessary to consider other extrahepatic factors that might result in increased hepatic BSP storage in pregnancy. With the method used in these studies, it is assumed that the quantity of BSP taken up into storage for each increment in plasma concentration of 1 mg per 100 ml is the same over a limited range of plasma concentration (1). It is conceivable that the capacity of the storage mechanism differs when changes in plasma concentration occur at low and high plasma levels of BSP. If the mechanism accounting for storage is composed of units with different affinities for BSP, components with high affinity would take up BSP first, thus at low plasma levels. Units with progressively lower affinity would remain as the quantity of BSP stored increases during rising plasma concentration. Thus lower values for *S* would be expected at higher plasma levels. In the present studies, average plasma levels of BSP during each hour of BSP infusion were somewhat lower in women in the last half of pregnancy than in control nonpregnant women (5.53, 4.18, and 7.27 mg per 100 ml in pregnant women compared to 8.95, 7.01, and 8.88 mg per 100 ml in control nonpregnant women, Tables I and II). Apparently, in the dog, *S* is relatively constant over a range of BSP levels in plasma of at least 3 to 12 mg per 100 ml (26). The plasma concentrations attained in the present studies fell well within this range, and if *S* behaves in the human as it does in the dog, it seems unlikely that the small differences in plasma levels could account for the large changes in *S* observed during pregnancy.

The degree of protein-binding of BSP in plasma may exert an influence on uptake of BSP into storage. It is possible that the quantity of BSP

stored in the liver is proportional to a small fraction of unbound dye in plasma rather than to total plasma concentration of BSP. Thus, increased unbound BSP in plasma of pregnant women might result in increased *S*. In preliminary studies (27), ultrafiltrates of plasma, obtained from both control and pregnant women, to which BSP had been added *in vitro* revealed no free BSP in plasma water with total BSP concentrations in plasma of up to 20 mg per 100 ml. These observations suggest, therefore, that differences in protein-binding over the range of plasma BSP concentrations attained in the present studies did not influence values of *S*.

A rise in *S* might be considered to result from a decrease in  $T_m$ . This seems unlikely, however, for in at least three situations—in patients with the Dubin-Johnson syndrome (2), after acute ligation of the common bile duct in dogs (26), and during administration of 17 $\alpha$ -ethyl-19-nortestosterone (37)—*S* has remained normal when  $T_m$  is decreased markedly.

It seems reasonable to conclude, therefore, that an increase in *S* during the last half of pregnancy indicates that the relative storage capacity of the liver for BSP increases, and furthermore, that this alteration is due to an intrinsic change in the liver. Such an increase in *S* could be accounted for by an increase in hepatic mass, by increased storage capacity of each unit of liver tissue, or by both. As for increased hepatic mass in normal pregnancy in the human, few data are available for analysis. A review of autopsy information derived from women dying after a brief illness during pregnancy in our hospital revealed an average liver weight of approximately 1,700 g, with no significant difference in liver mass during early and late pregnancy. Thus, an increase in size of the liver does not explain the rise in *S*. Normally, BSP is concentrated in human liver three to four times above plasma concentration, on the average (2). Although the nature of the concentrating mechanism is not known—whether it is a liver protein with greater affinity for BSP than plasma protein, an active transport mechanism, or something else—, an increase in capacity of this mechanism in each unit of liver tissue is the most likely explanation for the rise in *S* during pregnancy.

The implication of this observation appears to extend beyond hepatic uptake of BSP. Obviously, the human liver is not usually exposed to BSP. A rise in storage capacity for this dye could hardly be considered a specific response, therefore. It seems reasonable to expect that increased hepatic storage capacity for a variety of compounds, including drugs, will be found during pregnancy.

The fall in  $T_m$  in the last half of pregnancy may also be accounted for by several possibilities. First, if the liver holds on to BSP more avidly, less BSP may be available for excretion. The observation that  $S$  remained increased at a time when  $T_m$  was normal in several postpartum patients suggests another mechanism, however. Second, interference with BSP conjugation might result in decreased  $T_m$ . Previous work indicated that conjugation of BSP with glutathione (28–30), a process catalyzed by a liver enzyme (31), does not influence the extent to which BSP is stored in the liver (32). BSP  $T_m$  is dependent on conjugation, however, since conjugated BSP is delivered into bile at a faster rate than free BSP (32). The components of the BSP-glutathione conjugating system have not been examined in human liver during pregnancy. In the rat, a decrease in BSP conjugating enzyme activity with normal levels of glutathione has been found during the last 5 days of gestation (14). When liver glutathione levels have been maintained, however, a moderate decrease in enzyme activity has not resulted in a decrease in BSP  $T_m$  in the rat (32). Whether conjugation is impaired as a result of a critical decrease in enzyme activity or glutathione content in the human liver must await further studies. An analysis of BSP compounds appearing in plasma was carried out in order to detect any major changes in conjugation that might occur in the present studies. BSP conjugates were observed in plasma during the prolonged infusions of BSP (Tables I to III). The proportion of total plasma BSP accounted for by BSP conjugates during each hour of infusion was almost the same in patients during the last half of pregnancy and in control nonpregnant women (5.8, 21.6, and 20.9% in pregnancy, as compared to 6.4, 26.4, and 24.4% in controls). The small differences between preg-

nant and control patients in the averages for each hour were not significant statistically. Since the quantity of BSP infused was approximately the same in pregnant and nonpregnant subjects, the data above suggest that major changes in BSP conjugation in pregnancy did not affect  $T_m$ .

Finally, decreased  $T_m$  may result from an impairment of the transport processes involved in the delivery of both conjugated and free BSP from liver cells to bile. In this regard, it is of interest that a syndrome of obstructive jaundice has been observed during the latter part of pregnancy (33–36). After delivery, the obstructive features disappear, and hepatic function returns to normal. Obstructive jaundice may recur with subsequent pregnancies. The nature of the intrahepatic disturbance appears to involve impairment of biliary secretory mechanisms. It is tempting to speculate that the decrease in BSP  $T_m$  observed in the present studies reflects a similar response of the liver, although obviously a less marked one, to changes occurring normally during pregnancy. According to this view, obstructive jaundice might result either from an increase in quantity of the factor that accounts for decreased BSP  $T_m$  in normal pregnancy, or from a greater sensitivity of biliary secretory mechanisms to normal amounts of this factor found in pregnancy.

Analysis of various mechanisms that might account for both the rise in  $S$  and the fall in  $T_m$  in pregnancy appears to indicate that alterations in two independent hepatic systems are involved. Furthermore, since the changes in hepatic BSP-removal mechanisms occur during the last half of pregnancy, and then return to or toward normal in the first week after delivery, it seems likely that they are mediated by increased hormone levels in pregnancy, probably of estrogens, progesterone, or both. With decrease in hormone levels after delivery, BSP-removal mechanisms return toward normal.

#### SUMMARY

Hepatic sulfobromophthalein sodium (BSP)-removal mechanisms have been appraised quantitatively during pregnancy and the first week postpartum in terms of 1) the relative storage ca-

capacity for BSP,  $S$ , defined as the number of milligrams of dye taken up into storage for each increment of plasma concentration of 1 mg per 100 ml and 2) the maximal excretory rate of BSP into bile,  $BSP\ T_m$ , in milligrams per minute.  $S$  rose 122% during the last half of pregnancy, then returned to or toward normal during the first week postpartum. In contrast,  $BSP\ T_m$  decreased 27% in the last half of pregnancy, then rapidly increased to normal levels after delivery. These changes appear to reflect alterations in two independent hepatic mechanisms.

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