URINARY CONCENTRATING ABILITY IN PREGNANT WOMEN WITH ASYMPTOMATIC BACTERIURIA*

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Studies of patients with acute and chronic pyelonephritis have suggested that impairment of urinary concentrating ability may be an early feature of this disease (1-3). In these reports the diagnostic criteria for pyelonephritis have included clinical and pyelographic findings together with an abnormal urinary sediment with or without positive cultures. More recently, increasing emphasis has been placed on significant bacteriuria determined by quantitative urine cultures as a criterion for active pyelonephritis (4). In some subjects in whom bacteriuria has been discovered for the first time by quantitative urine cultures, the diagnosis of pyelonephritis has then been established from the clinical and laboratory findings (5); but in other individuals with asymptomatic bacteriuria, the normal urinary sediment and pyelographic findings, and even postmortem microscopic examination of the kidneys, have not supported a clinical suspicion of silent pyelonephritis (5, 6).

The present study was undertaken to determine whether an abnormality in concentrating ability might exist in healthy individuals with numerically significant bacteria in the urine. Pregnant women were chosen for study, since the incidence of bacteriuria is reported to be high in this group (7), whereas conditions known to impair concentrating ability, such as malnutrition, electrolyte disturbances and degenerative renal diseases, would not be encountered commonly among young, pregnant women. The present report concerns the finding of impaired urinary concentrating ability following 24 hours of fluid deprivation in pregnant women

with asymptomatic bacteriuria, compared with a suitable control group.

METHODS

The subjects were healthy, pregnant women attending the Beth Israel Hospital Prenatal Clinic. A detailed report concerning bacteriuria and pyelonephritis in this group has been presented elsewhere (8). The technique used to obtain clean-voided urine cultures was as follows. The perineum was cleansed with sponges immersed in a 1:1,000 solution of benzalkonium chloride. A voided specimen was then collected in a sterile bottle and promptly refrigerated. Within 24 hours quantitative urine cultures were performed with limited identification of those bacteria appearing in significant numbers as described previously (5). Pregnant women whose cultures revealed approximately 100,000 or more bacteria per ml in a random, clean-voided urine specimen obtained at the first clinic visit were requested to return for a repeat culture. If the second, random, cleanvoided urine specimen again contained 100,000 or more bacteria per ml a 24 hour fluid-deprivation concentration test was carried out. Subjects were told that a urinary tract infection might be present and were instructed to refrain from all fluids from breakfast on one day until after completion of the test the next morning; dry meals were permitted. Initially only a single, clean-voided urine specimen for culture and determination of osmolality was obtained at the termination of the 24 hours of dehydration. Later in the study a 24 hour urine collection during the period of dehydration was also obtained for determination of the endogenous creatinine clearance and the osmolal clearance. Subjects were asked to keep the 24 hour urine collection refrigerated; no preservatives were used. A venous blood specimen was obtained at the end of the period of dehydration for determination of creatinine and osmolality. All samples were refrigerated until analyzed. Osmolality determinations were performed within 24 hours for most specimens.

Pregnant women of age and duration of pregnancy comparable to those with bacteriuria, but with low bacterial counts in clean-voided urine specimens, were chosen as control subjects. It was difficult to match the groups exactly with respect to parity since the incidence of bacteriuria rose sharply with the number of prior pregnancies (7, 8). However, as many grandmultiparous

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Data for Group A subjects with significant bacteriuria TABLE I

										Serum after	after		24-Ho	ur urine d	24-Hour urine during fluid deprivation	deprivat	ion
					Random CVS*	1 CVS*	Final CVS 2	Final CVS after fluid deprivation	ivation	fluid dep	rivation	Orine/ serum					Cosm/
Subject	Age	Grav.	Para.	Wks†	Date	Culture	Date	Culture	Osmol- ality	Osmol- ality	Creat- inine	osmol- ality	Solute‡	Creat.‡	Cosm‡	Cort	X 186
	311.5					Bact./ml§		Bact./ml§	mOsm/ kg H ₂ O	mOsm/ kg H ₂ O	mg/ 100 ml		μOsm/ min	mg/ min	ml/min	z	
14	29	ß	4	26	11/18/58 11/26/58	over 10 ⁶ 306,000	12/ 3/58	over 106	816	290	99.0	2.81					
2A	22	6	2	16	11/18/58 11/26/58 11/29/58	over 106 over 106 730,000	12/ 3/58	1,000	006	286	0.78	3.15					
3A	31	က	-	20	11/18/58 12/ 5/58	over 10 ⁶ 78,000	12/10/58	over 106	480	318	0.81	1.51					
4A	27	S	ဗ	19	12/9 /58 12/31/58	over 106 over 106	12/17/58	over 106	675	294	0.81	2.30					
5A	19	4	3	34	12/30/58	298,000	1/ 6/59	over 106	669	285	0.92	2.45					
6A	22	2	-	23	1/13/59 1/28/59	over 106 over 106	1/21/59	over 106	096	300	96'0	3.20					
7.A	21	-	0	19	1/13/59 1/28/59 2/18/59	91,000 P 91,000 P 576,000 P	1/22/59	247,000 P	780	280	0.99	2.79					
8A	20	1	0	16	3/31/59 4/5/59	132,000 113,000	4/ 7/59	323,000	636	285	1.00	2.23					
9A	27	S	ဗ	27	4/ 7/59 4/22/59	over 106 over 106	4/11/59	435,000	570	282	0.93	2.03					
10A	30	9	S	22	4/ 7/59 4/22/59	over 106 over 106	4/16/59	550,000	789	295	0.82	2.67					
11A	20	7	-	14	6/30/59 7/14/59	368,000 972,000	7/16/59	over 106	570	295	0.80	1.93					
12A	41	9	S	26	12/ 8/59 1/ 8/60	305,000 339,000	1/12/60	over 106	260	294	0.74	0.88	554	0.91	1.88	123	1.53
13A	30	9	ဗ	10	1/26/60 2/ 1/60	over 106 over 106	2/ 5/60	over 106	1,000	296	0.90	3.38	467	0.67	1.58	74	2.14
14A	32	6	∞	17	$\frac{2}{9}$, $\frac{9}{60}$	over 10 ⁶ 387,000	3/ 2/60	320,000	488	293	96.0	1.67	326	0.91	1.11	95	1.17
15A	24	က	7	15	2/23/60 3/ 2/60	695,000 120,000	3/15/60	139,000	723	297	0.86	2.43	570	0.75	1.92	87	2.21
16A	27	∞	7	31	4/ 5/60 4/ 9/60	over 106 over 106	4/16/60	over 106	729	290	0.62	2.51	594	0.94	2.05	152	1.35
17A	31	•	Ŋ	19	4/12/60 4/16/60	over 10 ⁶ 473,000	4/21/60	427,000	899	300	0.92	3.00	322	99.0	1.07	72	1.49
18A	24	7	-	25	9/27/60 11/ 4/60	over 106 over 106	11/29/60	over 10°	757	295	0.96	2.57	283	0.81	96'0	84	1.14
19A	25	ь	7	30	11/29/69 12/ 6/60	over 106 over 106	12/ 9/60	over 106	912	300	1.06	3.04	342	0.79	1.14	72	1.52
20A	30	8	7	35	11/29/60 12/ 6/60	over 106 over 106	12/15/60	over 106	477	301	0.00	1.58	382	0.91	1.27	101	1.26
Mean SD	27	4.2	2.9	22					706 193	294 7	0.87	2.41	427 121	0.82	0.42	56 26	0.39
* Clean	s papion.	ided specimen															

Clean-voided specimen.
Duration of pregnancy at time of concentration test.
Corrected to 1.73 cm² body surface area.
All E. coti, except TA, Proteus (P).
Included in Group A since all other specimens contained over 10⁵ E. coli/ml.

control subjects as could be obtained were included to make the groups as comparable as possible. To provide a similar incentive to refrain from fluids for 24 hours, all women were told that a urinary tract infection might be present.

Osmolalities were measured with a Fiske osmometer. Creatinine was determined by the method of Brod and Sirota (9). Since endogenous serum creatinine levels are low in pregnancy (10), a known amount of creatinine was added to some specimens to bring the photometric readings into a more desirable range, and this amount was substracted from the final calculation.

RESULTS

The results are considered separately for three different groups of subjects. The first group (A) was composed of 20 women whose urine specimens persistently contained over 100,000 gramnegative bacteria per ml, usually E. coli; none was encountered with persistently high counts of gram-positive organisms. These 20 women were considered to have bacteriuria indicative of bacterial multiplication within the urinary tract. Data for this group are shown in Table I. The second group (B) were 30 women whose urine specimens repeatedly contained fewer than 100,000 bacteria per ml. These organisms were presumed to represent contamination. Women with persistently low-count urine cultures were not considered to have bacteriuria and served as a control for Group A. Data for this low-count control group are shown in Table II. The third group (C) included 20 subjects whose urine specimens had more than 100,000 bacteria per ml on one occasion, whereas other specimens from the same individual at other times contained small numbers of bacteria. In general, the bacteria in Group C were grampositive, varied considerably in kind and concentration, and most likely represented contamination rather than true bacteriuria. However, occasionally gram-negative bacteria appeared in large numbers in one or more specimens and it was difficult to distinguish excessive contamination from transient bacteriuria. Data for Group C subjects are presented in Table III.

Women with asymptomatic bacteriuria (Table I) had significant impairment of urinary concentrating ability compared with control subjects in Group B without bacteriuria (Table II). Serum osmolality values after 24 hours' dehydration were the same in both groups. Nine of the 20 women in Group A had urinary osmolality values

below 700 mOsm per kg H₂O in the final specimen following dehydration; none of the women in the control Group B showed this degree of impairment. Subjects in Group C with occasional high bacterial counts in the urine, probably due to contamination, had maximal urinary osmolality values after 24 hours' dehydration, similar to those for control Group B. The impairment in concentrating ability observed in women with asymptomatic bacteriuria was highly significant (p < 0.01) compared with the control Group B, both with respect to the mean maximal urinary osmolality (706 mOsm per kg H₂O for Group A, compared with 935 mOsm per kg H₂O in Group B) or the mean maximal urine/serum osmolality ratio (2.41 for Group A compared with 3.16 for Group B). Except for the subject with the severest impairment (Subject 12 in Group A), symptoms of urinary frequency or nocturia did not reflect the degree of impairment of concentrating ability in the 24 hour concentration test.

Serum creatinine concentrations determined in all 20 women with bacteriuria in Group A and in 21 women in the control Group B did not differ significantly. Timed urine specimens during the 24 hour period of dehydration were obtained from 9 women in Group A and 19 women in Group B. The rates of solute excretion and osmolal clearance averaged about 9.5 per cent higher in the Group A women with bacteriuria, but these differences were not significant. The endogenous creatinine clearance averaged about 20 per cent lower in the bacteriuric group (0.05 >p > 0.02). The combined effects of a slightly higher osmolal clearance and slightly lower creatinine clearance resulted in a significantly higher percentage of filtered solute which was excreted for Group A as calculated from the ratio Cosm/ $C_{Cr} \times 100$ (mean value 1.53 for Group A, 1.10 for Group B, t = 2.98, p < 0.01). However, the maximal urine osmolality following fluid deprivation was not correlated with the C_{Osm}/C_{Cr} ratio.

Hypokalemia, hypercalcemia, and sickle cell disease, conditions known to be associated with impaired concentrating ability, were not present in these subjects. It is emphasized that women with bacteriuria as a group could not be distinguished from those without bacteriuria on the basis of history, physical examination (including blood pressure), pyuria, or proteinuria (8).

TABLE II
Data for Group B subjects without significant bacteriuria

Serum oserum serum serum serum serum ality Solute‡ Creat.‡ Com‡ Co‡ a 3.48					Final CV	Final CVS after fluid deprivation	ion	Serum after fluid deprivation	after ivation	Urine/	24-Ho	ur urine	during fl	24-Hour urine during fluid deprivation	ation
Bact./ml mO_{SPM} mO_{SPM} mO_{SPM} mu_{S}	Age Grav. Para. Wks† Date Culture D	Random CVS* Date Culture	l lean		ate	Culture	Osmol- ality	Osmol- ality	Creat- inine	serum osmol- ality	Solute‡	Creat.‡	Cosm‡	Cort	Xǰ™ X100
59,000 78,175 48 134 48 134 48 150 48 134 48 150 <th< td=""><td>yrs Bact./ml</td><td>Bact./ml</td><td>Bact./ml</td><td></td><td></td><td>Bact./ml</td><td>/mosm/</td><td>1</td><td>mg/</td><td></td><td>/wsom</td><td>/šm</td><td>/lm</td><td>min</td><td></td></th<>	yrs Bact./ml	Bact./ml	Bact./ml			Bact./ml	/mosm/	1	mg/		/wsom	/šm	/lm	min	
33,000 753 294 295 3.13 3.13 3.13 3.13 3.13 3.13 3.13 3.13 3.13 3.13 3.13 3.13 3.13 3.13 3.13 3.13 3.13 3.14 <	27 5 3 21 1/27/59 3,000 2/ 20 1 22 12/30/58 2,000 1/ 20 1 0 14 12/16/58 2,000 1/ 22 1 0 21 2/10/59 16,000 2/ 27 3 2 11 2/17/59 8,000 2/	1/27/59 3,000 12/30/58 2,000 12/16/58 2,000 2/10/59 16,000 2/17/59 8,000		22112	6/59 14/59 6/59 21/59 26/59	59,000 1,000 7,000 11,000 73,000 A. aerogenes	954 954 984 936 1,173		# 000	3.48 3.42 3.28 3.94 3.33	r: ::	z z			
1,000 746 296 0.64 2.52 436 0.81 1.47 127 4,000 937 298 1.26 3.40 289 1.71 111 <t< td=""><td>24 2 1 17 3/ 3/59 606 3/ 27 3 1 20 3/10/59 4,000 3/ 24 3 2 14 4/14/59 3,000 4/ 29 2 2 3 4/14/59 2,000 4/ 30 5 4 16 3/21/60 1,000 3/</td><td>3/ 3/59 606 3/10/59 4,000 4/17/59 3,000 4/14/59 2.000 3/21/60 1,000</td><td></td><td>ww44w</td><td>19/59 17/59 21/59 22/59 29/60</td><td>23,000 39,000 76,000 E. coli 57,000 E. coli 2,000</td><td>753 924 999 891 926</td><td>297 295 310 297 301</td><td>0.69</td><td>2.54 3.13 3.22 3.00 3.08</td><td>442</td><td>0.93</td><td>1.47</td><td>135</td><td>1.09</td></t<>	24 2 1 17 3/ 3/59 606 3/ 27 3 1 20 3/10/59 4,000 3/ 24 3 2 14 4/14/59 3,000 4/ 29 2 2 3 4/14/59 2,000 4/ 30 5 4 16 3/21/60 1,000 3/	3/ 3/59 606 3/10/59 4,000 4/17/59 3,000 4/14/59 2.000 3/21/60 1,000		ww44w	19/59 17/59 21/59 22/59 29/60	23,000 39,000 76,000 E. coli 57,000 E. coli 2,000	753 924 999 891 926	297 295 310 297 301	0.69	2.54 3.13 3.22 3.00 3.08	442	0.93	1.47	135	1.09
270 945 289 0.72 3.27 331 0.86 1.15 1.19 8 1,036 286 266 3.67 497 1.17 1.55 2,000 885 298 1.11 2.97 665 1.27 1.56 114 1,000 830 297 0.67 2.76 270 0.84 0.91 161 1,000 830 297 0.75 2.86 4.24 1.26 1.27 1.56 114 448 859 209 0.87 2.86 4.24 2.86 3.49 0.65 1.27 1.25 4,000 1,004 287 0.87 2.86 3.49 0.63 1.20 7.2 4,000 1,044 299 0.87 2.86 0.96 3.24 474 1.09 1.36 4,000 1,000 2.97 0.87 2.44 1.09 1.36 1.21 1,000 2.87 <td< td=""><td>20 3/24/60 2,000 24 3/24/60 6,000 20 10/4/60 16,000 18 10/14/60 2,000 20 10/19/60 1,000</td><td>3/24/60 2,000 3/24/60 6,000 10/4/60 16,000 10/14/60 2,000 10/19/60 1,000</td><td></td><td>£,5000</td><td>29/60 30/60 19/60 11/60</td><td>1,000 4,000 23,000 118 3,000</td><td>746 827 983 900 1,129</td><td>296 298 289 318</td><td>0.64 0.93 1.26 0.75 1.07</td><td>2.52 2.78 3.40 3.02 3.55</td><td>436 509 289 347 467</td><td>0.81 1.03 1.39 0.81 1.03</td><td>1.47 1.71 1.00 1.16</td><td>127 111 110 108 96</td><td>1.16 1.54 0.91 1.07</td></td<>	20 3/24/60 2,000 24 3/24/60 6,000 20 10/4/60 16,000 18 10/14/60 2,000 20 10/19/60 1,000	3/24/60 2,000 3/24/60 6,000 10/4/60 16,000 10/14/60 2,000 10/19/60 1,000		£,5000	29/60 30/60 19/60 11/60	1,000 4,000 23,000 118 3,000	746 827 983 900 1,129	296 298 289 318	0.64 0.93 1.26 0.75 1.07	2.52 2.78 3.40 3.02 3.55	436 509 289 347 467	0.81 1.03 1.39 0.81 1.03	1.47 1.71 1.00 1.16	127 111 110 108 96	1.16 1.54 0.91 1.07
21,000 820 297 0.67 2.76 2.76 0.84 0.91 1.25 1,000 839 291 0.73 2.88 4.24 6.91 1.25 448 859 308 1.08 2.79 2.39 0.98 0.78 91 23,000 966 2.87 0.71 3.37 364 0.93 1.20 72 46,000 1.044 2.99 0.63 3.24 474 1.09 1.59 121 1,000 967 2.98 0.90 3.24 474 1.09 1.59 121 1,000 2.94 2.99 0.63 3.24 474 1.09 1.59 121 1,000 2.93 0.90 2.50 2.43 0.80 0.84 89 1,000 2.93 0.69 3.07 3.43 0.91 1.30 1.20 1,000 2.93 0.89 0.80 3.07 3.81 0.94	20 2 1 24 10/20/60 1,400 10/2 30 3 1 15 10/11/60 EMB neg.\$ 10/2 20 1 0 17 10/18/60 1,000 10/2 30 5 23 10/4/60 1,000 10/1 27 3 2 27 10/18/60 1,000 10/1	10/20/60 1,400 10/11/60 EMB neg.\$ 10/18/60 1,000 10/4/60 7,000 10/18/60 1,000		10/2	2/80 0/00 0/00 0/00	270 8 52 22,000 600	945 1,050 923 885 916	289 286 315 298 295	0.72 0.66 0.81 1.11 0.62	3.27 3.67 2.93 2.97 3.11	331 497 615 465 282	0.86 1.02 1.24 1.27 1.00	1.15 1.74 1.95 1.56 0.96	119 155 153 114 161	0.97 1.12 1.27 1.37 0.60
23,000 966 287 0,71 3.37 364 0.98 1.27 138 46,000 10,44 299 0,63 3.49 168 0,52 0,56 83 12 964 298 0,90 3.24 474 1.09 1.59 1.59 1,000 293 0,90 2.50 243 0,80 0,84 89 935 293 0,81 3.16 387 0,97 1.30 120 118 10 0.18 0,40 124 0,22 0,41 27	27 3 2 17 10/27/60 4,000 11/ 21 1 10/14/60 9,000 10/2 20 5 4 17 10/25/60 232 10/2 20 2 1 15 10/35/60 838 11/ 25 4 3 30 11/18/60 148 11/2	10/27/60 4,000 10/14/60 9,000 10/25/60 232 10/31/60 838 11/18/60 148		110/2	1/60 6/60 1/60 1/60	21,000 1,000 68 448 1,000	820 839 1,242 859 829	297 291 293 308 290	0.67 0.73 0.66 1.08 0.87	2.76 2.88 4.24 2.79 2.86	270 239 349	0.84 0.98 0.63	0.91 0.78 1.20	125 91 72	0.73 0.86 1.67
293 0.81 3.16 387 0.97 1.30 120 10 0.18 0.40 124 0.22 0.41 27	26 6 3 18 11/21/60 240 11/2 30 5 4 26 11/18/60 19,000 11/2 11 30 11/23/60 84 11/2 28 5 4 25 11/25/60 1,000 12/ 26 3 2 30 11/28/60 1,000 12/	11/21/60 240 11/13/60 19,000 11/23/60 84 11/25/60 1,000 11/28/60 1,000		11/2	3/60 1/60 1/60 6/60	23,000 46,000 2 1,000	966 1,044 967 724 900	287 299 298 290 293	0.71 0.63 0.90 0.90	3.37 3.49 3.24 2.50 3.07	364 168 474 243 575	0.98 0.52 1.09 0.80	1.27 0.56 1.59 0.84 1.96	138 83 121 89 177	0.92 0.67 1.31 0.94 1.11
10 0.18 0.40 124 0.22 0.41 27	25 3.1 1.9 21						935	293	0.81	3.16	387	0.97	1.30	120	1.10
							118	10	0.18	0.40	124	0.22	0.41	27	0.35

* † ‡ See footnotes to Table I. § No growth on eosin-methylene-blue agar plate.

TABLE III Data for Group C subjects with high-count urine cultures, probably due to contamination

				Random CVS*	Fina	Final CVS after fluid deprivation		fluid deprivation		
Grav. P	Para.	Wks†	Date	Culture	Date	Culture	Osmol- ality	Osmol- ality		
				Bact./ml		Bact./ml	/mosm/	mOsm/		1
•	~	24	$\frac{1}{6/59}$	193,000 Proteus 344,000 diphth.	1/13/59	248,000 Staph.	1,125	300	3.75	
	_	17	$\frac{1/27}{59}$	144,000 diphth. 100,000 <i>Skaph</i> .	2/ 5/59	634,000 diphth.	1,035	280	3.70	
	_	23	2/3/59	14,000	2/10/59	108,000 <i>Staph.</i> & diphth.	1,131	292	3.87	
•	8	25	2/24/59	143,000 A. aerogenes	3/ 7/59	162,000 diphth.	1.110	293	3.70	
	_	24	2/24/59	2,000	3/4/59	123,000 Staph.	1,065	297	3.59	
•	4	17	3/24/59	566,000 Staph.	4/ 1/59	315,000 diphth.	1,146	283	4.05	
		20	4/28/59 5/20/59 6/3/59 6/22/59	117,000 A. aerogenes 37,000 E. coli 50,000 A. aerogenes 16,000 diphth.	5/12/59	101,000 A. aerogenes & diphth.	1,050	298	3.52	
		17	7/20/59	459,000 yeasts, Staph. & diphth.	7/22/59	over 10° yeasts, Staph. & diphth.	1,098	288	3.81	
_	0	17	9/16/59	31,000 Staph.	9/20/29	151,000 Staph.	1.010	288	3.51	
	1	70	10/13/60	3,000	10/15/60	128,000 diphth.	949	294	3.23	
	7	10	10/ 4/60	EMB‡ 1 colony E. coli	10/19/60	115,000 <i>Staph.</i> & diphth.	788	291	2.71	
		27	10/11/60	71,000 diphth.	10/19/60	over 10° Staph. & diphth.	1,000	297	3.37	
	_	19	1/20/59	700,000 diphth.	1/29/59	39,000	996	280	3.45	
- ,	7	29	12/ 9/58	388,000 Enterococci	12/18/59	4,000	1,020	300	3.40	
	-0	24 17	2/10/59 2/17/59	107,000 A. aerogenes 124,000 A. aerogenes	2/16/59 2/24/59	38,000 37,000 <i>Staph</i> . & yeasts	1,074 1,209	294 290	3.65	
	0	70	2/11/59	114,000 Staph.	2/25/59	76,000 A. aerogenes & Staph.	993	302	3.29	
	7	19	2/ 3/59	133,000 Staph.	2/11/59	25,000 Staph. & A. aerogenes	744	292	2.55	
		24	$\frac{10}{25}/60$ $\frac{10}{31}/60$	over 10° E. coli over 10° E. coli	11/8/60	1,000	835	289	2.89	
	4	16	11/22/60	2,000	11/30/60	312,000 Staph. & diphth.	669	296	2.36	
2.6	1.4	21				•	1,002 139	292 6	3.43	
* † See footnotes to Table I.				‡ Eosin-methylene-blue ag	ar plate.					1
	6 dable I.		1	1 17 2 25 1 24 4 17 1 20 1 20	1/28/59 1 17 1/28/59 1 23 2/3/59 2 25 2/24/59 4 17 3/24/59 1 24 2/24/59 1 24 2/24/59 1 20 4/28/59 1 1 20 4/28/59 0 17 9/16/59 1 17 1/20/59 2 10 10/4/60 1 27 10/17/60 1 24 2/10/59 0 20 2/17/59 2 19 2/3/59 1 24 10/25/60 1 24 10/25/60 1 24 10/25/60 1 24 11/20/59 1 24 2/10/59 1 24 2/10/59 1 24 2/10/59 1 24 2/10/59 1 24 2/17/59 1 24 10/25/60 1 14 21 11/22/60	1 178/59 344,000 diphth. 1 1/27/59 144,000 diphth. 2 25 2/24/59 144,000 diphth. 2 25 2/24/59 144,000 4. aerogenes 1 24 2/24/59 2,000 4 17 3/24/59 2,000 2 17 3/24/59 37,000 E. coli 2 4/28/59 117,000 A. aerogenes 6/22/59 37,000 E. coli 1 17 7/20/59 37,000 yeasts. 2 10 10/4/60 EMB‡ 1 colony E. coli 1 27 10/17/60 71,000 diphth. 2 2 9 12/9/58 388,000 Enterococci 1 24 2/10/59 107,000 A. aerogenes 0 20 2/17/59 114,000 Staph. 2 2 9 12/9/58 388,000 Enterococci 1 24 2/10/59 107,000 A. aerogenes 0 20 2/17/59 114,000 Staph. 1 24 10/25/60 over 10° E. coli 1 24 10/25/60 over 10° E. coli 1 24 2/3/59 133,000 Staph. 1 24 10/25/60 over 10° E. coli 1 24 21 16 11/22/60 2,000 1.4 21 ‡ Eosin-methylene-blue agar	1 17 1/28/59 344,000 diphth. 1 23 2/27/59 114,000 diphth. 2 22 25 2/24/59 144,000 diphth. 2 25 2/24/59 140,000 Skaph. 2 25 2/24/59 143,000 A. aerogenes 3/7/59 4 17 3/24/59 2,000 2 25 2/24/59 2,000 3 4/59 4 17 3/24/59 31,000 A. aerogenes 5/12/59 6/23/59 31,000 A. aerogenes 5/12/59 6/23/59 117,000 A. aerogenes 5/12/59 1	1/28/59 344,000 diphth. 1 23 2/3/59 144,000 diphth. 2 25 2/24/59 144,000 diphth. 2 25 2/24/59 143,000 4. aerogenes 3/759 162,000 diphth. 2 25 2/24/59 143,000 4. aerogenes 3/759 123,000 diphth. 3/24/59 2,000 Saph. 4/159 123,000 diphth. 3/24/59 117,000 4. aerogenes 3/12/59 113,000 diphth. 2 4/28/59 117,000 4. aerogenes 5/12/59 113,000 diphth. 3/24/59 37,000 E. cohi & E. Cohi	1 17 1/28/59 344,000 diphth. 1 17 1/28/59 144,000 diphth. 2 12 1/27/59 140,000 Slaph. 2 12 2/24/59 140,000 Slaph. 2 12 2/24/59 140,000 Slaph. 3 1/300 Slaph. 4 17 3/24/59 2,000 4 17 3/24/59 31,000 E.colic serveness 3/7/59 153,000 Slaph. 1 1 20 4/28/59 117,000 E.colic serveness 5/12/59 101,000 A. aerogeness 1,050 6/22/59 110,000 A. aerogeness 5/12/59 101,000 A. aerogeness 1,050 6/22/59 110,000 Slaph. 2 10 10/20/59 450,000 Slaph. 3 10 10 10/20/59 450,000 Slaph. 3 10 10 10/20/59 110,000 Slaph. 3 11,000 Slaph. 4 16 11,122/60 2,000 11,300 Slaph. 4 16 11,22/60 2,000 Slaph. 5 12,32/32 Slaph. 5 12,32	1 17 1/28/59 344,000 diphth. 2 / 5/59 634,000 diphth. 1,035 280 2/27/59 144,000 diphth. 2 / 5/59 634,000 diphth. 1,035 280 2/27/59 100,000 Supht. 2 / 10,000 Supht. 2 / 14,000 diphth. 1,110 293 2 / 2/24/59 2 / 2,000 3 / 4/59 115,000 Supht. 1,110 293 3 / 2/24/59 2 / 2,000 Supht. 3 / 2,000 Supht. 1,146 283 5 / 2,000 Supht. 3 / 2,000 Supht. 4 / 2,000 Supht. 4 / 2,000 Supht. 4 / 2,000 Supht. 3 / 2,000 Supht. 3 / 2,000 Supht. 3 / 2,000 Supht. 3 / 2,000 Supht. 4 / 2,000 Supht. 3 / 2,000 Supht. 3 / 2,000 Supht. 3 / 2,000 Supht. 3 / 2,000 Supht. 4 / 2,000 Supht. 4 / 2,000 Supht. 3 / 2,000 Supht. 4 / 2,000 Supht. 5 /

TABLE IV Routine urinalysis at first prenatal clinic visit for Group A subjects with bacteriuria (see Table I)

						Urinary se	diment	
Subject	Date	Sp gr	Protein*	Sugar†	WBC	RBC	Casts	Bact.‡
						no./high-pou	vered field	
1A	11/18/58	qns	0	0	8-16	0	0	4+
2A	11/18/58	1.022	0	0	0	0	0	3+
3A	11/18/58	1.012	0	0	occ.	0-2	0	3
4A	12/ 9/58	1.020	0	0	occ.	occ.	0	3+
5A	12/30/58	qns	0	0	1-3	occ.	0	4+
6A	1/13/59	1.023	0	0	occ.	0	0	3+
7A	1/13/59	1.015	0	0	0	0	0	0
8A	3/31/59	1.012	0	0	0	0	0	4+
9A	4/ 7/59	1.012	0	0	1-3	0	0	4+
10A	4/ 7/59	1.020	0	0	8-10	0	0	1+
11A	6/30/59	1.020	0-tr.	0	occ.	0	0	3+
12A	12/ 8/59	1.007	0	0	20-25	occ.	0 .	1+
13A	1/26/60	1.028	0 -	0	0	0	0	0
14A	2/ 9/60	1.008	0	0	0	0	0	1+
15A	2/23/60	1.011	0	0	0	0	0	1+
16A	4/5/60	1.010	0	0	0	0	0	4+
17A	4/12/60	1.014	0	0	0	0	0	3
18A	9/27/60	1.016	0	0	many	0	0	2
19A	11/29/60	1.016	0	0	14-16	0	0	1+
20A	12/15/60	qns	1-2+	0	many	0–1	0	$\overline{4} +$

^{*} Prior to 1960, beta naphthalene sulfonic acid precipitation method; since 1960, using Uristix. † Prior to 1960, using Clinitest; since 1960, using Uristix. ‡ Graded qualitatively, 0 to 4+.

Routine analyses at the first clinic visit for the 20 Group A subjects with bacteriuria are recorded in Table IV. Cylinduria was absent, pyuria was variable, and slight proteinuria was present in 2 women.

DISCUSSION

The validity of classifying subjects as having bacteriuria or not on the basis of bacterial counts in clean-voided specimens deserves comment. The basis for such a classification rests on the assumption that if bacteria are present within the urinary tract they will multiply to large numbers in the bladder urine which serves as a good culture medium. In contrast, bacteria added to sterile urine as contamination in clean-voided specimens usually are considerably fewer in number. In general, this hypothesis has been verified (4-6, 11-20), and exceptions clearly outlined (21), but agreement has not been universal (22, 23). The choice of the number of bacteria as a dividing line between bacteriuria and contamination is based on statistical probabilities. In agreement with many other workers in this field, I have found 100,000 bacteria per ml to represent a reasonable dividing line sufficiently sensitive to include most instances of bacteriuria and yet discriminatory against most instances of contamination. For example (8), in 33 pregnant women with over 100,000 gramnegative bacteria per ml in an initial random, cleanvoided urine specimen, a second clean-voided urine culture gave the same result in 21 women (63.6 per cent). After two successive cultures showing over 100,000 gram-negative bacteria per ml, a third culture, or more, gave the same result in over 85 per cent of the women. Thus, the chances that women in Group A in this study, with a minimum of two and usually three separate cultures containing over 100,000 gram-negative bacteria per ml, would remain in this high-count group with additional cultures would be about 85 per cent. The high-count urine cultures for subjects in Group C, however, were considered to represent contamination, since additional cultures were not confirmatory. The finding of normal concentrating ability in these Group C subjects is in accord with this interpretation.

The finding of definite impairment in concentrating ability (maximal urinary osmolality less than 700 mOsm per kg H₂O) with fluid deprivation in 9 of 20 pregnant women with asympto-

matic bacteriuria may be a reflection of early silent pyelonephritis in these subjects. In Subject 9A, with bacteriuria and a maximal urine osmolality of 570 mOsm per kg H₂O during pregnancy, studies 7 months post partum revealed improvement in concentrating ability to 800 mOsm per kg H₂O in association with spontaneous cessation of bacteriuria. However, symptomatic, acute pyelonephritis developed in this subject 8 months post partum, which was not treated, and at 8.5 months post partum maximal urinary osmolality was again reduced to 620 mOsm per kg H₂O with return of E. coli bacteriuria. Treatment to suppress bacteriuria then resulted in improvement in maximal urinary osmolality to 870 mOsm per kg H₂O at 10 months post partum. In Subject 8A, both bacteriuria with E. coli and impaired concentrating ability persisted post partum (maximal urine osmolality 636 mOsm per kg H₂O during pregnancy, 725 mOsm 2 months post partum). Spontaneous clearing of bacteriuria post partum in Subject 15A was associated with improvement in concentrating ability from a maximal urinary osmolality of 723 mOsm per kg H2O during pregnancy to 1,000 mOsm 3.5 months post partum. These few serial studies suggest that impaired concentrating ability may be a reversible result of infection rather than a pre-existing, fixed defect which predisposes to infection. Similarly, Winberg (3) has reported that impaired concentrating ability occurring with acute, nonobstructive urinary tract infections in children possibly lasted for 4 to 6 weeks after symptoms and urinary findings cleared. The presence of silent pyelonephritis in pregnant women with asymptomatic bacteriuria is also suggested by their high incidence of the subsequent development of acute pyelonephritis (7, 8). Impaired concentrating ability with bacteriuria may also be attributed to an increase in the hydronephrosis of pregnancy due to ureteritis and ureteral reflux in the absence of renal infection, but a clear distinction between pyelonephritis and infection limited to the ureter cannot easily be made.

It is possible that screening for both bacteriuria and impaired concentrating ability might effectively select those individuals destined to develop acute pyelonephritis of pregnancy. This is supported by the subsequent development of acute pyelonephritis in Subjects 9A, 10A, 12A and

18A, all with some degree of impairment of concentrating ability and bacteriuria, whereas in the remaining subjects in Group A, symptomatic pyelonephritis during pregnancy did not occur. If the incidence of bacteriuria among pregnant women is taken as 5 per cent (7, 8) and 40 per cent of such women have impaired concentrating ability, then silent pyelonephritis might be present in about 2.0 per cent of all pregnant women. Long-term studies are clearly indicated to determine whether chronic pyelonephritis develops in pregnant women with asymptomatic bacteriuria.

The mechanism by which impaired concentrating ability occurs with asymptomatic bacteriuria is not clear from the limited data obtained. creatinine clearance values are somewhat low in both Groups A and B. This may reflect incompleteness of the collections and use of a less specific method for serum creatinine or the effect of upright posture during most of the collection period. In view of the relatively normal serum creatinine concentrations and endogenous creatinine clearances, severe reduction in glomerular filtration rate (GFR) appears unlikely, although it is possible that one kidney may have been affected more than the other. It is known that small reductions in filtration rate may affect the concentrating mechanism (24). The slightly higher solute excretion in bacteriuric women may be an artifact, since it is not certain that all women kept the 24 hour urine collection refrigerated. Osmolality of urine specimens left at room temperature has been observed to rise and this process may be accelerated in the presence of large numbers of bacteria. Additional and serial renal function studies are necessary to define the interrelation of GFR, solute excretion and concentrating ability in subjects with bacteriuria.

Calculation of food intake was made by dietitians for all the women in this study and showed no difference for women with and without bacteriuria. Weight gain during pregnancy was comparable for both groups. Thus, it is unlikely that protein deprivation was an important factor in the development of impaired concentrating ability.

An experimental model was sought to study the effects of renal infection on the concentrating mechanism. Bacteriuria and pyelonephritis were produced by the direct injection of a strain of *Proteus* into the urinary bladder of the rat (25).

Impaired concentrating ability was observed, but the animals were severely uremic. Attempts to produce bacteriuria without altering GFR in order to study the direct effects of infection on the concentrating mechanism have not yet been successful. Even in experimental, unilateral pyelonephritis, which prevents the development of uremia, the involved side has shown reduction in filtration rate (26).

SUMMARY

- 1. Pregnant women with asymptomatic bacteriuria were found to have significant impairment of urinary concentrating ability when compared with a control group without bacteriuria.
- 2. Impaired concentrating ability most likely is secondary to silent, active pyelonephritis.
- 3. The mechanism of the impairment remains to be determined. Direct involvement of the renal concentrating mechanism by infection, an increase in the hydronephrosis of pregnancy due to infection and malfunction of the ureters, and changes in the concentrating mechanism secondary to slight reductions in glomerular filtration rate may be important factors.

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