

STUDIES OF THE PRINCIPLE IN LIVER EFFECTIVE IN PERNICIOUS ANEMIA. IV. THE THERAPEUTIC ACTIVITY OF ITS MULTIPLE FACTORS ^{1,2}

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Early in the course of an investigation of the principle in liver effective in pernicious anemia certain observations suggested the possibility that the therapeutic activity of liver extract might depend upon the presence of several chemically distinct substances. The evidence for this view was the fact that continued purification of therapeutically active liver extract, in the absence of significant losses and of destructive procedures, resulted in partial or complete extinction of therapeutic activity. On the other hand, the admixture with such highly purified materials of other fractions derived from liver extract resulted in the recovery of therapeutic activity. These latter fractions we have termed *accessory* factors, for they appeared to augment the activity of the *primary* factor (or factors), while in the absence of the primary factor they were therapeutically inert. This communication describes in detail the observations mentioned above, part of which have already been presented in a preliminary report (1).

METHODS AND MATERIALS

The patients studied were suffering from classical Addisonian pernicious anemia in relapse. Complications such as severe combined system disease and hemorrhage were absent. No infections occurred during the periods of observations, except for cystitis in three patients (Cases 7, 9, and 12, Table I). During the prolonged periods

of study the patients' diet was of a mixed type, including meat or fish once daily, but devoid of yeast, liver, kidney, and tripe. Prior to the administration of therapeutically active fractions the blood level of each patient was established by adequate control periods, as noted in Table I.

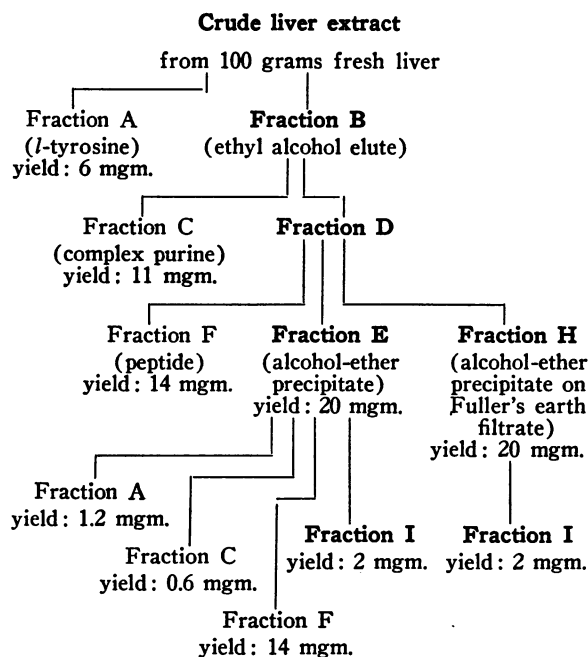


FIG. 1. DERIVATION OF PURIFIED LIVER FRACTIONS

The percentage of reticulocytes in the capillary blood was estimated by the wet method, 1000 erythrocytes being counted. Erythrocyte, hemoglobin, and hematocrit determinations were made on venous blood drawn without stasis, and rendered incoagulable by heparin. The number of erythrocytes was determined with pipettes and counting-chambers certified by the Bureau of Standards. The volume of packed red blood cells was measured in the Wintrobe hematocrit. The hemoglobin was determined by the Stadie-Wu method (12). The reticulocytes were counted

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TABLE I
Data of Figure 2

Curve	Patient	Length of period	Date of period	Fractions administered					Calculated amount of primary fraction administered per day	Rate of administration	Control period preceding administration
				E	H	A	C	F			
		days		total number grams fresh liver from which extract derived					grams fresh liver from which extract derived		days
1	1	20	Dec. 18, 1935 to Jan. 7, 1936	200		100	20		10	Every 2d day	1
2	2	30	April 16 to May 16, 1936		900	0	0	900	30	Every 10th day	2
3	3	28	April 17 to May 15, 1936		300	300	300	0	11	Every 10th day	16†
4	3	15	April 2 to April 17, 1936		370	0	0	0	25	Single	1
5	4	30	Dec. 28, 1935 to Jan. 27, 1936	300		110	200		10	Every 2d day	10‡
6	4	9	Dec. 19 to Dec. 28, 1935	100		0	0		11	Every 2d day	1
7	5	14	Mar. 20 to April 3, 1936		100	0	0	100	7	Single	1
8	1	20	Feb. 5 to Feb. 25, 1936	200		*	200		10	Every 2d day	50‡
9	6	18	Dec. 9 to Dec. 27, 1935	240		0	0		13	Every 2d day	10
10	7	8	April 30 to May 8, 1936		100	100	100	100	13	Single	7
11	4	10	Jan. 27 to Feb. 6, 1936	100		100	100		10	Every 2d day	40‡
12	8	24	Sept. 27 to Oct. 21, 1935	240		240	240		10	Every 2d day	3
13	9	8	May 1 to May 9, 1936		100	100	100	100	13	Single	11
14	1	20	Jan. 7 to Jan. 27, 1936	200		300	200		10	Every 2d day	21‡
15	10	10	June 5 to June 15, 1936		100	100	100	100	10	Single	75
16	11	21	Oct. 11 to Nov. 1, 1935	200		200	200		10	Every 10th day	6
17	12	41	Oct. 2 to Nov. 12, 1936		600	600	600	600	15	Every 2d day during first 20 days, every 10th day thereafter	29
18	13	14	Oct. 25 to Nov. 8, 1935	100		100	100		7	Single	22
19	2	27	May 16 to June 12, 1936		300	300	300	300	11	Every 10th day	32‡
20	1	16	Feb. 25 to Mar. 12, 1936	100		100	100		6	Every 2d day during first 8 days	70‡
21	1	16	Mar. 23 to April 8, 1936	100		†	100		6	Every 2d day during first 8 days	97‡
22	6	12	Dec. 27, 1935 to Jan. 8, 1936	170		170	170		14	Every 2d day	28‡

* 100 mgm. Fraction A orally daily.

† 1.0 gram Fraction A orally daily.

‡ Includes preceding experimental period.

daily and the venous blood constituents were studied, in most cases, on alternate days.

The derivation from commercial liver extract (Fraction G of Cohn et al. (2)) of the materials discussed in this communication is presented in Figure 1. In this figure the fractions containing the primary factor are depicted by heavy type. They are Fractions B, D, E, H, and I. The details of the preparation of Fractions A, B, and C have been previously described (3, 4). From Fraction D the primary factor has been brought down in both of the amorphous fractions E and H. From Fraction D, also, was separated Fraction F (an accessory factor) by precipitation with rhodanilic acid (5). The sources of primary factor discussed in this communication are Fraction E and Fraction H. While Fraction E contains both primary factor and Fraction F, as well as very small amounts of Fractions A and C, Fraction H, although amorphous, is completely devoid of Fractions A, C, or F. Fraction I was obtained from either Fraction E or from Fraction H, by precipitation with Reinecke salt and subsequent regeneration, as a microcrystalline sulfate in a yield of 2 mgm. from 100 grams of fresh liver (6). Studies of the therapeutic action of Fraction I will be presented in a later communication.

The accessory factors are represented by Fractions A, C, and F. Fraction A, obtained in a yield of 6 mgm. from 100 grams of liver, has been identified as *l*-tyrosine (3). In most of the experiments recorded below crystalline commercial *l*-tyrosine (Kahlbaum or Eastman Kodak) has been employed instead of Fraction A isolated from liver. Fraction C, obtained in a yield of 11 mgm. from 100 grams of liver, has been identified as a complex purine (3) and was administered in a crystalline state. Fraction F, a peptide, was prepared either directly from Fraction D or from Fraction E, in a yield of 14 mgm. from 100 grams of liver (5), and has been administered as a regenerated crystalline rhodanilate.

Throughout the experiments recorded below Fractions E (with the exception of the material administered to Patient 10, Table I) and H have been obtained from the same original supply of each fraction prepared in a large amount. The following quantities of the various fractions have been administered, as derived from 100 grams of

fresh liver: Fraction A, 11 mgm.; Fraction C, 6 mgm.; Fraction F, 14 mgm.; Fraction E, 20 mgm.; and Fraction H, 20 mgm.

Unless otherwise noted, all of the fractions were sterilized by boiling for one minute on the weakly acid side (alkaline to methyl red and acid to phenol red), and were administered by intramuscular injection.

The therapeutic activity of the primary factor, with and without the accessory factors

The therapeutic activity of the primary factor *without*³ the three accessory factors, administered as either Fraction E or H, was studied in six patients during nine periods of observation. These fractions were administered during periods varying from nine to thirty days in length, in amounts derived from 7 to 30 grams of liver per day. The effects of these fractions upon erythrocyte production are depicted in the left-hand part of Figure 2. The primary factor as either Fraction E or H, *together with* the three accessory factors, was administered to eleven patients during thirteen periods of observation from eight to forty-one days in length, in amounts derived from 6 to 15 grams of liver per day. Included among these latter patients were four to whom had been previously administered primary factor without the accessory factors. In the right-hand part of Figure 2 are depicted the individual erythrocyte regeneration curves following the administration of the primary factor together with the three accessory factors. The fractions administered, the amounts, and other relevant data bearing upon these erythrocyte regeneration curves are noted in Table I.

It is evident from inspection of Figure 2 that, in the *absence* of the three accessory factors, the effect of either Fraction E or H upon erythrocyte production (with the exception of Curve 2; *vide infra*) was either slight and of short duration (Curves 1, 3, 4, 7, 9), or was entirely lacking (Curves 5, 6, 8). On the other hand, after the administration of the primary factor *together with* Fractions A, C, and F, the erythrocyte responses showed a high degree of activity.

³ Although Fraction E contains Fraction F, and very small quantities of Fractions A and C (Figure 1), for the purpose of the present discussion it is considered in conjunction with Fraction H. Fraction H is completely devoid of Fractions A, C, and F.

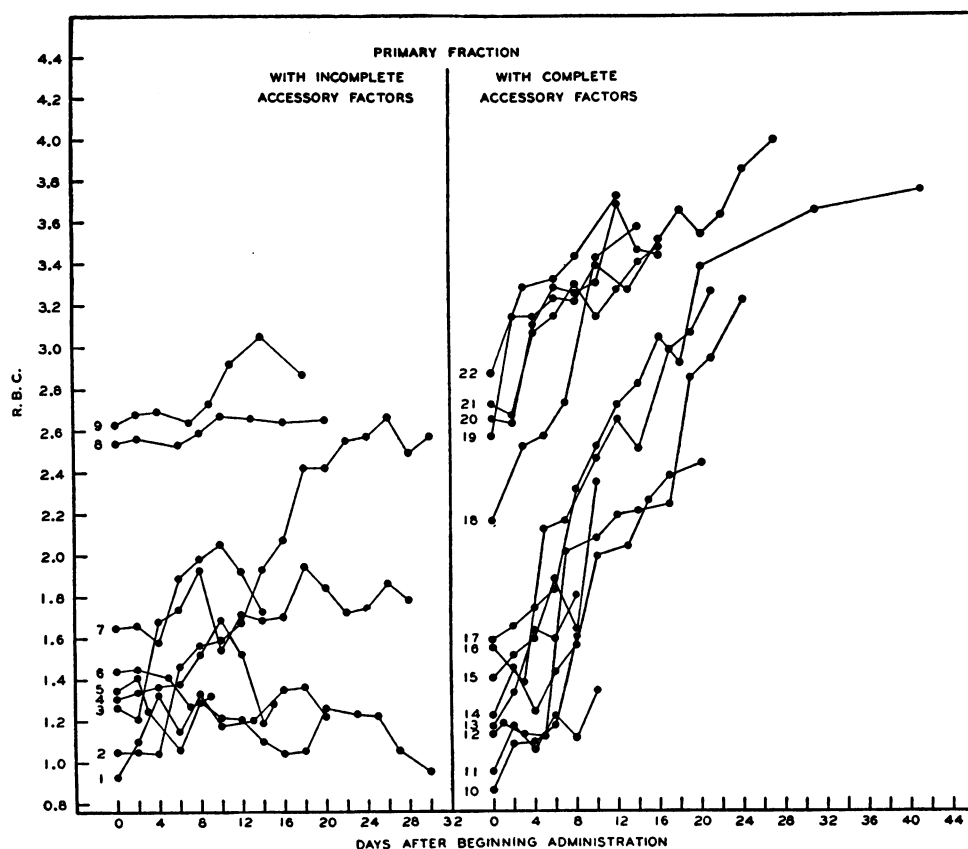


FIG. 2. ERYTHROCYTE REGENERATION CURVES FOLLOWING THE ADMINISTRATION OF PRIMARY FACTOR WITH INCOMPLETE AND WITH COMPLETE ACCESSORY FACTORS. FRACTIONS ADMINISTERED, QUANTITIES, AND OTHER DATA CONTAINED IN TABLE I

TABLE II
Comparative reticulocyte responses to primary factor with and without accessory factors

	Primary factor with incomplete accessory factors						Primary factor with complete accessory factors			
	E	H	H	E	H	E	H	E	E	H
Fraction.....	1	2	3	4	5	6	7	8	11	12
Patient.....	1	2	4	5	7	9	10	12	16	17
Curve number in Figure 2.....	100	300	370	100	100	200	100	100	100	200
Total amount fresh liver from which extract derived, grams.....	-20 grams every 2d day*	Single dose	Single dose	-20 grams every 2d day	Single dose	-40 grams every 2d day	Single dose	-20 grams every 2d day	Single dose	-40 grams every 2d day
Rate of administration.....	0.93	1.05	1.32	1.44	1.65	2.62	0.86	1.15	1.56	1.57
R.B.C. at beginning, millions per cu. mm.	1.19	1.58	1.69	1.32	2.05	2.73	1.60	2.08	2.46	2.52
R.B.C. at end, millions per cu. mm.....	7.8	31.0	16.4	3.0	9.8	4.4	24.6	25.0	23.8	18.1
Reticulocytes at peak, per cent.....	34	31	25	24	20	9	35	28	21	21
Predicted reticulocyte peak, per cent....	6th	6th	7th	6th	6th	8th	5th	7th	4th	8th
Day of reticulocyte peak.....	10	10	10	9	10	9	8	10	9	10
Length of period, days.....										

* The abbreviation -20 stands for "derived from 20 grams of liver."

All of the curves of Figure 2 are represented in the average erythrocyte regeneration curves of Figure 3. To facilitate interpretation the curves representing initial erythrocyte levels below 2.0 million have been averaged separately from those representing initial levels above 2.0 million. It is evident that the administration of the primary factor, together with Fractions A, C, and F, resulted in a rate of erythrocyte regeneration markedly greater than that produced by the primary factor in the absence of all three of the accessory factors, despite the administration, in the latter instance, of considerably larger quantities of pri-

mary factor. Indeed, the administration of both primary and accessory factors resulted in a rate of erythrocyte production closely approximating that induced by the intramuscular administration of an identical average daily dosage (derived from 11 grams of liver) of a commercial liver extract recently studied by Murphy (7).

Data concerning reticulocyte production following the administration of the primary factor *alone* were obtained in six patients, whose initial erythrocyte levels permitted a possible reticulocyte response. These observations are presented in Table II. For comparison there are included the

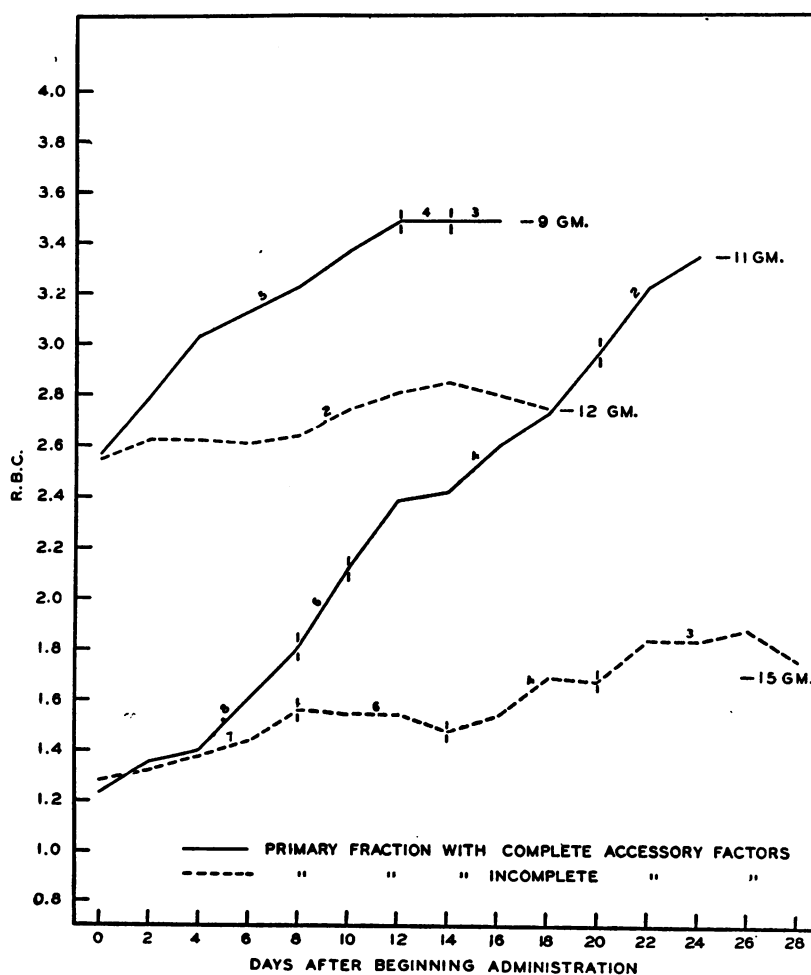


FIG. 3. AVERAGES OF ALL ERYTHROCYTE REGENERATION CURVES CONTAINED IN FIGURE 1, AT TWO DIFFERENT INITIAL ERYTHROCYTE LEVELS

The number of separate experiments included in each average is represented by the numerals directly above each curve; the periods corresponding to these numerals are defined by the small vertical lines. The quantity at the end of each curve denotes the calculated average daily amount, in terms of fresh liver from which the extract was derived, of primary fraction administered.

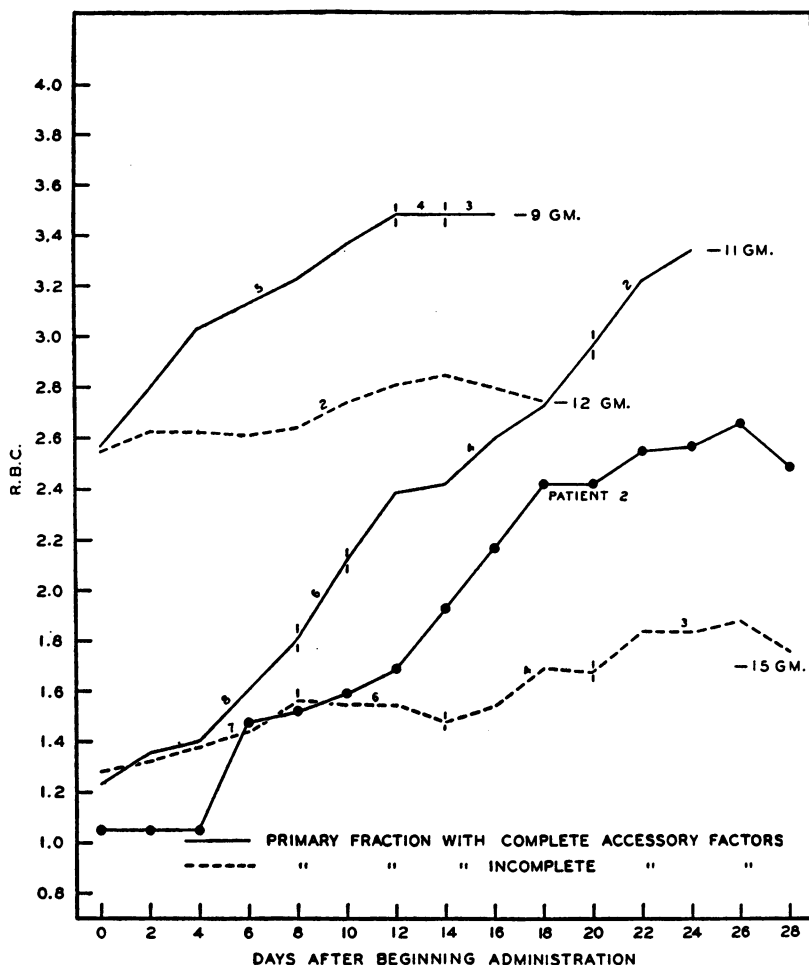


FIG. 4. ERYTHROCYTE RESPONSE TO ADMINISTRATION OF MASSIVE AMOUNT (— 300 GRAMS EVERY TENTH DAY) OF PRIMARY FACTOR WITH INCOMPLETE ACCESSORY FACTORS (PATIENT 2)

This erythrocyte regeneration curve is superimposed on the curves of Figure 3, for comparison with average response to administration of smaller amount of primary factor together with complete accessory factors. See text.

predicted reticulocyte peaks, based upon unpublished data of the authors, following the intramuscular injection of commercial liver extract derived from 100 grams of liver. It is evident that Fraction E or Fraction H, when administered in an amount derived from 100 or 200 grams of liver, during the first ten days, induced markedly submaximal reticulocyte responses, regardless of the rate of administration (single dose in Patient 5, divided doses in Patients 1, 4, and 6). After the administration of a massive amount of Fraction H (derived from 300 grams of liver) a maximal response followed in Patient 2, but an even

larger amount (derived from 370 grams of liver) administered to Patient 3 was followed by a submaximal response. On the other hand, in four patients treated with moderate amounts of Fractions E or H, *together* with the three accessory factors, the reticulocyte responses were approximately maximal. (See Table II.)

The effects of massive amounts of the primary factor with incomplete accessory factors

The therapeutic effects of large amounts of the primary factor *alone* were studied during a prolonged period in one patient. The erythrocyte

regeneration curve of this case (Patient 2) is included in Figure 2 (Curve 2), and is also presented in Figure 4, in order to facilitate comparison with the average erythrocyte regeneration curves of Figure 2. To this patient (Figure 4) at an initial erythrocyte level of 1.05 million, were administered Fractions H and F, each derived from 300 grams of liver, in a single dose. The same amount of each fraction was again administered on the tenth and on the twentieth days, totaling material derived from 900 grams of liver during thirty days. The reticulocyte response was maximal, reaching a peak of 31.0 per cent on the sixth day. During the first ten days the erythrocytes rose to 1.58 million, by the thirtieth day they reached a level of 2.5 million. It is apparent, from the data of Figure 4, that the erythrocyte response to the administration of large amounts (—30 grams per day)⁴ of the primary factor, without the three accessory factors, was not as great as the average erythrocyte response to smaller amounts (—11 grams per day) of primary factor together with the accessory factors. At the end of this period Fractions H, A, C, and F, each derived from 100 grams of liver, were administered to this patient, and the same dosage was repeated on the tenth and on the twentieth days. The erythrocytes continued to rise, reaching a count of 4.0 million on the twenty-seventh day (Curve 19, Figure 2).

The augmentative action of each accessory factor

In the studies described above all three of the accessory factors, Fractions A, C, and F, were administered together with the primary factor (Fraction E or H). That each of these fractions may act as an accessory factor was suggested by certain indirect evidence (1). Direct evidence for this view was furnished by the following observations.

The administration of an adequate amount of primary factor together with Fractions C and F, but with an incomplete amount of Fraction A, was followed, in two patients, by no erythrocyte response. To Patient 4 (Curve 5, Figure 2), at an initial erythrocyte level of 1.3 million, were administered on alternate days Fractions E — 20

grams (derived from 20 grams of fresh liver), C — 10 grams, and A — 2 grams, during the first ten days; during the second ten days the dosage on alternate days was E — 20 grams, C — 10 grams, and A — 10 grams; and during the last ten days, E — 20 grams, C — 20 grams, and A — 10 grams. It is seen that even after the increase of Fraction C, in the absence of a complete amount of Fraction A, during the last ten days, the erythrocytes continued to fall, reaching 0.95 million. To the patient were then administered on alternate days doses of Fractions E — 20 grams, C — 20 grams, and A — 20 grams, during the following ten days, and, as depicted in Curve 11, Figure 2, the erythrocytes rose to 1.34 million, accompanied by clinical improvement. Similar observations were made in Patient 1, at an initial erythrocyte level of 2.54 million. As shown in Figure 6, to this patient were administered on alternate days doses, during a period of twenty days, of Fractions E — 20 grams and C — 20 grams, as well as 100 mgm. of tyrosine (Fraction A), *orally*, daily. The erythrocytes remained stationary. In the next succeeding period the same amounts of Fractions E and C were continued, but Fraction A — 20 grams, was administered on alternate days by *intramuscular injection*. A satisfactory erythrocyte response promptly followed.

The accessory action of Fraction C, in the presence of an adequate amount of the other factors, was suggested by the following observations upon Patient 1, presented in Figure 5. During these observations over a period of forty days Fraction E — 20 grams was administered on alternate days. During the first twenty days Fraction C — 2 grams was also administered at the same rate. During the first ten days no additional Fraction A was injected (other than that contained in Fraction E; see Figure 1). It is seen that during this first period the reticulocytes rose to a peak of 9.2 per cent on the eighth day, followed by a rise of erythrocytes from 0.92 million to 1.2 million. Slight clinical improvement took place. During the second period of ten days, Fraction A — 20 grams, administered on alternate days, was added to the previous dosage of Fractions E and C. In response to these materials the reticulocytes did not continue to fall, but rose again to a peak of 9

⁴ The abbreviation — 30 grams per day stands for "derived from 30 grams of liver per day."

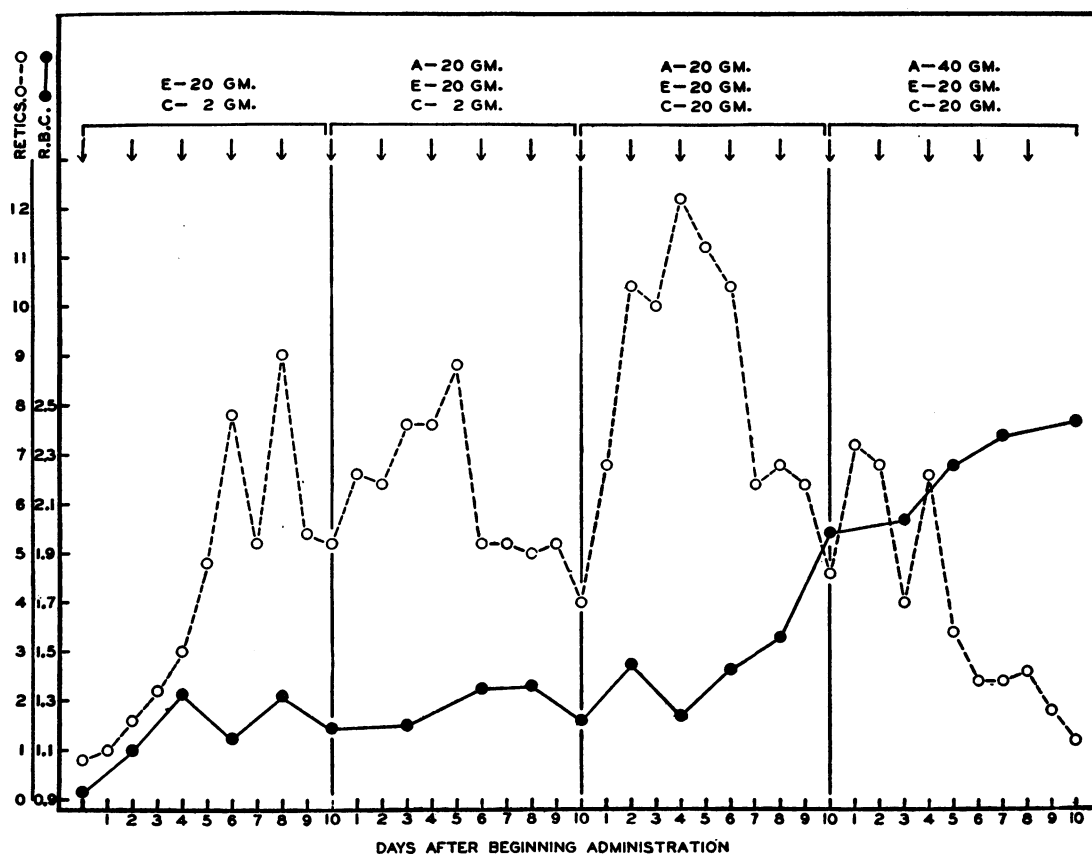


FIG. 5. PATIENT 1. FRACTION C AS AN ACCESSORY HEMATOPOIETIC FACTOR

The quantities above each period refer to the amounts of fractions injected on the days denoted by each arrow. Throughout the four periods Fraction E — 20 grams was administered on alternate days. During the first two periods Fraction C — 2 grams was administered on alternate days. There were only slight reticulocyte and erythrocyte responses in the first period in the absence of Fraction A and no further erythrocyte rise in the second period after addition of Fraction A — 20 grams on alternate days. The increase of Fraction C to — 20 grams in the third period resulted in the greatest rise of reticulocytes and satisfactory rise of erythrocytes, continuing in the fourth period.

per cent; the erythrocytes remained stationary, and the clinical condition was unchanged. During the third period the dosage of Fraction C on alternate days was increased from — 2 to — 20 grams, while the same amounts of Fractions E and A were continued. A third and distinctly orderly reticulocyte response, with a maximum of 12.4 per cent on the fourth day, was induced, and the erythrocytes rose from a level of 1.2 million to one of 2.0 million. Marked clinical improvement accompanied the changes in the blood. During the fourth period of ten days the continued administration on alternate days of the same fractions, except for an increase of the dosage of Fraction A from — 20 to — 40 grams, was ac-

companied by a continued rise of the erythrocytes. Thus the continuous administration of primary factor, as Fraction E, together with Fraction A, but with minimal amounts of Fraction C, induced only slight reticulocyte and erythrocyte responses. A ten-fold increase in the amount of Fraction C (third period, Figure 5), however, was followed by a reticulocyte response of greater magnitude, and by a satisfactory gain of erythrocytes. That Fractions E, A, and C, each derived from 20 grams of liver, were therapeutically more effective than Fractions E — 20, A — 20, and C — 2, is suggested in view of the discussion of double reticulocyte responses by Minot and Castle (8).

The accessory action of Fraction F, in the

presence of the primary factor together with Fractions A and C, was indicated by the following observations. To Patient 3 (Curve 4, Figure 2) at an initial erythrocyte level of 1.3 million was administered a single dose of Fraction H — 370 grams. (It is to be recalled that Fraction H, in contrast with Fraction E, is completely devoid of Fractions A, C, and F.) During the following fifteen days the erythrocytes rose slightly but then receded to a final value of 1.28 million. During the succeeding twenty-eight days (Curve 3, Figure 2), to Fraction H — 100 grams were added Fractions A — 100 grams and C — 100 grams. These amounts were administered every tenth day. On the seventh day the reticulocytes rose to a peak of 24.4 per cent, followed by a rise of the erythrocytes to 1.93 million on the eighth day. Thereafter, the erythrocytes rose no further, and by the twenty-eighth day had declined to 1.78 million. At this point commercial liver extract was administered, followed by rapid rise of the erythrocytes. Thus, in the complete absence of Fraction F, Fractions H, A, and C induced a rise of erythrocytes of only 0.5 million during a period of twenty-eight days. On the other hand, after the addition of Fraction F, Fractions H, A, and C, administered in similar dosage, induced satisfactory erythrocyte responses in five other patients (Curves 10, 13, 15, 17, and 19, Figure 2).

The inactivity of the accessory factors in the absence of the primary factor

In Table III are presented data concerning the negative therapeutic activity of the three accessory factors, Fractions A, C, and F, in the *absence* of the primary factor. After the administration of the stated amounts of accessory factors in a single dose observations of the reticulocytes and of the erythrocytes were made during periods ranging from seven to twenty days. In subsequent periods either commercial liver extract, or partially purified experimental liver extract, was administered, with resulting satisfactory responses in each case. It is seen that neither Fractions A, C, or F individually or together (Patients 10, 20, and 21, Table III) induced significant rises of reticulocytes or erythrocyte responses, in patients who subsequently reacted to the administration of crude liver extracts.

Parenterally contrasted with orally administered tyrosine as an accessory factor

The experimental evidence discussed above, that as little as 6 mgm. of *l*-tyrosine (Fraction A — 100 grams liver), when administered by intramuscular injection, acted as an accessory factor, is difficult to reconcile with the apparent fact that the patients who formed the subjects of this investigation derived several *grams* of tyrosine daily from ingested protein. The patients consumed a normal amount of protein, severe diarrhea was absent, no gross evidence of defective protein digestion was manifest, and during the experimental periods most of the patients gained in weight.

In the hope of throwing some light on this problem the following study was undertaken. Patient 1, following the observations recorded in Figure 5, remained without treatment for one week, during which the erythrocyte level remained stable. The experiment presented in Figure 6 was then instituted. During the first period of twenty days Fractions E — 20 grams and C — 20 grams were administered on alternate days. During this period also 100 mgm. of commercial *l*-tyrosine was administered *orally* every day. No change in the patient's clinical condition took place, and during the twenty days the erythrocytes rose from 2.55 million to only 2.65 million. During the first ten days of the following period the administration on alternate days of the same basic fractions, E — 20 grams and C — 20 grams, was continued, but only 1.2 mgm. of tyrosine (A — 20 grams) was *injected* intramuscularly on alternate days. However, a sharp rise of the erythrocytes to 3.48 million in sixteen days ensued, accompanied by clinical improvement.

During the nineteen days following the last treatment the erythrocytes declined to 2.72 million. At this point the same basic fractions E — 20 grams and C — 20 grams were administered, and continued on alternate days for ten days. During this period 1.0 gram of tyrosine was ingested daily. An erythrocyte response promptly followed, reaching in sixteen days 3.48 million. It is evident that the slope of the erythrocyte curve following the daily oral administration of 1.0 gram of tyrosine is very similar to that following the parenteral administration on alternate days of 1.2 mgm. of tyrosine.

TABLE III
The inactivity of the accessory factors in the absence of the primary factor

Fraction.....	A		C			A + C		F	A + C + F		
	14	15	16	17	18	19	16		21	20	10
Patient.....											
FIRST PERIODS											
Responses to administration of accessory factors											
R.B.C. at beginning, millions per cu. mm.....	1.29	2.11	2.46	1.74	1.80	1.69	3.10	2.46	2.99	2.42	1.35
R.B.C. at end, millions per cu. mm.....	1.29	2.12	2.47	1.51	1.75	1.51	2.90	2.42	3.03	2.51	1.40
Maximum reticulocyte count, per cent.....	3.4	3.8	1.2	4.0	3.4	2.0	2.4	1.4	3.0	1.2	4.6
Length of period, days.....	13	8	9	9	18	7	19	12	10	12	20
Date of period.....	Mar. 29 to April 11, 1934	April 19 to April 27, 1934	Jan. 3 to Jan. 12, 1935	Feb. 12 to Feb. 21, 1935	May 1 to May 19, 1936	Jan. 26 to Feb. 2, 1935	Feb. 6 to Feb. 25, 1935	April 26 to May 8, 1935	Jan. 13 to Jan. 23, 1936	May 8 to May 20, 1935	April 28 to May 18, 1936
Amount fresh liver from which extract derived, grams.....	137	150	100	200	500	90	150	900	100	125	300
SECOND PERIODS											
Responses to administration of therapeutically active materials											
Material administered.....	C.l.e.*	E.l.e.†	E.l.e.	C.l.e.	E.l.e.	E.l.e.	E.l.e.	E.l.e.	E.l.e.	E.l.e.	E.l.e.
R.B.C. at beginning, millions per cu. mm.....	1.29	2.12	2.47	1.51	2.08	1.51	2.90	2.54	3.03	2.54	1.40
R.B.C. at end, millions per cu. mm.....	2.07	2.46	2.82	3.07	3.03	2.28	3.26	3.12	3.50	3.12	2.35
Reticulocyte peak, per cent.....	31.9	9.6	7.8	26.4		26.6	6.6	6.8	3.2	6.8	12.0
Length of period, days.....	12	9	9	8	10	9	9	10	10	10	10
Date of period.....	April 11 to April 23, 1934	April 27 to May 6, 1934	Jan. 12 to Jan. 21, 1935	Feb. 21 to Mar. 1, 1935	June 10 to June 20, 1936	Feb. 2 to Feb. 11, 1935	Feb. 25 to Mar. 6, 1935	June 10 to June 20, 1935	Jan. 23 to Feb. 2, 1936	June 10 to June 20, 1935	May 18 to May 28, 1936
Amount fresh liver from which extract derived, grams.....	100	77	88	2000	100	200	150	100	100	100	200

* Commercial liver extract.

† Experimental liver extract.

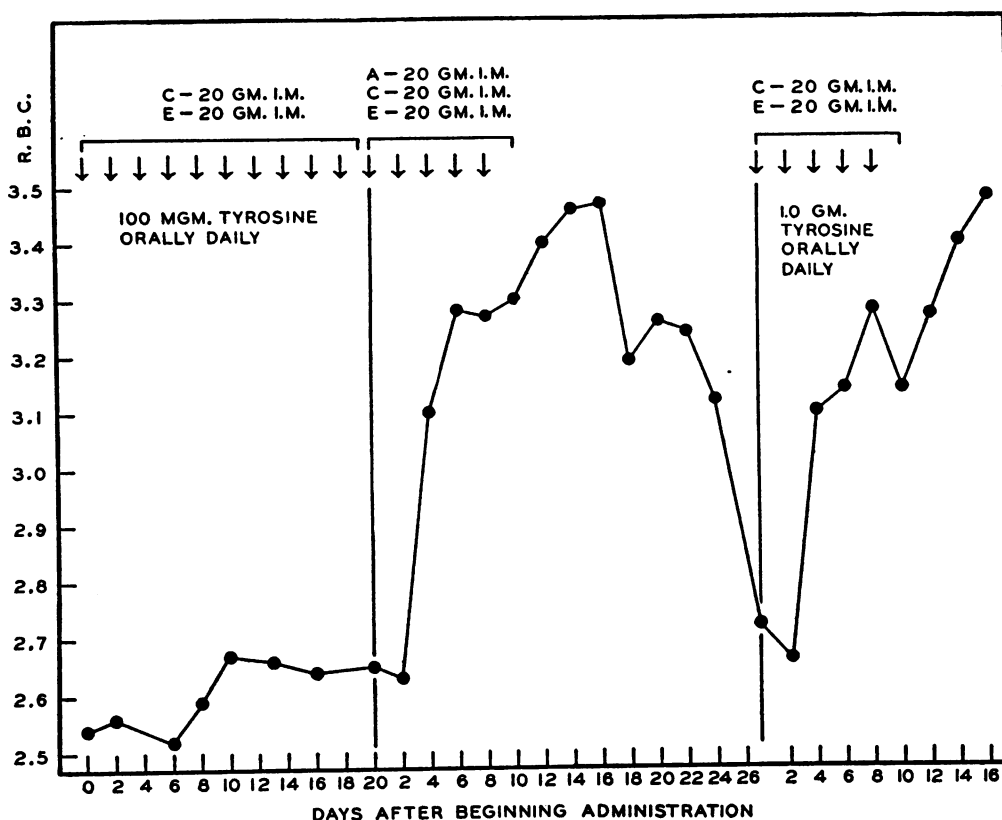


FIG. 6. PATIENT 1. PARENTERALLY CONTRASTED WITH ORALLY ADMINISTERED TYROSINE (FRACTION A) AS AN ACCESSORY HEMATOPOIETIC FACTOR

The quantities above each period refer to the amounts of fractions injected on the days denoted by each arrow. Fractions E and C were injected at a uniform rate in each of three periods. There was no erythrocyte response during the first period of twenty days while 100 mgm. of tyrosine was administered orally daily, followed by satisfactory rise of erythrocytes in the second period after 1.2 mgm. tyrosine was injected on alternate days. Following cessation of treatment, erythrocytes declined to previous level. In the third period, there was another satisfactory erythrocyte response after daily oral administration of 1.0 gram tyrosine.

It is thus suggested by these data that in this particular patient, at least, 100 mgm. of tyrosine ingested daily in addition to the tyrosine derived from the diet, was not equivalent in accessory hematopoietic activity to 0.6 mgm. (per day) of parenterally administered tyrosine, while a tenfold increase of the orally administered tyrosine was as active as the injected tyrosine.

DISCUSSION

The observations described above suggest that the complete therapeutic action of relatively crude liver extract in pernicious anemia was induced only by the presence in the extract of several chemically distinct substances. The evidence for

this hypothesis has been derived partly from data concerning reticulocyte production, but principally from data concerning erythrocyte regeneration during prolonged periods. One fraction has been termed a *primary* factor, the other fractions *accessory* factors. The primary factor alone, when administered in an amorphous impure state (Fractions E or H), exerted at most only a moderate therapeutic effect. The accessory factors (Fractions A, C, and F) were individually and collectively completely inert. On the other hand, the administration of the primary factor together with the three accessory factors was followed by satisfactory clinical improvement and by a rapid rate of erythrocyte regeneration.

It might be assumed that the satisfactory therapeutic activity of the four fractions together depended upon additive effects of a substance common to all. That this is unlikely is evidenced by the data of Table III. Fractions A, C, and F were individually and collectively administered in amounts sufficiently great, so that had there been contamination with the primary factor a response would have occurred. All of the evidence, rather, suggests that Fractions A, C, and F *augmented* the activity of a primary factor. Furthermore, it might be assumed that the entire therapeutic activity of liver extract resides in a single substance, and that the inadequate effects of our primary factor were due to the administration of insufficient amounts. Against this interpretation is the fact that even after the administration of an amount of primary factor derived from as much as 30 grams of liver per day (Curves 2 and 4, Figure 2), instead of the usual dosage of extract derived from 10 grams of liver per day, the resulting clinical and hematopoietic effects were inferior to those that followed the administration of smaller amounts (— 10 grams) of primary factor, *together with* like amounts of the three accessory factors.

We do not possess evidence that all possible accessory factors in crude liver extract are represented by Fractions A, C, and F. Nor do the present experimental data permit the conclusion that the four factors described are sufficient to induce a *complete* clinical and hematopoietic remission, and to maintain such a remission over a prolonged period. The longest periods over which continued observations have been made are the following. Patient 2, treated with Fractions H, A, C, and F, reached an erythrocyte level of 4.0 million in 27 days (Curve 19, Figure 2). Patient 12, whose course is depicted in Figure 2 (Curve 17), was treated with Fractions H, A, C, and F; at the end of 60 days the erythrocytes numbered 4.46 million.

Observations in one patient presented above have indicated that the accessory hematopoietic action of *l*-tyrosine (Fraction A), parenterally administered in an amount no greater than 0.6 mgm. per day, surpassed that effected by the tyrosine presumably derived from ingested protein, together with a daily oral ration of 100 mgm. of

l-tyrosine. These contrasting effective dosages might be due to either defective absorption of orally administered tyrosine, to abnormal destruction of tyrosine in the intestine, or to an abnormality of utilization of absorbed tyrosine. No data are at hand which bear on these possibilities.

The *minimal quantities* of all four factors that induce a satisfactory response have not been determined. With Fraction E as the primary factor the smallest total dosage administered was 26 mgm. of solids per 10 days (Figure 2, Curve 18). With Fraction H as the primary factor the smallest dosage administered was 51 mgm. of solids, per 10 days (Figure 2, Curve 15).

The above quantities are of interest in relation to the purified liver extracts recently described by other workers. Very similar amounts of total solids (per 10 days), with resulting satisfactory initial erythrocyte gains, were administered by Dakin, Ungley and West (9). On the other hand, Strandell, Poulsson and Schartum-Hansen (10) have recently reported hematopoietic responses following the use of purified amorphous material consisting of 0.35 mgm. of solids derived from 100 grams of liver, material that was prepared by Laland and Klem (11). Strandell and his collaborators administered this material to four patients, in the following dosages: Case I, total of 2.1 mgm. over a period of 42 days; Case II, 2.1 mgm. over a period of 32 days; Case III, 0.7 mgm. over a period of 9 days; and, Case IV, 0.7 mgm. over a period of 10 days. Although these authors conclude that 0.7 mgm. of their material "has a very good antianemic effect" their data, in our opinion, do not entirely substantiate this conclusion. Thus in Case I, although no initial erythrocyte count is given, on the seventh day the erythrocytes numbered 1.28 million, rising to 3.30 million at the end of 42 days, a distinctly submaximal response. In Case II the initial count is given as 0.91 million, rising to 2.1 million on the sixth day, but reaching only 2.45 million on the thirty-second day. In Case III the erythrocytes rose from 1.24 million to 1.4 million in 9 days. In Case IV the erythrocytes rose from 1.19 to 1.42 million in 10 days. These data, it seems to us, though indicating some therapeutic activity, are similar to the responses induced by our primary factor, in the absence of

the accessory factors (Figures 2 and 3). It has already been pointed out that the material of Laland and Klem shows certain chemical similarities to our Fraction I, derived from the primary factors, Fractions E or H (6).

SUMMARY AND CONCLUSIONS

Studies of the therapeutic activity of purified liver extract in pernicious anemia suggest that the hematopoietic effect may be exerted by an augmentative action of at least three chemically distinct *accessory* factors upon the activity of a *primary* factor. Of the three known accessory factors one is *L*-tyrosine, another contains a complex purine, and the third is a peptide. The accessory factors are completely devoid of the primary factor, and without the addition of the primary factor are therapeutically inert. The primary factor has been studied in an amorphous state. Its chemical nature is undetermined. Without the addition of the three accessory factors the primary factor is therapeutically only slightly active.

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